

Maternal versus paternal inheritance of a 132 bp 11p15.5 microdeletion affecting *KCNQ1OT1* and associated phenotypes

In this brief communication, we address the possible associations of a 132 bp deletion within the antisense gene *KCNQ1OT1* on 11p15.5 to growth abnormalities. We question its recently suggested connection of paternal inheritance with Silver-Russell syndrome (SRS; OMIM #180860) and demonstrate maternal inheritance of the same variant in a 16-year-old girl with a tumour of the left leg. She also had mild isolated hemihypertrophy, suggesting a possible link to Beckwith-Wiedemann syndrome (BWS; OMIM #130650). Our findings underscore the complexity of 11p15.5 structural variations and the diffidence required when assessing their pathogenicity.

Genetic alterations of 11p15.5 imprinted gene clusters (regulated by imprinting centres H19/IGF2:IG-DMR (IC1) and *KCNQ1OT1*:TSS-DMR (IC2)) contribute to the phenotypically inverse growth disorders Beckwith-Wiedemann syndrome (BWS; OMIM #130650) and Silver-Russell syndrome (SRS; OMIM #180860).^{1,2}

BWS shows locus heterogeneity within the 11p15.5 chromosomal region, where IC2 loss of methylation, IC1 gain of methylation, 11p15.5 UPD/CNV or *CDKN1C* pathogenic variants may lead to a wide spectrum of BWS features ranging from isolated cancer predisposition³ to severe dysmorphism.² The predominant cause of SRS is IC1 loss of methylation, with UPD7, 11p15.5 CNVs or *CDKN1C* variants constituting rarer causes.^{4,5}

As recently summarised by Eggermann *et al*,⁵ eight individuals with microdeletions in IC2 have been observed in five families. The deletions, all spanning 60 kb or more, affected the *KCNQ1OT1* gene, the IC2, and part of the exonic sequence of *KCNQ1*. Paternal loss of

KCNQ1OT1 should lead to expression of paternal *CDKN1C*, theoretically impeding growth. Among the eight individuals, five had paternally inherited deletions (three showing SRS phenotype and two asymptomatic) and three had maternally inherited deletions (all showing BWS phenotype).

In a sixth family, they presented a 132 bp deletion located in the non-coding gene *KCNQ1OT1*, which overlapped with intron 11 of *KCNQ1* (*KCNQ1OT1*: NR_002728.3:n.134_265del; *KCNQ1*: LRG287t1:c.1514+37 654_1514+37 785 del; Chr11(GRCh37):g.2,720,964–2721,095del). The proband, a 4-year-old girl, inherited this alteration paternally and scored 2 of 6 on the Netchine-Harbison scale (postnatal biometrics and prominent forehead). Thus, the affected girl did not meet the clinical criteria for the diagnosis of SRS. The authors postulate that her short stature is the result of paternal expression of *CDKN1C*. They also suggest that the father was unaffected as his deletion was inherited maternally and *KCNQ1OT1* is therefore already silenced.

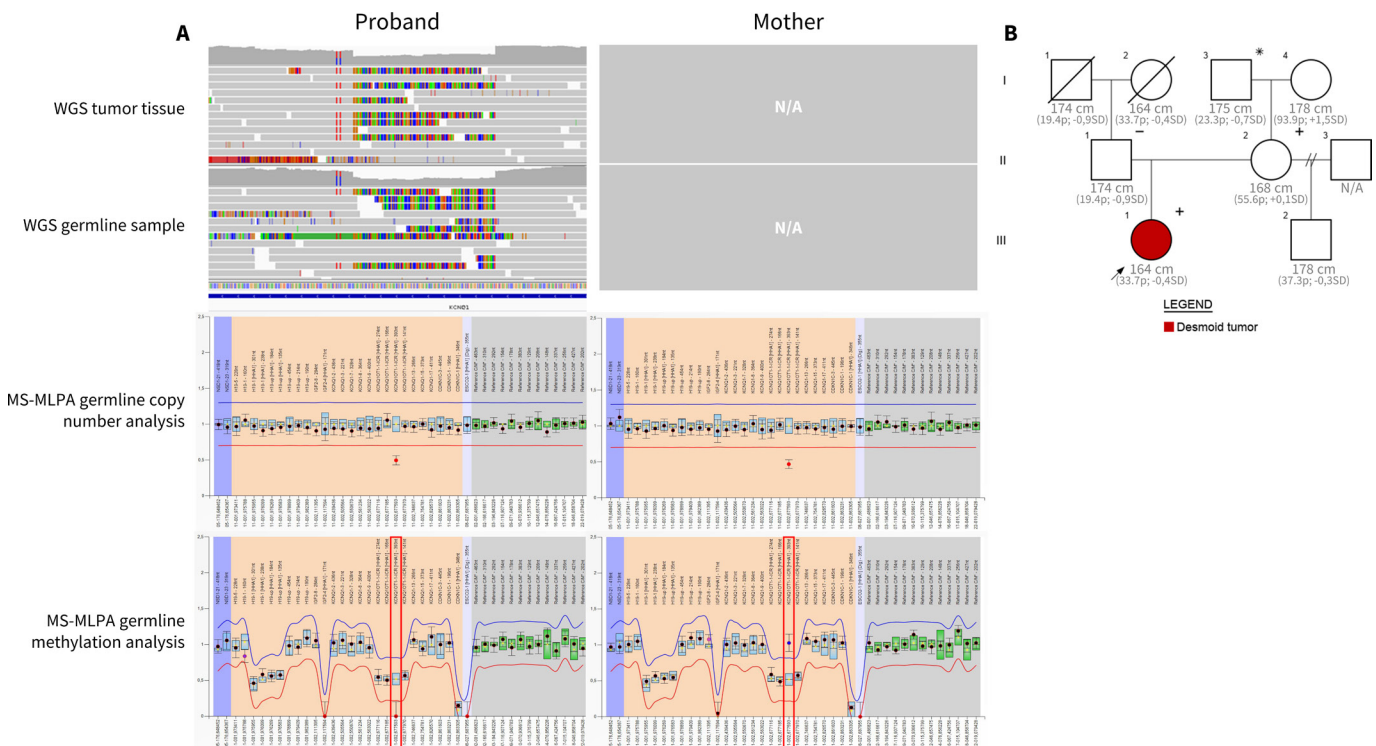


Figure 1 (A) Evidence of the 132 bp deletion in the proband (on the left) and her mother (on the right). In the proband, the deletion was evident in both germline and tumour whole genome sequencing (WGS) data, showing a distinct coverage drop and split reads. As expected, methylation-specific multiplex ligation-dependent probe amplification analysis confirmed the deletion with a single probe located in this genomic region (*KCNQ1OT1*-393bp, 07 172 L06781). Heterozygous deletion of the same probe was identified in the mother. In the proband, the observed hypomethylation of the probe (highlighted by red rectangle) indicates a deletion on the maternal allele (confirmed); while, in the mother, hypermethylation (highlighted by red rectangle) of the probe indicates a deletion on the paternal allele (surmised). (B) Pedigree with the arrow indicating the proband. Carriers of the 132 bp deletion were either confirmed (+) or surmised (*). Proband's father tested negative (–). Adult height with percentile and SD for the gender are presented below each subject.

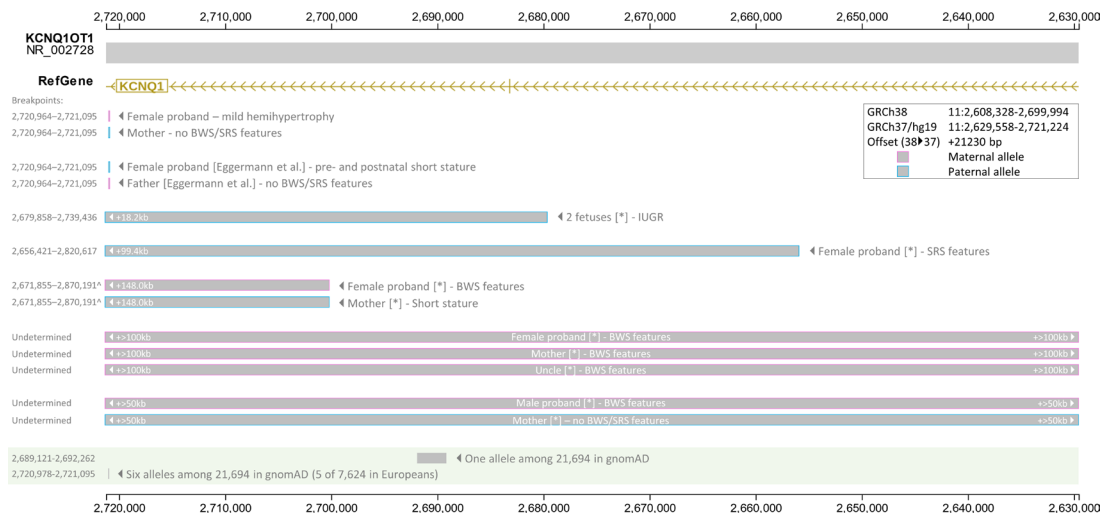


Figure 2 Diagram illustrating reported heterozygous deletions within or across the *KCNQ1OT1* gene region on chromosome 11 (hg19 location on the axes at the top and bottom; GRCh38 liftover offset is indicated in the box on the right along with gene coordinates). The deletion reported in our study and in the study by Eggermann *et al* are presented at the top with breakpoints, location indicated by grey bar, individual with the genotype and their phenotype shown going from left to right. Below, overlapping deletions reported in the literature are listed (*based on overview by Eggermann *et al*).⁵ Numbers at the tail ends of the deletions indicate breakpoint up or down stream from the shown region. Maternal and paternal inheritance is indicated by pink and blue outlines, respectively. All heterozygous deletions within the *KCNQ1OT1* gene reported in gnomAD (SV V.2.1) are listed along with number of carriers in the light green box. There were no homozygotes.

We report a seventh family harbouring precisely the same 132bp deletion described in the family presented by Eggermann *et al*. The proband was identified as part of the STAGING project (Sequencing Tumor And Germline DNA – Implications and National Guidelines), a national cohort of Danish paediatric cancer patients, offering extensive prospective sequencing of germline DNA. The first 198 patients have been published, including the sequencing and bioinformatics pipeline methods.⁶

The 15 11/12 years old female proband was included in STAGING⁶ following a diagnosis of an unspecified tumour located in the right popliteal region. Whole genome sequencing (WGS) of a needle biopsy from the tumour revealed a somatic *CTNNB1* (NM_001904.4: c.121A>G, p.(Thr41Ala)) gain-of-function variant strongly associated with desmoid fibromatosis tumours cooperated by histopathology. The fibromatosis mass was deemed unsuited for radical surgery due to involvement of the tibial nerve and major vessels. Clinical and radiological progression of the lesion during a wait-and-see approach prompted initiation of chemotherapy with methotrexate and vinblastine followed by tyrosine kinase inhibitors, which both proved largely ineffective (figure 1). Alternative limb-sparing surgical therapies are now being explored.

The proband had an uncomplicated birth at 37 weeks of gestation with a birth weight of 3200g (−0.37 z). She

never experienced feeding difficulties. She presented with right-sided toe walking at 15 months of age and was diagnosed with a likely causative congenital shortening of the right Achilles tendon, surgically corrected at age 4. She had menarché around age 10 years and stopped growing at age 13 9/12 years, with a final height of 164 cm (33.7 percentile, −0.4SD). She had both foot (left foot two European sizes smaller) and leg length (left leg 0.5 cm longer) discrepancy, although the latter falls within normal range. She presented no other features of BWS.

Methylation-specific multiplex ligation-dependent probe amplification analysis (MS-MLPA, ME030-C3, MRC Holland, Amsterdam, The Netherlands) was carried out to verify the 132bp germline deletion, and a heterozygous deletion of a single probe located in the same region (*KCNQ1OT1*-393bp, 07172-L06781) was detected in the proband and her mother. MS-MLPA of the proband's mother indicated paternal inheritance of the deletion (hypermethylation). The breakpoints of the 132bp deletion were obtained from WGS of the germline and tumour DNA and were identical to the deletion of the previously family reported by Eggermann *et al* (figure 2).

According to the mechanism proposed by Eggermann *et al*, the mother of the proband of our study and the proband described in their study should both display an Silver-Russell syndrome phenotype, caused by the paternally inherited


132bp deletion. However, at 46 years of age, the mother of the proband of our study reports no significant health issues and she is of normal height (168 cm; 55.6 percentile, 0.1SD), fulfilling none of the Netchine-Harbisson criteria. This suggests that the deletion presents with incomplete penetrance, is benign or mosaic.

The proband of our study displays a phenotype that overlaps somewhat with BWS; mild isolated hemihypertrophy and a shortened Achilles tendon (previously reported in BWS⁷). Finally, *CTNNB1* gain-of-function variants are known to drive a significant proportion of paediatric BWS cancers.⁸ Among 5213 childhood cancers reported in the St. Jude Cloud,⁹ 63 had somatic *CTNNB1* variants; medulloblastoma (39/722), Wilms' (10/252), adrenal cortical carcinoma (4/41), hepatoblastoma (3/16) and rhabdomyosarcoma (2/60). While medulloblastomas are not linked to BWS, the four other types constitute the hallmark cancers associated with the syndrome.

Importantly, a 117bp deletion (Chr 11 (GRCh37): g.2,720,978–2721,095del) fully overlapping with the 132bp deletion reported here and by Eggermann *et al* was detected in 6 out of 10847 individuals in gnomAD, with 5 out of 3812 Europeans being carriers.¹⁰

We have elaborated on the phenotypes associated with a rare germline 132bp deletion within *KCNQ1OT1*, suggesting a, perhaps incidental, link to tumour predisposition. Overall, our findings have

direct implications for the genetic counselling of families with this variant. Existing empirical evidence is inconsistent, and we consider the variant to be of unknown significance. Further investigations of small deletions within this region are required to clarify any link to BWS/SRS-related disorders.

Ulrik Kristoffer Stoltze ^{1,2}
Thomas Van Overeem Hansen,¹
Jesper Sune Brok,² Karen Grønsvov,¹
Zeynep Tumer,^{1,3} Lise Barlebo Ahlborn,⁴
Kjeld Schmiegelow,² Karin A W Wadt^{1,3}

¹Department of Clinical Genetics, Rigshospitalet, Copenhagen, Denmark

²Department of Pediatrics and Adolescent Medicine, Rigshospitalet, Copenhagen, Denmark

³Department of Clinical Medicine, Rigshospitalet, Copenhagen, Denmark

⁴Center for Genomic Medicine, Rigshospitalet, Copenhagen, Denmark

Correspondence to Dr Ulrik Kristoffer Stoltze; ulrik.kristoffer.stoltze@regionh.dk

Correction notice This article has been corrected. The name of author Zeynep Tumer has been amended.

Acknowledgements Research project nurses, Anja Hirche and Solvej Kullegaard, collected consent, samples and data from patient and family.

Contributors UKS, TVOH, KG, AZT, KS and KAWW contributed to the overall planning, conduct and reporting of the work described in the article. UKS conceptualised the study, did the primary data analysis and wrote the manuscript. KG and AZT performed MLPA-MS analyses and contributed to the conduct and reporting of the work. JSB was the lead clinical investigator and contributed to the conduct and reporting of the work. LBA ran genomic and bioinformatic analysis on tumour samples and contributed to the conduct and reporting of the work. UKS and KAWW are guarantors and accept full responsibility for the finished work and/or the conduct of the study, had access to the data and controlled the decision to publish. All authors contributed to the review and final approval of the manuscript.

Funding Funding was received from BørneCancerFonden, the European Regional Development fund (Interregional Childhood Oncology Precision Medicine Exploration (iCOPE)). This work is part of Childhood Oncology Network Targeting Research, Organisation & Life expectancy (CONTROL) and supported by Danish Cancer Society (R-257-A14720) and the Danish Childhood Cancer Foundation (2019-5934).

Competing interests Authors had no competing interests related to this work. In the past 36 months, the following authors received speaker fees: TVOH (Pfizer), KS (Jazz Pharmaceuticals, Servier, Amgen, and Medscape) and KAWW (Pfizer).

Patient consent for publication Consent obtained directly from patient(s).

Ethics approval Ethical approval was obtained through the regional scientific ethical committee (the Ethical Scientific Committees for the Capital Region, H-15016782) and the Danish Data Protection Agency

(RH-2016-219, I-Suite no: 04804). Participants gave informed consent to participate in the study before taking part.

Provenance and peer review Not commissioned; externally peer reviewed.



OPEN ACCESS

Open access This is an open access article distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited, appropriate credit is given, any changes made indicated, and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>.

© Author(s) (or their employer(s)) 2023. Re-use permitted under CC BY-NC. No commercial re-use. See rights and permissions. Published by BMJ.



Check for updates

To cite Stoltze UK, Hansen TVO, Brok JS, et al. *J Med Genet* 2023;**60**:128–130.

Received 11 November 2021

Accepted 1 March 2022

Published Online First 30 June 2022

J Med Genet 2023;**60**:128–130.

doi:10.1136/jmedgenet-2021-108335

ORCID iD

Ulrik Kristoffer Stoltze <http://orcid.org/0000-0001-5862-3292>

REFERENCES

- Wakeling EL, Brioude F, Lokulo-Sodipe O, O'Connell SM, Salem J, Bliet J, Canton APM, Chrzanoska KH, Davies JH, Dias RP, Dubern B, Elbracht M, Giabicani E, Grimberg A, Grønsvov K, Hokken-Koelega ACS, Jorge AA, Kagami M, Linglart A, Maghnie M, Mohnke K, Monk D, Moore GE, Murray PG, Ogata T, Petit IO, Russo S, Said E, Toumba M, Tümer Z, Binder G, Eggermann T, Harbison MD, Temple IK, Mackay DJG, Netchine I. Diagnosis and management of Silver-Russell syndrome: first international consensus statement. *Nat Rev Endocrinol* 2017;**13**:105–24.
- Brioude F, Kalish JM, Mussa A, Foster AC, Bliet J, Ferrero GB, Boonen SE, Cole T, Baker R, Bertolotti M, Cocchi G, Coze C, De Pellegrin M, Hussain K, Ibrahim A, Kilby MD, Krajewska-Walasek M, Kratz CP, Ladusans EJ, Lapunzina P, Le Bouc Y, Maas SM, Macdonald F, Öunap K, Peruzzi L, Rossignol S, Russo S, Shipster C, Skórka A, Tatton-Brown K, Tenorio J, Tortora C, Grønsvov K, Netchine I, Hennekam RC, Prawitt D, Tümer Z, Eggermann T, Mackay DJG, Riccio A, Maher ER. Expert consensus document: clinical and molecular diagnosis, screening and management of Beckwith-Wiedemann syndrome: an international consensus statement. *Nat Rev Endocrinol* 2018;**14**:229–49.
- Fiala EM, Ortiz MV, Kennedy JA, Glodzik D, Fleischut MH, Duffy KA, Hathaway ER, Heaton T, Gerstle JT, Steiner P, Shukla N, McNeer N, Tkachuk K, Bouvier N, Cadoo K, Carlo MI, Latham A, Dubard Gault M, Joseph V, Kemel Y, Kentsis A, Stadler Z, La Quaglia M, Papaemmanuil E, Friedman D, Ganguly A, Kung A, Offit K, Kalish JM, Walsh MF. 11p15.5 epimutations in children with Wilms tumor and hepatoblastoma detected in peripheral blood. *Cancer* 2020;**126**:3114–21.
- Brioude F, Oliver-Petit I, Blaise A, Praz F, Rossignol S, Le Jule M, Thibaud N, Fausat A-M, Tauber M, Le Bouc Y, Netchine I, Jule ML, Bouc YL. *CDKN1C* mutation affecting the PCNA-binding domain as a cause of familial Russell Silver syndrome. *J Med Genet* 2013;**50**:823–30.
- Eggermann T, Kraft F, Lausberg E, Ergezinger K, Kunstmann E. Paternal 132 bp deletion affecting *KCNQ1OT1* in 11p15.5 is associated with growth retardation but does not affect imprinting. *J Med Genet* 2021;**58**:173–6.
- Byrjalsen A, Hansen TVO, Stoltze UK, Mehrjouy MM, Barnkob NM, Hjalgrim LL, Mathiasen R, Lautrup CK, Gregersen PA, Hasle H, Wehner PS, Tuckuviene R, Sackett PW, Laspiur AO, Rossing M, Marvig RL, Tommerup N, Olsen TE, Scheie D, Gupta R, Gerdes A-M, Schmiegelow K, Wadt K. Nationwide germline whole genome sequencing of 198 consecutive pediatric cancer patients reveals a high incidence of cancer prone syndromes. *PLoS Genet* 2020;**16**:e1009231.
- Poole RL, Leith DJ, Docherty LE, Shmela ME, Gicquel C, Splitt M, Temple IK, Mackay DJG. Beckwith-Wiedemann syndrome caused by maternally inherited mutation of an OCT-binding motif in the IGF2/H19-imprinting control region, ICR1. *Eur J Hum Genet* 2012;**20**:240–3.
- Fukuzawa R, Hata J-ichi, Hayashi Y, Ikeda H, Reeve AE. Beckwith-Wiedemann syndrome-associated hepatoblastoma: Wnt signal activation occurs later in tumorigenesis in patients with 11p15.5 uniparental disomy. *Pediatr Dev Pathol* 2003;**6**:299–306.
- McLeod C, Gout AM, Zhou X, Thrasher A, Rahbarinia D, Brady SW, Macias M, Birch K, Finkelstein D, Sunny J, Mudunuri R, Orr BA, Treadway M, Davidson B, Ard TK, Chiao A, Swistak A, Wiggins S, Foy S, Wang J, Sioson E, Wang S, Michael JR, Liu Y, Ma X, Patel A, Edmonson MN, Wilkinson MR, Frantz AM, Chang T-C, Tian L, Lei S, Islam SMA, Meyer C, Thangaraj N, Tater P, Kandali V, Ma S, Nguyen T, Serang O, McGuire I, Robison N, Gentry D, Tang X, Palmer LE, Wu G, Suh E, Tanner L, McMurry J, Lear M, Pappo AS, Wang Z, Wilson CL, Cheng Y, Meshinchi S, Alexandrov LB, Weiss MJ, Armstrong GT, Robison LL, Yasui Y, Nichols KE, Ellison DW, Bangur C, Mullighan CG, Baker SJ, Dyer MA, Miller G, Newman S, Rusch M, Daly R, Perry K, Downing JR, Zhang J. St. Jude cloud: a pediatric cancer genomic Data-Sharing ecosystem. *Cancer Discov* 2021;**11**:1082–99.
- Collins RL, Brand H, Karczewski KJ, Zhao X, Alfoldi J, Francioli LC, Khera AV, Lowther C, Gauthier LD, Wang H, Watts NA, Solomonson M, O'Donnell-Luria A, Baumann A, Munshi R, Walker M, Whelan CW, Huang Y, Brookings T, Sharpe T, Stone MR, Valkanas E, Fu J, Tiao G, Laricchia KM, Ruano-Rubio V, Stevens C, Gupta N, Cusick C, Margolin L, Taylor KD, Lin HJ, Rich SS, Post WS, Chen Y-DI, Rotter JJ, Nusbaum C, Philippakis A, Lander E, Gabriel S, Neale BM, Kathiresan S, Daly MJ, Banks E, MacArthur DG, Talkowski ME, Genome Aggregation Database Production Team, Genome Aggregation Database Consortium. A structural variation reference for medical and population genetics. *Nature* 2020;**581**:444–51.