Variable skeletal phenotypes associated with biallelic variants in PRKG2

The 100 000 Genomes Project (100KGP) is a UK-wide initiative that has a goal of using whole genome sequencing (WGS) to identify genetic causes of rare inherited diseases and embed the use of this technology within the NHS. Using data from this resource alongside international gene-matching efforts, four individuals from two independent families were identified harbouring homozygous frameshift or stop-gain variants in PRKG2, a recently described skeletal dysplasia gene. Detailed clinical and radiological assessments helped extend the phenotypic range associated with this autosomal recessive condition while functional studies indicated that both variants had a similar impact on FGF-induced MAPK signalling.

PRKG2 encodes the cyclic guanosine monophosphate dependent protein kinase II (cGKiI), which acts downstream of the natriuretic peptide receptor-C/natriuretic peptide (NPR-B/CNP). NPR-B is encoded by NPR2, biallelic variants in which are responsible for acromesomelic dysplasia, Maroteaux type (AMDM; MIM 602875). Rodent models further implicated PRKG2 in skeletal development and cGKiI deficiency was shown to be the cause of the dwarfism phenotype observed in Angus cattle. Building on support from pathway analysis and model organisms, a recent study showed that biallelic PRKG2 variants can result in acromesomelic dysplasia, PRKG2-type (AMDP) in humans, adding PRKG2 to a list of >400 genes associated with genetic skeletal disorders. As only two affected individuals were reported, it is important that the full clinical range of this condition is described.

In this study, we searched for rare biallelic PRKG2 variants using data from the 100KGP via the LabKey application available within Genomic England’s research environment. Researchers can apply for access online (www.genomics.english. co.uk/join-a-gecip-domain). Initial filtering employed a 1% population allele frequency threshold based on data from the 1000 Genomes Project as well as in-house frequency information. An additional family was identified via a network of collaborators and variants were classified using ACMG criteria (online supplemental table 1).

In family 1, WGS and subsequent Sanger sequencing uncovered a homozygous pathogenic PRKG2 variant, NM_006259.3:c.2282dup (p.Asp761Glufs*34; online supplemental figure 1) in three brothers referred with spondyloepiphyseal dysplasia (figure 1A). Interestingly, the middle-affected brother (F1-IV-6) also has type 1 osteogenesis imperfecta (OI). An early clinical exome sequencing study found that for 4.6% of cases with a molecular diagnosis, more than one gene was contributing to a blended phenotype. Complex cases such as these are expected to be more common in highly consanguineous families where large regions of homozygosity (ROHs) make up a significant proportion of the genome; however, for F1-IV-6 the secondary diagnosis of OI was due to a COL1A1 frameshift, which had arisen de novo. OI was suspected in this child because of multiple fractures in childhood (arm as an infant, wrist aged 8 and thoracic T6 wedge fracture) combined with blue sclerae. It is certainly possible that the coexistent OI may have had an impact on the severity of the phenotype in this individual, not least because his height was more significantly reduced than for his two brothers and OI (type 1) is a known cause of reduced stature in its own right.

In family 2, exome sequencing for a girl with acromesomelic dysplasia revealed a homozygous pathogenic PRKG2 variant c.1705C>T; p.(Arg569*) (online supplemental figure 2), observed previously in a patient with similar clinical and radiological features. Comparison of the available genomic data for F2-V-3 and the previously published case was not able to detect a shared haplotype across the PRKG2 locus. However, exome sequencing has limited resolution to detect small regions of identity by descent and so a founder mutation cannot be ruled out. Given the differing ethnicities and the fact that c.1705C>T lies at a CpG dinucleotide, the recurrence of c.1705C>T being due to separate mutational events seems a more likely scenario.

Both variants described here are extremely rare; p.(Asp761Glufs*34) is absent from gnomAD (https://gnomad.broadinstitute.org), while p.(Arg569*) is present as a singleton allele. In both families, the disease-causing variants lay within large ROHs (online supplemental table 2). Pathogenic variants are overrepresented in the largest ROHs and it has been proposed that lying in one of the top 10 such regions can be used as evidence supporting pathogenicity. While the p.(Arg569*) variant has already been demonstrated to affect the downstream MAPK pathway, the p.(Asp761Glufs*34) in family 1 is likely to be disruptive given the switch of the final Asp-Phe residues for 33 alternative amino acids at the C terminus. In silico modelling highlights the structural importance of this region, in particular the final Phe762 residue (figure 1B, supplementary methods; interactive version at https://michelangelo.sgc.ox.ac.uk/j/prkg2).

To functionally confirm the pathogenicity of the newly identified p.(Asp761Glufs*34) variant, we first analysed cGKiI expression by western blot analysis. Plasmid construction for p.(Asp761Glufs*34) involved a sequential PCR strategy (supplemental methods), with the previously characterised p.(Arg569*) variant employed as a positive control. For both variants, cGKiI was detected at the predicted size (figure 1C), although at dramatically reduced levels (≥80%) compared with the wild type (figure 1D). Next, we evaluated whether the p.Asp761Glufs*34 mutant was able to inhibit FGF2-induced MAPK pathway by analysing its ability to induce phosphorylation of Raf-1 at Ser-338 and ERK1/2, as described previously. Wild-type cGKiI downregulated MAPK signalling by reducing ERK1/2 activation through the upstream phosphorylation of Raf-1 at Ser-338 and ERK1/2, as described previously. However, the p.Asp761Glufs*34 mutant failed to phosphorylate Raf-1 at Ser-338 and thus, reduced FGF2-induced ERK1/2 phosphorylation (figure 1E–F), similar to results for the p.Arg569* variant.

Detailed phenotypic information is provided for both families and compared with the two published cases (online supplemental table 2, figure 3). Radiological findings for F2-V-3 were very similar to those observed for ‘Probands 1’ described previously, which is unsurprising given that both individuals harbour the same homozygous p.(Arg569*). In contrast, for PRKG2 family 1 there was a consistent radiographic phenotype distinct from previously reported AMDP and AMDM. The three brothers reported here (F1-IV-3, IV-6 and IV-7) had no evidence of acromesomelic shortening, except for mild shortening of toes observed for individual F1-IV-7. The main findings were platyspondyly with anterior vertebral body projections, long slender femoral necks and some metaphyseal irregularity (most evident in the radius and ulna) and striations (figure 2). The metaphyses of the distal phalanges were somewhat cone-shaped in one child, but not...
Genotype-phenotype correlations

Figure 1  Pedigrees, structural modelling and the effects of the PRKG2 variants on cGKII protein levels/MAPK pathway regulation. (A) Simplified pedigrees and segregation of variants in PRKG2 and COL1A1 in two families with rare skeletal dysplasias. More detailed pedigrees are shown in online supplemental figure 4. AMD, acromesomelic dysplasia (mild); ND, not determined; OI, osteogenesis imperfecta; SMD, spondylometaphyseal dysplasia; WT, wild-type; *, WGS performed as part of 100KGP; +, exome sequencing. The COL1A1 variant was initially detected by targeted sequencing in 2011 but confirmed to have arisen de novo by WGS. (B) Structure of cGKII (wild type: turquoise) with overlay of the mutant, p.Asp761Glufs*34 (salmon) extension and inset of Phe762 residue. The protein kinase domain is regulated by two cyclic nucleotides binding (CNB) domains. The predicted C-terminal extension would fall between CNB-A domain and the protein kinase domain and is likely to interfere with the activation of the latter by the former, were it to be stable, a conclusion not supported by in silico predictions. In fact, the extension results in a deleterious amino acid change of a core residue, Phe762, to a leucine (inset). Also visible is the hydrogen bond between the terminal carboxylate and Thr519, whereas the amide bond between Leu762 and Leu763 is forced away in order to best accommodate the subsequent residues. (C) Immunoblotting results for cGKII (upper panel) and GAPDH as an endogenous control (lower panel) of cell lysates extracted from transiently transfected HEK293T cells. Both human cGKII mutants as well as wild-type (WT) proteins were detected at their predicted size: R569*: 65.1 kDa and D761Efs*34: 91.1 kDa (calculated by using the ExPASy online tool, https://web.expasy.org). (D) Densitometry quantification of cGKII showing that there is an 80% reduction in expression of the two mutants compared with WT. (E) Western blots of phosphorylated Raf-1 and ERK1/2 proteins of the MAPK pathway showed that neither of the mutants were able to phosphorylate c-Raf at Ser43 and therefore downregulate ERK activation compared with WT in response to FGF2 induction in transiently transfected HEK293 cells. (F) Densitometry quantification of pMAPK 44/42 protein revealed that neither R569* nor D761Efs*34 mutants were able to downregulate FGF2-induced ERK1/2 activation compared with WT, in transiently transfected HEK293 cells in the presence of 8-pCPT-cGMP. Three biological experiments were performed, and significance values are represented as *p<0.05, **p<0.01, ***p<0.001 and ****p>0.0001. EV, empty vector; T−, untransfected cells.

pronounced, generalised or associated with shortening, as seen in AMDM. In summary, family 1 exhibited a skeletal phenotype characterised by spondylometaphyseal dysplasia, rather than acromesomelic dysplasia as expected in AMDP and AMDM.

Interestingly, the PRKG2 locus has been identified in several genome-wide association studies on height (www.ebi.ac.uk/gwas/genes/PRKG2). Therefore, the description of this now confirmed Mendelian condition constitutes an additional example of rare variants in a gene causing a severe condition, where common variants in the same gene are associated with a related trait. In summary, analysis of 100KGP data combined with gene-matching efforts identified four affected individuals with biallelic loss of function variants in PRKG2, extending the
The patients described here were the only individuals harbouring severe biallelic PRKG2 variants across all rare disease areas within the 100KGP. These data include 295 patients recruited due to an unexplained skeletal dysplasia and therefore our results are consistent with this condition being extremely rare in humans.


Figure 2 Radiographic findings in two families with PRKG2 variants: radiographs of left upper limb (A) and lower limb (B) in a 26-month-old boy (F1-IV-7) from family 1. The long bones are stocky in appearance but there is no disproportion within the limbs. (C) Pelvic radiograph at age 4 in same child shows development of long, slender femoral necks. (D) Lateral spinal radiograph at age 4 show generalised mild platyspondyly with small central anterior projections of the vertebral bodies, and hypoplasia of the L2 vertebral body. (E) Left hand radiograph at age 11 in same child shows no brachydactyly; there is mild metaphyseal chondrodysplasia evident in the distal radius and particularly the ulna, with some metaphyseal striations (black arrow); subtle coning of the distal phalangeal metaphyses is evident (white arrows), without associated shortening. Pelvic (F) and lateral spine (G) radiographs in middle affected sibling (F1-IV-6) in family 1 showing similar features of long slender femoral necks and platyspondyly with anterior vertebral body projections. Osteopaenia is also evident; this child also has type 1 osteogenesis imperfecta due to a de novo pathogenic variant in COL1A1.

Additional radiology is available for F1-IV-3 in online supplemental figure 5 which shows similar results to those for F1-IV-7. (H) Left hand radiograph in female child (F2-V-3, aged 10 years) from family 2 showing generalised brachydactyly. (I) Right upper limb radiograph also from F2-V-3 demonstrates mild disproportionate shortening of the radius and ulna relative to the humerus (mesomelic shortening). (J) Pelvic radiograph from F2-V-3 demonstrates mildly elongated femoral necks. (K) Lateral spine radiograph from the same individual demonstrates mild platyspondyly with small anterior vertebral body projections.

Figure 2 Radiographic findings in two families with PRKG2 variants: radiographs of left upper limb (A) and lower limb (B) in a 26-month-old boy (F1-IV-7) from family 1. The long bones are stocky in appearance but there is no disproportion within the limbs. (C) Pelvic radiograph at age 4 in same child shows development of long, slender femoral necks. (D) Lateral spinal radiograph at age 4 show generalised mild platyspondyly with small central anterior projections of the vertebral bodies, and hypoplasia of the L2 vertebral body. (E) Left hand radiograph at age 11 in same child shows no brachydactyly; there is mild metaphyseal chondrodysplasia evident in the distal radius and particularly the ulna, with some metaphyseal striations (black arrow); subtle coning of the distal phalangeal metaphyses is evident (white arrows), without associated shortening. Pelvic (F) and lateral spine (G) radiographs in middle affected sibling (F1-IV-6) in family 1 showing similar features of long slender femoral necks and platyspondyly with anterior vertebral body projections. Osteopaenia is also evident; this child also has type 1 osteogenesis imperfecta due to a de novo pathogenic variant in COL1A1.

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phenotypic range of this condition to include spondylometaphyseal dysplasia. The patients described here were the only individuals harbouring severe biallelic PRKG2 variants across all rare disease areas within the 100KGP. These data include 295 patients recruited due to an unexplained skeletal dysplasia and therefore our results are consistent with this condition being extremely rare in humans.
Contributors

ATP and JCT conceived the project. ATP, BB-P, MPF, FD-G, TL, RM and KEH performed data analysis. BB-P and FD-G generated experimental data. The 57 members of GERC performed WGS and provided access data via a secure research environment. MBT, ADC, EGK, MD, AW, PW, KB, KE, RM, HT and DS recruited the patients and collected/interpreted clinical information. ATP drafted the manuscript, which was revised and approved by all.

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Competing interests

None declared.

Patient consent for publication

Consent obtained from parent(s)/guardian(s)

Ethics approval

The 100K Genomes Project has approval from the HRA Committee East of England (Cambridge South REC: 14/EE/1112).

Provenance and peer review

Not commissioned; externally peer reviewed.

Supplemental material

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### Table S2: Clinical details and variant information for four individuals from two independent families with biallelic variants in PRKG2.

| Family/individual | No. (Gender) | Parental relationship | Parents’ stature | Mother | Father | Ethnicity | Consanguinity (parental) | Allele Frequency | ROH region (chr4, size, Genomes Project, Truseq PCR-Free High Throughput library) | Allele Frequency | Homozygous variant | Allele Frequency | Homozygous variant | Allele Frequency | Homozygous variant | Allele Frequency | Homozygous variant | Allele Frequency | Homozygous variant | Allele Frequency | Homozygous variant | Allele Frequency | Homozygous variant | Allele Frequency | Homozygous variant |
|-------------------|-------------|-----------------------|------------------|--------|--------|-----------|--------------------------|----------------|--------------------------|----------------|-------------------|----------------|-------------------|----------------|-------------------|----------------|-------------------|----------------|-------------------|----------------|-------------------|----------------|-------------------|----------------|-------------------|----------------|-------------------|
| Family 1          | IV-3(M/F)   | Mother 160 cm (-0.60 SD); Father 177 cm (0 SD); | Normal, except for broad thumbs | Normal | Normal | Yes (double 2nd cousins) | Yes (3rd cousins) | .42 (PMID: 33559681) | chr4:25,732,624-85,677,887 | .35 (PMID: 33559681) | p.(Glu591Serfs*175) | .19 (PMID: 33559681) | p.(Lys450*175) | .15 (PMID: 33559681) | p.(Glu591Serfs*175) | .19 (PMID: 33559681) | p.(Lys450*175) | .15 (PMID: 33559681) | p.(Glu591Serfs*175) | .19 (PMID: 33559681) | p.(Lys450*175) | .15 (PMID: 33559681) |
| Family 2          | IV-6(M/F)   | Mother 147 cm ( -2.50 SD); Father 155 cm (-2.10 SD); | Normal | Normal | Normal | Yes (double 1st cousins) | Yes (3rd cousins) | .42 (PMID: 33559681) | chr4:53,687,014-85,677,887 | .35 (PMID: 33559681) | p.(Glu591Serfs*175) | .19 (PMID: 33559681) | p.(Lys450*175) | .15 (PMID: 33559681) | p.(Glu591Serfs*175) | .19 (PMID: 33559681) | p.(Lys450*175) | .15 (PMID: 33559681) | p.(Glu591Serfs*175) | .19 (PMID: 33559681) | p.(Lys450*175) | .15 (PMID: 33559681) |
| Family 3          | IV-7(M/F)   | Mother 171 cm (1.90 SD); Father 158.5 cm (-2.5 SD); | Normal | Normal | Normal | Yes (double 2nd cousins) | Yes (3rd cousins) | .42 (PMID: 33559681) | chr4:25,732,624-85,677,887 | .35 (PMID: 33559681) | p.(Glu591Serfs*175) | .19 (PMID: 33559681) | p.(Lys450*175) | .15 (PMID: 33559681) | p.(Glu591Serfs*175) | .19 (PMID: 33559681) | p.(Lys450*175) | .15 (PMID: 33559681) | p.(Glu591Serfs*175) | .19 (PMID: 33559681) | p.(Lys450*175) | .15 (PMID: 33559681) |
| Family 4          | V-3(M/F)    | Mother 140.8 cm (-4.9 SD); Father 143.8 cm (-3.11 SD); | Normal, except for broad thumbs | Normal | Normal | Yes (3rd cousins) | Yes (3rd cousins) | .42 (PMID: 33559681) | chr4:25,732,624-85,677,887 | .35 (PMID: 33559681) | p.(Glu591Serfs*175) | .19 (PMID: 33559681) | p.(Lys450*175) | .15 (PMID: 33559681) | p.(Glu591Serfs*175) | .19 (PMID: 33559681) | p.(Lys450*175) | .15 (PMID: 33559681) | p.(Glu591Serfs*175) | .19 (PMID: 33559681) | p.(Lys450*175) | .15 (PMID: 33559681) |

#### Notes
- Variants were classified using the Mainz Criteria for genotypes and a recently updated criteria for genotype classifications.
- The quality of evidence was assessed using the GRADE system and the level of evidence is shown in brackets after each genotype.
- The table includes a summary of the clinical features, variant information, and sequencing methods for the four individuals from two independent families with biallelic variants in PRKG2.
SUPPLEMENTARY INFORMATION

Variable skeletal phenotypes associated with biallelic variants in PRKG2

Figure S1: Sanger sequencing electropherogram validating the PRKG2 variant in Family 1 NM_006259.3: c.2282dupA; p.(Asp761Glufs*34) which lies close to the wild-type C-terminus. Heterozygosity in parental samples was already confirmed by WGS.

Figure S2: Sanger sequencing electropherogram showing the PRKG2 variant in Family 2 NM_006259.3: c.1705C>T; p.(Arg569*) to be homozygous in the proband and heterozygous in a parental sample.
Recessive skeletal dysplasia associated with PRKG2

Figure S3: Photos showing clinical features of note. A) Photo showing of the toes in Family 1 which were unremarkable for F1-IV-3 (top) and F1-IV-6 (middle), but with mild shortening seen for F1-IV-7 (bottom). B) Photo showing short/broad fingers in individual F2-V-3 (aged 10 years). C) Photo showing subtle dysmorphic features in F2-V-3, with thick eyebrows, synophrys and a broad nasal bridge.
Recessive skeletal dysplasia associated with \textit{PRKG2}

\textbf{Figure S4:} Full pedigrees and segregation of variants in \textit{PRKG2} and \textit{COL1A1} for 2 families with rare skeletal dysplasias. Pedigrees are shown in same order as the simplified versions in Figure 1A. SMD, Spondylometaphyseal dysplasia; OI, Osteogenesis imperfecta; AD, Acromesomelic dysplasia (mild); WT, wild-type; ND = not determined; *, genome sequencing performed as part of 100KGP, +, exome sequencing.
Recessive skeletal dysplasia associated with PRKG2

Figure S5: Additional radiological images for individual F1-IV-3. A) Chest radiograph aged 13; normal appearances. Lateral lumbar spine (B) radiograph and lateral thoracic spine radiograph aged 13. There is mild generalised platyspondyly with more pronounced height reduction in the dorsal aspects of the vertebral bodies. D) Hand and forearm aged 13. E) Hand and wrist aged 18; there is no brachydactyly.

Supplementary Methods

In silico modelling of PRKG2 frameshift variant

This composite model of the cGMP-dependent protein kinase 2 was obtained by combining several parts. The kinase domain was predicted by I-Tasser\(^1\), while the cGMP binding domains were the crystal structures PDB:5C6C and PDB:5BV6.\(^2\) The in silico calculations were done with Pyrosetta.\(^3\) Further details and the full code is available at https://github.com/matteoferla/PRKG2_analysis. The interactive version of the model is provided using the MichelaNGLo tool.\(^4\)

Plasmid construction
Recessive skeletal dysplasia associated with PRKG2

The empty Myc-DDK-tagged expression pCMV6-Entry plasmid as well as the human PRKG2 (NM_006259.2; pCMV6-PRKG2-WT) wild type plasmids were obtained from Origene Technologies (Rockville, MD). The PRKG2 p.R569* mutant was generated as previously reported. The p.D761Efs*34 mutant was generated by two sequential PCRs in order to add the extra nucleotide sequence to the C-terminus of the wild type sequence. The PCR-1 primers were 5’ CCGGCAGATCGCCTGGGAATGTTCACTGAAACCACTATCTAAGCACCAGATGGACAC3’ (forward) and 5’ CTTCTGTAGAGTACAGGCAGTAATCAACTTTTCTTCTGTCAAGGTCTTTATCCCAGCCTGATAGC 3’ (reverse). The product of PCR-1 was then submitted to a second PCR (PCR-2) to add the remaining nucleotides to complete the mutated sequence. The same forward primer was used in PCR-2 whilst 5’ GTACGGTTAATACCTGAAAAGAAAATAATGTGGATTATTTATCGCCTGCCCTCTCTGTAGATACA 3´ was employed as the reverse primer. The PCR product was then cloned into a pCR2.1 vector by TA cloning method following manufacturer’s instructions (Life Technologies, Carlsbad, CA). Next, the cloned fragment was subcloned into pCMV6 vector using the pCMV6-PRKG2-WT construct digested with SgfI and MluI enzymes (Thermo Fisher Scientific) resulting in the pCMV6-PRKG2-D761Efs*34 mutant plasmid. The construct was verified by Sanger sequencing.

**MAPK pathway analysis**

Cell culture, transient transfections, Western blots and densitometry quantification were performed as previously described.

**Table S1: Variant interpretation using ACMG guidelines.**

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<th>Family ID</th>
<th>Family 1</th>
<th>Family 2</th>
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<td>Variant</td>
<td>NM_006259.3:c.2282dup p.(Asp761Glufs*34)</td>
<td>NM_006259.3:c.1705C&gt;T p.(Arg569*)</td>
</tr>
<tr>
<td>ACMG evidence codes</td>
<td>PVS1 (applied with caution as near 3’ end but N.B. 33 additional residues and WB data in Fig 1C,D supports use), PS3 (MAPK signalling affected, Fig 1E,F), PM2 (absent in gnomAD), PP1 (cosegregation, Fig 1A), PP3 (structural modelling, Fig 1B)</td>
<td>PVS1, PS3 (Díaz-González et al and now replicated), PM2 (singleton allele in gnomAD 2.1.1), PP3 (in silico e.g. CADD=37)</td>
</tr>
</tbody>
</table>

| ACMG classification | Pathogenic | Pathogenic |

**Table S2:** Clinical details and variant information for four individuals from two independent families with biallelic variants in PRKG2. Results are compared to two cases reported by Díaz-González et al 2020. Sequencing methods in Family 2 are as described by Pagnamenta et al 2021. NA, not available; ROH, region of homozygosity. Variant annotation is based on NM_006259.3. SDs for adults heights were calculated using https://tall.life/height-percentile-calculator-age-country. For Family 1 we used the mean adult height obtained from the Royal College of Paediatrics and Child Health’s standard growth chart

*N.B. This table is provided as a separate xlsx file.*
Recessive skeletal dysplasia associated with PRKG2

References