



EDITOR'S
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SHORT REPORT

Mutations in *PRRT2* result in paroxysmal dyskinesias with marked variability in clinical expression

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ABSTRACT

Background Paroxysmal dyskinesias (PDs), a clinically and genetically heterogeneous group of episodic movement disorders, include kinesigenic PD (PKD), exercise-induced PD (PED) and non-kinesigenic PD (PNKD). These disorders are all transmitted as autosomal dominant traits with incomplete penetrance. Several PD-related genetic disorders, including PKD and familial infantile convulsions with paroxysmal choreoathetosis (ICCA), mapped to the same region on chromosome 16. Independent genetic studies have recently revealed that PKD can be caused by loss-of-function mutations in the proline-rich transmembrane protein 2 gene (*PRRT2*). We tested the hypothesis that other forms of PDs are also due to *PRRT2* mutations.

Methods/results The whole genomic region of *PRRT2* was sequenced in six Han Chinese families and 15 sporadic cases of PD-related phenotypes. The previously reported mutation, c.649dupC (p.R217Pfs*7), was found in two families with PKD, one family with ICCA, one family with PNKD-like phenotype, and two sporadic cases with PED. In an additional ICCA family, a novel frameshift mutation, c.904dupG (p.D302Gfs*38), was identified. A missense mutation, c.913G→A (p.G305R), and a synonymous substitution, c.1011C→T (p.G337G), were also detected in two sporadic PKD cases.

Conclusion This study shows that PKD, ICCA and some other PD-related phenotypes are part of the same phenotypic spectrum, caused by mutations in *PRRT2*. This underscores the complexity of the phenotypic consequences of *PRRT2* mutations.

INTRODUCTION

Paroxysmal dyskinesias (PDs) are a group of disorders characterised by episodes of involuntary movements that correspond to attacks of choreoathetosis or dystonia. They can be classified into three main categories: paroxysmal kinesigenic dyskinesia (PKD), paroxysmal exercise-induced dyskinesia (PED) and paroxysmal non-kinesigenic dyskinesia (PNKD).¹ PKD (OMIM 128200), also known as paroxysmal kinesigenic choreoathetosis or episodic kinesigenic dyskinesia 1, represents the most common type of PD, presenting with recurrent and brief attacks of dystonia and choreoathetosis. The diagnostic criteria for idiopathic PKD² includes sudden movements as the trigger, a short duration of attacks (<1 min), lack of loss of consciousness or pain during attacks, a good response to antiepileptic drug treatment, exclusion

of other organic diseases, and an onset age between 1 and 20 years. In contrast with PKD, PED is a neurological symptom with wide phenotypic variability and is usually triggered by prolonged physical exertion.³ PNKD (OMIM 118800) would not be precipitated by sudden movements, but alcohol, caffeine, stress, sleep deprivation and exercise can lower the threshold of attacks.⁴ Mutations in the myofibrillogenesis regulator-1 gene (*MR1*) have been revealed in several PNKD families.^{5–7}

PD may be variably associated with epilepsy.⁸ Benign familial infantile seizures (BFIS1, OMIM 601764; BFIS2, OMIM 605751), also called benign familial infantile convulsions, is mostly mentioned, as it can occur independently of or concurrently with PD.^{9–11} BFIS is an autosomal dominant disorder described as afebrile seizures occurring between 3 and 12 months of age, with good response to medication and no neurological sequelae.¹² Familial infantile convulsions with paroxysmal choreoathetosis (ICCA, OMIM 602066) shares overlapping clinical features with BFIS and PKD.^{11 13} Epilepsy may also coexist with PED.¹⁴ Autosomal recessive rolandic epilepsy with PED and writer's cramp (RE-PED-WC, OMIM 608105) shows phenotypic similarities to ICCA.¹⁵ Very interestingly, PKD, BFIS2, ICCA and RE-PED-WC all map to the same pericentromeric region on chromosome 16, suggesting that they are probably allelic disorders.^{8 12 13 15–17}

Two Chinese groups have recently identified *PRRT2*, encoding proline-rich transmembrane protein 2, as a causative gene for PKD.^{18 19} We thus performed mutation screening of *PRRT2* in our collected families and sporadic cases affected with PKD or other PD-related phenotypes.

SUBJECTS AND METHODS

We ascertained six Han Chinese families (see online supplementary figure 1 for pedigrees) and 15 sporadic cases of PKD, ICCA, PED or PNKD-like phenotypes (details in online supplementary table 1).

Clinical findings in affected individuals in families A and B are consistent with a diagnosis of ICCA. Individuals were considered as affected if they had either BFIS or PD, or if they had both a history of BFIS and PD. Nine affected individuals in family A were available for clinical evaluation. The traits of BFIS and PD in the family were

unconnected, with six and three individuals having isolated BFIS and PD, respectively. Nearly all cases of BFIS manifested as deviation of eyes to one side, an altered consciousness, and bilateral limb jerks with hypertonia. The seizures occurred mostly when awake and were isolated except that, in two cases (IV-7 and V-1), seizures happened during sleep and were clustered. None of the interictal EEGs showed epileptiform abnormalities between attacks, and CT scanning or MRI were normal in the cases examined. Both of the two affected individuals in family B had BFIS and PD. Notably, the characteristics of choreoathetosis in these two families were consistent with PED. The age of onset was 8–10 years, and remission seemed to be around 20 years. Choreoathetosis was of the dystonic type, occurring at rest, or could be provoked by anxiety, fatigue or exercise after a few seconds. The limbs of one side were affected, with short duration of attacks lasting <5 min. Response to antiepileptic drugs varied in both families.

Affected individuals in family C had a PNKD-like phenotype. The trunk and limbs on both sides were involved, with a long duration. The onset age of PD was about 7–8 years, except in the proband (IV-2), where the age was as young as 2 years with seemingly more severe symptoms. The age of remission was quite different, ranging from 2 to over 50. Alcohol, anxiety and fatigue were described as precipitants by affected individuals. Drinking alcohol was a definite trigger for III-5. Attacks could occur as often as 10–20 times per day. III-5 and IV-2 of family C described an obvious benefit of sleep during episodes. The anti-convulsive drugs were not effective as suggested before.⁴ The proband also had migraine. III-7, in particular, showed a typical generalised tonic-clonic type of seizure with normal EEG. The epilepsy began at the age of 20 with automatic remission in 2 years. Unfortunately, she, as well as II-2 and III-1, refused to participate in our genetic studies. No history of central nervous system disease or damage was found in any of the participating patients, and psychomotor development was normal on neurological examination.

In families D–F, clinical presentations of most affected individuals met the diagnostic criteria for PKD in all aspects.¹ Among the 15 sporadic cases, 10 had PKD (cases 1, 2, 4, 5, 6, 8, 9, 10, 12 and 13), four had PED (cases 3, 11, 14 and 15), and one had PNKD-like phenotype (case 7) (online supplementary table 1).

After obtaining informed consent, we collected peripheral blood samples for DNA from 11 individuals (six affected and five unaffected) in family A, the probands of families B–F, and all the 15 sporadic cases. For mutation screening of the *PRRT2* gene, we PCR-amplified the entire gene into four overlapping fragments and sequenced them directly after purification.¹⁹ We also

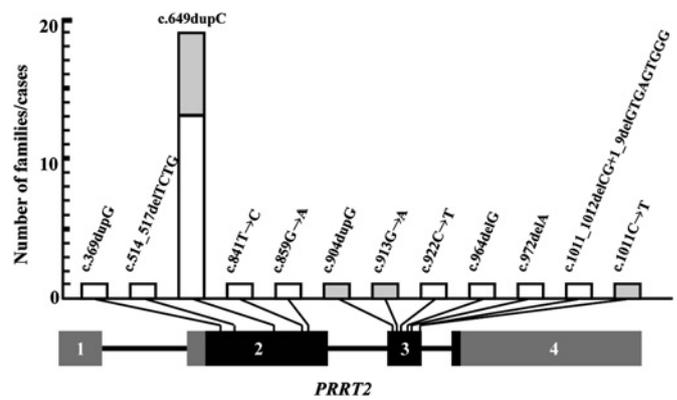


Figure 1 Gene structure of *PRRT2* with a summary of identified mutations in paroxysmal kinesigenic dyskinesia, infantile convulsions with paroxysmal choreoathetosis, and other paroxysmal dyskinesia-related phenotypes. For each identified mutation, its relative position in the *PRRT2* gene and the numbers of families/cases detected with the mutation are indicated. Light-grey bars represent mutations reported in the present study.

performed genomic sequencing of *PRRT2* in 192 unrelated Han Chinese controls.

RESULTS

We sequenced *PRRT2* in one affected individual per family and identified the reported c.649dupC (p.R217Pfs*7) mutation^{18 19} in families A, C, E and F (figure 1 and table 1). This mutation was detected in all available affected individuals of family A. In family B, we found a novel single base duplication (c.904dupG), producing a frameshift and premature termination codon (p.D302Gfs*38) (figure 1, table 1 and online supplementary figure 2). Sequence analysis of *PRRT2* in sporadic cases identified the c.649dupC (p.R217Pfs*7) mutation in two cases with PED, and a novel missense mutation, c.913G→A, in one with PKD (figure 1, table 1 and online supplementary figure 2). This missense mutation replaces a non-polar glycine (G) (online supplementary figure 3) with a basic arginine (R) at amino acid residue 305 (p.G305R). We also found a synonymous substitution, c.1011C→T (p.G337G), in a sporadic case with PKD. None of the identified mutations was found in the 192 controls. Consistent with the other two parallel reports on PKD,^{18 19} the mutations involved loss of function and suggest haploinsufficiency as the molecular mechanism.

To determine whether the PNKD-like phenotype in family C was caused by a mutation in *MR1*, we also sequenced this gene and found no pathogenic mutations (data not shown).

Table 1 Major clinical features in families/cases with identified *PRRT2* mutations

Subject	Main phenotype	Accompanied features	Trigger of PD	Mutation DNA	Amino acid
Family A	ICCA	–	Ax, Ex	c.649dupC	p.R217Pfs*7
Family B	ICCA	–	Ax, Ex	c.904dupG	p.D302Gfs*38
Family C	PNKD-like	Migraine	Ah, Ft, Ax	c.649dupC	p.R217Pfs*7
Family E	PKD	–	Sm	c.649dupC	p.R217Pfs*7
Family F	PKD	Migraine, FC	Sm	c.649dupC	p.R217Pfs*7
Case 5	PKD	FC	Sm	c.1011C→T	p.G337G
Case 13	PKD	–	Sm	c.913G→A	p.G305R
Case 14	PED	–	Ex	c.649dupC	p.R217Pfs*7
Case 15	PED	–	Ex, St	c.649dupC	p.R217Pfs*7

Ax, anxiety; Ex, exercise; FC, febrile convulsions; Ft, fatigue; ICCA, infantile convulsions with paroxysmal choreoathetosis; PD, paroxysmal dyskinesia; PED, paroxysmal exercise-induced dyskinesia; PKD, paroxysmal kinesigenic dyskinesia; PNKD, paroxysmal non-kinesigenic dyskinesia; Sm, sudden movement; St, startle.

DISCUSSION

With the use of next-generation sequencing, two Chinese groups have recently identified *PRRT2* as a causative gene for PKD.^{18 19} Chen and colleagues identified loss-of-function *PRRT2* mutations in all their eight study families.¹⁸ In three of their four study families, Li and colleagues found *PRRT2* mutations, including two frameshifts and one missense.¹⁹ They also showed that *PRRT2* was mutated in approximately one-third of the sporadic PKD cases (10/29).¹⁹ In the present study, we sequenced the *PRRT2* gene in three familial and 10 sporadic PKD cases, and detected the previously reported mutation, c.649dupC (p.R217Pfs*7),^{18 19} in two families, and a novel missense mutation in a sporadic case (table 1). Taken together, these results indicate that *PRRT2* mutations are responsible for most familial PKD and only a subset of sporadic PKD, and suggest the presence of genetic heterogeneity in PKD.

Previous genetic linkage studies in multiple families had suggested that *BFIS2*, *ICCA* and *RE-PED-WC* may be allelic with PKD.^{8 12 13 15–17} We have found *PRRT2* mutations in two Chinese families with *ICCA*, indicating that *ICCA* is indeed part of the same phenotypic spectrum caused by mutations in *PRRT2*. It is conceivable that *PRRT2* mutations are also responsible for *BFIS2* and *RE-PED-WC*. We unexpectedly identified *PRRT2* mutations in a Chinese family with a PNKD-like phenotype and in two sporadic cases of PED (table 1), further expanding the phenotypic spectrum caused by mutations in the same *PRRT2* gene and underscoring the complexity of the phenotypic consequences of *PRRT2* mutations. Our work, together with the previous genetic studies,^{8 12 13 15–17} provides genetic evidence that PKD, *ICCA* and other PD-related phenotypes may represent a phenotypic continuum.

We and others^{18 19} have together identified 12 distinct *PRRT2* mutations in familial and sporadic cases of PKD, *ICCA* or other PD-related phenotypes (figure 1). No obvious genotype–phenotype correlation was observed. Most strikingly, the c.649dupC (p.R217Pfs*7) mutation was detected in 19 of the 30 cases with *PRRT2* mutations (16 familial and 14 sporadic cases) (figure 1). We later typed a CA repeat sequence 139 kb centromeric to *PRRT2* in our familial and sporadic cases with the c.649dupC mutation (table 1)¹⁹ and found that six sporadic cases lacked the allele linked to the mutation in family A (data not shown). These facts, together with the haplotyping data of Chen and colleagues,¹⁸ suggest that the cytosine (C) tract at nucleotides 641–649 of the *PRRT2* cDNA (5'-CCCCCCCC-3') may be the hotspot for a single C duplication. Further haplotype analysis is still needed to exclude the possibility of a founder effect.

Frequent detection of truncating *PRRT2* mutations has suggested haploinsufficiency as the underlying mechanism.^{18 19} Li and colleagues also found three PKD-causing missense mutations in *PRRT2*.¹⁹ The c.913G→A (p.G305R) missense mutation detected in the present study (figure 1 and table 1) changed a highly conserved amino acid residue (online supplementary figure 3). Assessment using PolyPhen-2 (<http://genetics.bwh.harvard.edu/pph2/>) predicted a damaging effect of this mutation. The synonymous c.1011C→T (p.G337G) is located at a position very close to the exon–intron junction (figure 1), suggesting that it may affect splicing. Neither of the two substitutions was detected in our ethnically matched controls. At present, the pathogenicity of c.1011C→T (p.G337G) lacks supportive evidence.

In summary, we found *PRRT2* mutations in 38.1% (9/21) of our study cases (six family probands and 15 sporadic) and demonstrated that PKD, *ICCA* and other PD-related phenotypes

may be caused by mutation in the same *PRRT2* gene. Further molecular studies would provide insight into the connection between epilepsy and paroxysmal movement disorders.

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Competing interests None.

Patient consent Obtained.

Ethics approval This study was approved by the ethics committee of Peking Union Medical College Hospital, Chinese Academy of Medical Sciences.

Contributors QLi, LYC and XZ analysed all data and wrote the paper. QLi, ZQ, JYL, LS, WY and YL generated the experimental data, and QLi, XHW, QLu, XQZ, LQ, LWW and XQL provided all clinical findings and patient samples.

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Data sharing statement Primer sequences not given in the text are available upon request.

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