SHORT REPORT

Support of the histaminergic hypothesis in Tourette Syndrome: association of the histamine decarboxylase gene in a large sample of families

Iordanis Karagiannidis,1 Sandra Dehning,2 Paul Sandor,3 Zsanett Tarnok,4 Renata Rizzo,5 Tomasz Wolanczyk,6 Marcos Madruga-Garrido,7 Johannes Hebebrand,8 Markus M Nöthen,9 Gerd Lehmkuhl,10 Luca Farkas,4 Peter Nagy,4 Urszula Szymanska,6 Zachos Anastasiou,1 Vasileios Statthias,1 Christos Androutsos,11 Vaia Tsironi,11 Anastasia Koumoula,11 Csaba Barta,12 Peter Zill,2 Pablo Mir,7,13 Norbert Müller,2 Cathy Barr,3 Peristera Paschou1

ABSTRACT

Background Gilles de la Tourette Syndrome is a neurodevelopmental disorder that is caused by the interaction of environment with a complex genetic background. The genetic etiology of the disorder remains, so far, elusive, although multiple promising leads have been recently reported. The recent implication of the histamine decarboxylase (HDC) gene, the key enzyme in histamine production, raises the intriguing hypothesis of a possible role of histaminergic dysfunction leading to TS onset.

Methods Following up on the finding of a nonsense mutation in a single family with TS, we investigated variation across the HDC gene for association with TS. As a result of a collaborative international effort, we studied a large sample of 520 nuclear families originating from seven European populations (Greek, Hungarian, Italian, Polish, German, Albanian, Spanish) as well as a sample collected in Canada.

Results and Conclusions Interrogating 12 tagging SNPs (ISNP) across the HDC region, we find strong over-transmission of alleles at two SNPs (rs854150 and rs1894236) in the complete sample, as well as a statistically significant associated haplotypes. Analysis of individual populations also reveals signals of association in the Canadian, German and Italian samples. Our results provide strong support for the histaminergic hypothesis in TS etiology and point to a possible role of histamine pathways in neuronal development.

INTRODUCTION

Gilles de la Tourette Syndrome (TS) is a multifactorial neurodevelopmental disorder resulting from the complex interaction of environment with genetic background. TS is characterised by the appearance of multiple motor and vocal tics, and high comorbidity rates with other neuropsychiatric disorders of childhood such as obsessive compulsive disorder (OCD), attention deficit hyperactivity disorder (ADHD) and autism, suggesting the possibility of a common aetiological background. Due to lack of education of health professionals and the general public, TS remains poorly understood and severely underdiagnosed in many countries despite the fact that recent investigations have reported a prevalence of 0.4–1% in populations of European origin. Results from large-scale studies and collaborative efforts have only very recently started to become available (eg, refs. 2–4) and the genetic basis of the disorder remains, so far, elusive, although multiple chromosomal regions have been implicated, providing several promising leads that warrant further investigation.

Tics are thought to result from dysfunctions in cortical and subcortical regions that are involved in habit formation, including the basal ganglia and related cortical and thalamic structures. TS has traditionally been viewed as a disorder of dopaminergic neurotransmission; however, other transmitters and neuromodulators have also been implicated. In fact, the recent implication of the L-histidine decarboxylase (HDC) gene in TS aetiology has raised the intriguing hypothesis of the involvement of histaminergic neural pathways in the onset of the disorder. HDC is highly conserved throughout different species and catalyses the oxidative decarboxylation of histidine to histamine.6–8 Recent studies have also implicated the oxidative decarboxylation of histidine to histamine.8–10

Sequencing of all genes within the linked interval led to the discovery of a rare coding mutation, a premature termination codon (p. W317*, c.951G>A) in exon 9 of the HDC gene. The mutation could not be found in any of the 3000 control individuals of Western European origin who were screened. At the same time, resequencing of the coding region of HDC in 720 patients with TS and 360 controls revealed no additional nonsense variants, demonstrating the fact that the nonsense mutation identified in the index family is extremely rare.

Since the original study that implicated HDC in TS aetiology, Lei et al11 screened the HDC gene for exonic mutations in 100 Chinese Han patients with TS, and could only find three variants, not predicted to result in amino acid changes: a C>T intronic transition (IVS1 +52C>T), which did not affect the splicing site, a synonymous C>A transition (c.426C>A) in exon 4 and a synonymous G>A transition (c.1743G>A) in exon 12.9 The first genomewide association study for TS only produced a weak association signal close to the HDC

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gene (p value of 0.02 with rs7166052). However, a recent genomewide scan for de novo or transmitted rare CNVs in TS found enrichment of genes within histamine receptor signalling pathways. Furthermore, a genomewide study of 95 French Canadian trios with familial history of TS showed association to a chromosome 15 microsatellite marker (D15S1016) that lies within the same interval found to be linked with TS in the original study that implicated the HDC gene in TS aetiology.

Following up on the novel hypothesis of the involvement of HDC and histaminergic neural pathways in TS aetiology, we investigated the possible association of variation across the HDC gene in a large sample of families originating from Canada as well as multiple countries in Europe. We find significant association to the HDC gene in our sample, providing strong support for the role of histamine in the genesis and mediation of tics, and pointing to new lines of research in this area.

MATERIAL AND METHODS

Genetic association study samples

Five independently collected samples of TS nuclear families were analysed (total of 520 families). Details of age of onset per sample, sex distribution as well as comorbid OCD and ADHD are shown in online supplementary table S1. The Tourette Syndrome Genetics—Southern and Eastern Europe Initiative sample included European-descent families of Polish (36 trios), Italian (50 trios), Hungarian (84 trios), Greek (10 trios) and Albanian origin (6 trios). Assessment was performed by on-site clinicians using the tools provided by the TS Association International Consortium for Genetics. TS was ascertained according to DSM-IV-TR criteria for Italy, Hungary, Albania and Greece, and DSM-IV for Poland. Our study also included two independent samples of German descent. The first was the German Collaborative TS Research Group sample (96 trios), collected using DSM-III-R criteria. Our second sample of German ancestry was collected in Munich (69 trios), using DSM-IV-TR criteria. Our fourth European sample was of Spanish ancestry (19 families), collected using DSM-IV-TR criteria. Finally, we also analysed a single sample of TS trios collected in Canada (150 families), using DSM-III-R criteria. Differences between DSM-III-R, DSM-IV and DSM-IV-TR criteria are shown in online supplementary table S1. The Tourette Syndrome Genetics—Southern and Eastern Europe Initiative sample included European-descent families of Polish (36 trios), Italian (50 trios), Hungarian (84 trios), Greek (10 trios) and Albanian origin (6 trios). Assessment was performed by on-site clinicians using the tools provided by the TS Association International Consortium for Genetics. TS was ascertained according to DSM-IV-TR criteria for Italy, Hungary, Albania and Greece, and DSM-IV for Poland. Our study also included two independent samples of German descent. The first was the German Collaborative TS Research Group sample (96 trios), collected using DSM-III-R criteria. Our second sample of German ancestry was collected in Munich (69 trios), using DSM-IV-TR criteria. Our fourth European sample was of Spanish ancestry (19 families), collected using DSM-IV-TR criteria. Finally, we also analysed a single sample of TS trios collected in Canada (150 families), using DSM-III-R criteria. Differences between DSM-III-R, DSM-IV and DSM-IV-TR criteria are minimal; the upper age limit of onset is 18 in DSM-IV (and DSM-IV-TR) and 21 in DSM-III-R, and the ‘marked distress’ criterion, possibly pointing to more severe cases, only appears in DSM-IV (applied only to the Polish families). For all samples, collection was approved by local Ethics Boards and informed consent was taken from all participating individuals or their parents.

Selection of single nucleotide polymorphisms, genotyping and statistical analysis

Using the HapMap CEPH European population as reference, we selected tagging single nucleotide polymorphisms (SNPs) in order to capture variation for a 40 kb region including the HDC gene. A total of 12 tSNPs were selected (see online supplementary table S2) capturing variation at an additional nine SNPs with a mean r² of 0.968. All selected SNPs are intronic. Genotyping was performed using the KASP genotyping method, a competitive allele-specific assay based on Fluorescent Resonance Energy Transfer from LGC Genomics and Kbioscience (http://www.lgcgenomics.com). The average genotyping success was greater than 99%. The linkage disequilibrium (LD) structure of each gene in the different studied populations was analysed using Haploview.15 In order to test for association of the studied SNPs with the TS phenotype, the transmission test for linkage disequilibrium (TDT) was performed, as also implemented in Haploview.15 We report here association results on single SNPs as well as SNP haplotypes of haplotype blocks defined with the Gabriel et al.16 criteria (tests with one degree of freedom, as implemented by Haploview). In order to correct for multiple comparisons, 1000 permutations were performed (using Haploview) and the adjusted p values are also reported here (table 1).

RESULTS

Genotyping results for all SNPs conformed to Hardy–Weinberg equilibrium proportions and Mendelian transmission within families. Rare allele frequencies for each analysed sample are shown in online supplementary figure S1. Since the transmission disequilibrium test (TDT) is robust to population stratification, we were able to perform analysis for the complete sample that was available to us, as well as families within each individual population. Single SNP analysis for association with the TS phenotype in the complete sample (520 families) produced

Figure 1  Linkage disequilibrium structure of studied region covering the l-histidine decarboxylase (HDC) gene (chromosome 15: 48 316 000–48 356 000 NCBI build 36). Unrelated individuals from a sample of 520 nuclear families were analysed using Haploview and haplotype blocks were defined using the Gabriel et al.16 criteria and visualised with the software Haplot. HDC variants found in individuals with Gilles de la Tourette Syndrome (TS) in previous studies are also shown. The triangle indicates the position of the originally described mutation that led to a premature termination codon (p. W317*, c.951G>A) in a family with TS, and the stars indicate the position of variants described in Chinese Han patients with TS (not predicted to result in amino acid changes). Access the article online to view this figure in colour.
The studied sample consists of 520 TS nuclear families of Greek, Albanian, Italian, Hungarian, Polish, Spanish, German and Canadian origin. The permutation p value was computed after 1000 permutations of the datasets. Results for the associated haplotype blocks 1 and 2 are shown (figure 1, block 1:SNPs rs854150-rs854151; block 2:SNPs rs2070596-rs2070595-rs2853766-rs1894236). Statistically significant signals (permutation p-value lower than 0.05) are shown in bold. T:U, transmitted-untransmitted; tSNP, tagging SNP.

### Table 1: Transmission test for linkage disequilibrium for l-histidine decarboxylase variation tSNPs and haplotypes

<table>
<thead>
<tr>
<th>Rs#</th>
<th>Over-transmitted allele</th>
<th>T:U</th>
<th>χ²</th>
<th>p Value</th>
<th>Permutation p value</th>
</tr>
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<tbody>
<tr>
<td>rs854150</td>
<td>G</td>
<td>284:220</td>
<td>8.127</td>
<td>0.0044</td>
<td>0.0290</td>
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<td>rs854151</td>
<td>G</td>
<td>205:190</td>
<td>0.57</td>
<td>0.4504</td>
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<tr>
<td>rs2070596</td>
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<td>192:154</td>
<td>4.173</td>
<td>0.0411</td>
<td>0.2640</td>
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<tr>
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<td>232:224</td>
<td>0.14</td>
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<td>rs2853766</td>
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<td>215:174</td>
<td>4.321</td>
<td>0.0376</td>
<td>0.2370</td>
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<td>rs1894236</td>
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<td>201:143</td>
<td>9.779</td>
<td>0.0018</td>
<td>0.0150</td>
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<tr>
<td>rs854157</td>
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<td>1.962</td>
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<td>0.009</td>
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<td>1.948</td>
<td>0.1628</td>
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<tr>
<td>rs854163</td>
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<td>223:188</td>
<td>2.981</td>
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<tr>
<td>rs1365503</td>
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<td>249:223</td>
<td>1.432</td>
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<table>
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<tr>
<th>Haplotype Frequency</th>
<th>T:U</th>
<th>χ²</th>
<th>p Value</th>
<th>Permutation p value</th>
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<tr>
<td>Block 1</td>
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<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>CA</td>
<td>0.620</td>
<td>224.5 : 286.7</td>
<td>7.56</td>
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<tr>
<td>GG</td>
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<td>227.2 : 195.0</td>
<td>2.465</td>
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<td>GA</td>
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<td>123.3 : 87.1</td>
<td>6.212</td>
<td>0.0127</td>
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<td>Block 2</td>
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<td></td>
<td></td>
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<tr>
<td>TTGC</td>
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<td>243.0 : 273.2</td>
<td>1.761</td>
<td>0.1845</td>
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<td>TGAC</td>
<td>0.248</td>
<td>218.0 : 173.0</td>
<td>5.179</td>
<td>0.0229</td>
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<tr>
<td>ATGT</td>
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<td>2.346</td>
<td>0.1256</td>
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DISCUSSION

Following up on the recent evidence about the involvement of the HDC gene and histaminergic neural pathways in the aetiology of TS, we studied variation across HDC for association with TS in a large sample of 520 nuclear families from seven European populations (Greece, Hungary, Poland, Italy, Albania, Germany and Spain) as well as a sample collected in Canada. We employed a tSNPs approach and identified strong overtransmission of alleles at two SNPs (rs854150 and rs1894236). Strong association was also found with a five-SNP haplotype carrying the protective rs1894236 allele, as well as two-SNP haplotypes carrying rs854150. Both SNPs that were found to be associated with TS in our study reside in intronic regions of HDC. However, it is worth noting that rs1894236 lies under one of the Encyclopedia of DNA Elements (ENCODE) project peaks for an enhancer-associated histone mark (UCSC genome browser NCBI36/hg18, http://genome.ucsc.edu). The ENCODE project has systematically mapped regions of transcription, transcription factor association, chromatin structure and histone modification, and histone marks are specific chemical modifications in histone sequence which may, for instance, enhance gene expression.
transcription. Thus, this particular region may be directly involved in regulation of transcription of the HDC gene. Individual population analysis produced additional signals of positive association. Thus, our results strongly support the histaminergic hypothesis for TS development.

Histamine is an evolutionary conserved signalling molecule that plays a central role in gastric acid secretion, innate and acquired immunity and immunomodulation, bronchoconstriction, vasodilation and neurotransmission. The neuronal histaminergic system is involved in a number of basic physiological functions, such as circadian rhythmicity, energy metabolism, neuroendocrine homeostasis, stress, sensory and motor functions, cognition, attention, learning and memory. Neuronal histamine is exclusively synthesised in the tuberomamillary nucleus of the hypothalamus (TMN), while histamine-positive fibres have been described projecting to cortex, hippocampus, basal ganglia, thalamus and virtually all other regions of the central nervous system. HDC is the key enzyme for histamine production from histidine. In the brain, its mRNA is expressed exclusively in the posterior hypothalamus. The specific regulation of HDC gene expression in the brain is not understood. However, neuroactive peptides and steroids are among the factors that control HDC transcription and degradation in various tissues and contexts (eg, oxidative stress). Recently, diurnal variation in the levels of HDC mRNA in the TMN has been observed in agreement with a role of neuronal histamine in regulating day–night patterns.

Histamine binds to and acts through four known G protein-coupled receptors (H1 through H4) and a polyamine binding site on glutamatergic N-Methyl-d-Aspartate (NMDA) receptors. High densities of H2R and H3R are found in the basal ganglia, especially on the principal neurons of the striatum, the GABAergic medium spiny neurons (MSN). Furthermore, H3R mRNAs in the cortex and in the substantia nigra pars compacta indicate the presence of H3 heteroreceptors on the major inputs to the striatum. Thus, histaminergic innervations of anatomical structures that have been related to TS has been confirmed. Furthermore, H3R regulates a variety of neurotransmitters, including dopamine and serotonin. On the other hand, Hdc−/− deficient mice have several traits relevant to features of TS and have shown decreased brain histamine and increased sensitivity to stereotypic behaviours upon administration of dopamine agonists. Such stimulant-induced movements, including rearing, sniffing and biting have previously been proposed as a model of human tics.

So far, the histaminergic system has not received as much attention as other monoaminergic systems of the brain. Classically established as a ‘peripherally’ important mediator of inflammation, the importance of histamine in neurotransmission and its role in neuropsychiatric disorders are only recently starting to be appreciated. For instance, it has been found severely affected in age-related neurodegenerative diseases such as Parkinson disease (PD) and Alzheimer disease (AD), while histamine receptors are becoming a prime target for pharmaceutical research and development and a number of agents have entered into clinical assessment as cognition enhancers for ADHD, schizophrenia, AD and PD. Adding to the multiple facets of neural histamine, our findings further support the implication of histaminergic dysfunction in TS aetiology, providing new insight. An interesting next step could be the functional characterisation of RNA transcripts in samples that carry the associated HDC genetic variants in order to help elucidate their role and clarify the possible involvement of histamine pathways in neurodevelopmental disorders.

**Author affiliations**

1Department of Molecular Biology and Genetics, Democritus University of Thrace, Alexandroupoli, Greece
2Department of Psychiatry, Ludwig-Maximilians-University Munich, Munich, Germany
3Toronto Western Research Institute, University Health Network, Toronto, Ontario, Canada
4Vadaskert Clinic for Child and Adolescent Psychiatry, Budapest, Hungary
5Materno-Infantile and Radiological Science Department, University of Catania, Catania, Italy
6Department of Child Psychiatry, Medical University of Warsaw, Warsaw, Poland
7Unidad de Trastornos del Movimiento, Servicio de Neurologia y Neurofisiologia Clinica/Seccion de Neuropatologia, Instituto de Biomedicina de Sevilla, Hospital Universitario Virgen del Rocio/CSCIC Universidad de Sevilla, Seville, Spain
8Department of Child and Adolescent Psychiatry, University of Duisburg-Essen, Essen, Germany
9Department of Genomics, Life and Brain Center, Institute for Human Genetics, University of Bonn, Bonn, Germany
10Department of Child and Adolescent, Psychiatry of the University of Cologne, Cologne, Germany
11Child and Adolescent Psychiatry Clinic, Sismagno General Hospital of Attica, Athens, Greece
12Department of Medical Chemistry, Molecular Biology and Pathobiocchemistry, Semmelweis University, Budapest, Hungary
13Centro de Investigación Biomédica en Red sobre Enfermedades Neurodegenerativas (CIBERNED), Seville, Spain

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**Contributors**

IK performed experiments and wrote the paper. SD contributed samples, participated in the design of experiments and analysis and in manuscript preparation. PS contributed samples, participated in the design of experiments and analysis and in manuscript preparation. 7TZ contributed samples, participated in the design of experiments and analysis and in manuscript preparation. CB contributed samples, participated in the design of experiments and analysis and in manuscript preparation. PN contributed samples, participated in the design of experiments and analysis and in manuscript preparation. MM contributed samples, participated in the design of experiments and analysis and in manuscript preparation. US contributed samples, participated in the design of experiments and analysis and in manuscript preparation. RR contributed samples, participated in the design of experiments and analysis and in manuscript preparation. ZT contributed samples, participated in the design of experiments and analysis and in manuscript preparation. PS contributed samples, participated in the design of experiments and analysis and in manuscript preparation. CB contributed samples, participated in the design of experiments and analysis and in manuscript preparation. US contributed samples, participated in the design of experiments and analysis and in manuscript preparation. PZ contributed samples, participated in the design of experiments and analysis and in manuscript preparation. PM contributed samples, participated in the design of experiments and analysis and in manuscript preparation. NM contributed samples, participated in the design of experiments and analysis and in manuscript preparation. AB contributed samples, participated in the design of experiments and analysis and in manuscript preparation. PP conceived the study, designed experiments and analysis and in manuscript preparation.

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**Competing interests**

None.

**Ethics approval**

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REFERENCES