Recessive mutations in the CYP4V2 gene in East Asian and Middle Eastern patients with Bietti crystalline corneoretinal dystrophy

J Lin, K M Nishiguchi, M Nakamura, T P Dryja, E L Berson, Y Miyake

Background: Bietti crystalline corneoretinal dystrophy (BCD) is an autosomal recessively inherited disorder characterised by tiny yellowish glittering retinal crystals, choroidal sclerosis, and crystals in the peripheral cornea, associated with progressive night blindness. CYP4V2, encoding a member of cytochrome p450 (CYP450) protein family, was recently identified as the causative gene.

Methods: We recruited 11 unrelated patients with BCD and characteristic clinical features; eight of Japanese, two of Middle Eastern, and one of Chinese ancestry. Genomic DNA was extracted from peripheral blood leucocytes, and all 11 exons and the flanking intron splice sites of the CYP4V2 gene were amplified and sequenced. A complete ophthalmological examination was performed.

Results: We found recessive mutations in the CYP4V2 gene in all of the 11 patients. Two novel mutations, L173W and Q450X, were identified in a Japanese patient and two unrelated patients from Middle Eastern countries, respectively. Each patient was a homozygote. A previously reported mutation IVS6-8_810delinsGC was identified in seven unrelated Japanese patients and the Chinese patient with BCD. All patients with BCD shared a characteristic fundus appearance with numerous intraretinal crystal deposits and atrophy of the retinal pigment epithelium. However, the clinical findings, including electroretinograph recordings, indicated that there was considerable variation in the degree of visual dysfunction even among patients of similar ages carrying the same mutation.

Conclusions: Defects in CYP4V2 are the main cause of BCD. The IVS6-8_810delinsGC mutant allele may be especially prevalent among patients with BCD in East Asian countries, resulting from a single founder. Variation of disease severity suggests that environmental or additional genetic factors influence the course of the retinal disease.

Bietti crystalline corneoretinal dystrophy (BCD) is an autosomal recessively inherited disorder, which was first reported in 1937 by Bietti. It is characterised by multiple small glistening intraretinal crystalline deposits scattered throughout the posterior pole associated with progressive atrophy of the retinal pigment epithelium (RPE) and choroidal sclerosis. The crystalline deposits are also present in circulating lymphocytes, and in the limbal cornea in some patients. The disease typically manifests itself between the second and fourth decades of life with progressive night blindness, decreased visual acuity, and constriction of the visual fields. The disease progresses slowly, and the vision is severely impaired in the late stages. Although BCD is a relatively rare disease, the diagnosis is straightforward because of the characteristic clinical features. Patients with BCD have been reported from most parts of the world, but it appears to be more common in East Asian populations, especially in China and Japan.

The gene responsible for BCD has been mapped to chromosome 4q35, and within this region CYP4V2, encoding a member of cytochrome p450 (CYP450) protein family, was recently identified as the causative gene. The gene contains 11 exons spanning 21.7 kb, and is expressed in various tissues, most abundantly in the retina. The CYP450 protein family is a group of enzymes that use iron to oxidise various substrates, including potentially harmful substances, thereby making them more water soluble and promoting their biological processing. Some members of the CYP450 family are known to have a role in lipid metabolism, and it is possible that the protein encoded by CYP4V2 plays a role in processing lipids such as fatty acids or a role in steroid metabolism. This speculation is based on a published biochemical analysis of cultured lymphocytes from patients with BCD, which showed abnormally high levels of triglycerides and cholesterol, and the absence of two fatty acid binding proteins.

There has been only one previous report of mutations in CYP4V2 in BCD. We undertook this study to better understand the range of pathogenic mutations and to search for possible non-allelic heterogeneity. Here we report our mutation screen of the CYP4V2 gene in 11 unrelated patients with BCD of East Asian or Middle Eastern ancestry and the clinical findings in the patients.

PATIENTS AND METHODS

This study was performed in accordance with the tenets of the Declaration of Helsinki, and was approved by the internal review boards at the Nagoya University Graduate School of Medicine, Massachusetts Eye and Ear Infirmary, and Harvard Medical School. For the study, 11 unrelated patients who had been diagnosed with BCD on the basis of their characteristic clinical features were recruited. Eight of the patients were of Japanese ancestry and were ascertained at the Department of Ophthalmology, Nagoya University Graduate School of Medicine. Two patients of Middle Eastern ancestry and one of Chinese ancestry were from the Berman-Gund Laboratory for the Study of Retinal Degenerations, Harvard Medical School. None of the parents or the children of the patients were affected, and five of the unrelated patients were the offspring of consanguineous relationships. The ophthalmological evaluations included the measurement of best corrected visual acuity, biomicroscopy, ophthalmoscopy, fundus photography, Goldmann kinetic perimetry, and electroretinograms (ERGs). Full field ERGs were recorded with the previously published methods using separate protocols at Nagoya University and at Harvard Medical School. After informed consent was obtained, blood
samples were taken from the patients and, in some cases, their relatives. Normal subjects without a history of a retinal disease were recruited as controls.

Genomic DNA was extracted from peripheral blood leukocytes using standard methods. The exons containing the open reading frame of the CYP4V2 gene and the flanking intron splice sites were individually amplified by PCR. The amplified DNA fragments were purified and directly sequenced using a dye terminator cycle sequencing kit (version 3.1; Applied Biosystems; Foster City, CA, USA) and ABI automated DNA sequencers (model 3100; Applied Biosystems). The numbering of the bases of the cDNA and genomic sequences in this article is according to Genbank accession numbers XM_209612 and NC_000004, respectively.

Prediction of the structure of CYP4V2 was based on comparison and analysis of crystal coordinates from mammalian CYP2C5 (Protein Data Bank (PDB) ID 1dt6) and bacterial CYPBM3 (PDB ID 2hpd).

Partial alignment of amino acid sequences of six eukaryotic CYP4V2 homologues selected using an online analysis tool

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**Table 1** Summary of mutations in CYP4V2 and clinical information in patients with BCD

<table>
<thead>
<tr>
<th>ID</th>
<th>Ethnicity</th>
<th>Nucleotide*</th>
<th>Protein</th>
<th>Age</th>
<th>Sex</th>
<th>Vo (R, L)</th>
<th>vERG amplitudes</th>
<th>R, L (μV)</th>
<th>Corneal deposit</th>
</tr>
</thead>
<tbody>
<tr>
<td>252</td>
<td>Japanese</td>
<td>IVS6-8_810delinsGC</td>
<td>Skip exon 7</td>
<td>50</td>
<td>M</td>
<td>1.0, 0.6</td>
<td>120, ND</td>
<td></td>
<td></td>
</tr>
<tr>
<td>259</td>
<td>Japanese</td>
<td>IVS6-8_810delinsGC</td>
<td>Skip exon 7</td>
<td>52</td>
<td>F</td>
<td>0.5, 1.0</td>
<td>ND, 27</td>
<td></td>
<td></td>
</tr>
<tr>
<td>260</td>
<td>Japanese</td>
<td>IVS6-8_810delinsGC</td>
<td>Skip exon 7</td>
<td>46</td>
<td>F</td>
<td>0.7, 0.9</td>
<td>207, ND</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>265</td>
<td>Japanese</td>
<td>IVS6-8_810delinsGC</td>
<td>Skip exon 7</td>
<td>38</td>
<td>F</td>
<td>0.2, 1.5</td>
<td>274, 296</td>
<td></td>
<td></td>
</tr>
<tr>
<td>272</td>
<td>Japanese</td>
<td>IVS6-8_810delinsGC</td>
<td>Skip exon 7</td>
<td>52</td>
<td>M</td>
<td>0.9, 0.7</td>
<td>385, ND</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>303</td>
<td>Japanese</td>
<td>IVS6-8_810delinsGC</td>
<td>Skip exon 7</td>
<td>54</td>
<td>M</td>
<td>0.5, 0.5</td>
<td>311, ND</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>312</td>
<td>Japanese</td>
<td>IVS6-8_810delinsGC</td>
<td>Skip exon 7</td>
<td>47</td>
<td>F</td>
<td>1.2, 1.2</td>
<td>312, ND</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>270</td>
<td>Japanese</td>
<td>518T → R</td>
<td>L173W</td>
<td>54</td>
<td>F</td>
<td>1.0, 1.2</td>
<td>296, ND</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>288-001</td>
<td>Chinese</td>
<td>IVS6-8_810delinsGC</td>
<td>Skip exon 7</td>
<td>26</td>
<td>F</td>
<td>0.5, 0.6</td>
<td>329, 323</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>288-002</td>
<td>Middle Eastern</td>
<td>1348C → T</td>
<td>Q450X</td>
<td>21</td>
<td>F</td>
<td>0.4, 0.2</td>
<td>229, 206</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>288-003</td>
<td>Middle Eastern</td>
<td>1348C → T</td>
<td>Q450X</td>
<td>45</td>
<td>M</td>
<td>0.8, 0.05</td>
<td>71, 29</td>
<td>+</td>
<td></td>
</tr>
</tbody>
</table>

ND, not determined. *All patients were homozygotes except 288-001, who is a presumed compound heterozygote; vERG amplitudes from dark-adapted single white flash ERGs extracting mixed rod and cone responses. For the first eight patients (patients 252–270), ERGs were recorded at Nagoya University (normal range >314 μV) and, for the last three patients (patients 288-001, 288-002, and 288-003), at Harvard Medical School (normal range >350 μV). The predicted consequence of exon 7 skipping is in frame deletion of residues 268–329.
L173

| Human CYP4V2       | EQANLYKLE       |
| Mouse Cyp4v3       | EQANLYNKKLE     |
| Rat LOC361155      | EQANLYNKKLE     |
| Mosquito 1272634   | KQSYVRQLK      |
| Nematode 4m169     | EQSKLVQKLC      |
| Thale cress a3g25180 | SEVNTLHLY     |

Figure 2. Partial amino acid sequence alignment surrounding the site of the L173W mutation in six eukaryotic CYP4V2 homologues: human CYP4V2 (NP_997235.1), mouse Cyp4v3 (NP_598730.1), rat LOC361155 (XP_341441.1), mosquito 1272634 (XP_311534.1), nematode 4m169 (NP_502152.2), and thale cress a3g25180 (NP_189154.1). An arrow indicates the position of the residue altered by the novel missense mutation L173W.

Table 1. Location and frequency of mutations and polymorphisms in the CYP4V2 gene.

<table>
<thead>
<tr>
<th>Allele frequency</th>
<th>Homozygote patients with IVS6-8_810delinsGC</th>
</tr>
</thead>
<tbody>
<tr>
<td>G</td>
<td>0.43</td>
</tr>
<tr>
<td>A</td>
<td>1.00</td>
</tr>
<tr>
<td>delCT</td>
<td>1.00</td>
</tr>
<tr>
<td>T</td>
<td>1.00</td>
</tr>
<tr>
<td>delinsGC</td>
<td>1.00</td>
</tr>
<tr>
<td>A</td>
<td>0.23</td>
</tr>
<tr>
<td>G</td>
<td>0.43</td>
</tr>
</tbody>
</table>

RESULTS

We identified four different mutations, L173W (c.518T→G), IVS6-8_810delinsGC, H331P (c.992A→C), and Q450X (c.1348C→T), in the CYP4V2 gene. Mutations were present in each of the 11 unrelated patients with BCD who were evaluated (table 1). Every patient was homozygous except one (patient 288-001), who was a presumed compound heterozygote. Two of the mutations, L173W and Q450X, are previously unreported; they were identified in one Japanese patient (patient 270) and in two Middle Eastern patients (patients 288-002 and 288-003), respectively. A representative pedigree and DNA sequence chromatogram for patients harbouring these novel mutations are presented in fig 1.

The novel mutation L173W was interpreted as likely to be pathogenic for the following reasons. Firstly, leucine at position 173 is highly conserved among all six close eukaryotic homologues ranging from Homo sapiens to Arabidopsis thaliana (fig 2). Secondly, based on a structural analysis of the cytochrome p450 protein homologues CYPBM3 and CYP2C5, L173 resides within an α helix presumably interacting with at least four other downstream helices. Thus, a change of the leucine at position 173 to tryptophan may result in a gross structural alteration possibly affecting the proper folding of the protein. Thirdly, the mutation was not found in an analysis of 100 Japanese normal controls.

The novel mutation Q450X was considered to be a pathogenic null mutation, as mRNA containing the predicted early stop codon would most likely be subject to nonsense mediated decay without being translated into a protein. Even if the translated protein is produced, it would lack amino acid sequences that are expected to coordinate the haem iron that binds substrate. This change was also not found among 90 healthy control individuals from North America.

Seven other Japanese patients with BCD were homozygotes for the IVS6-8_810delinsGC mutation that affected the IVS6 splice acceptor site. This mutation has been reported in other Japanese and Chinese BCD patients. Analysis of lymphocyte mRNA from patients indicated that the mutation causes in frame skipping of exon 7. Our Chinese patient had two mutations, IVS6-8_810delinsGC and H331P, and was a presumed compound heterozygote. There were no family members available for this to be confirmed by segregation analysis. Another BCD patient with the same combination of CYP4V2 alleles has been previously reported.

We categorised three missense changes, L22V (c.64C→G), E259K (c.775C→T), and R433Q (c.1328G→A), as likely non-pathogenic because they were all found in patients with other pathogenic recessive mutations in CYP4V2. An isoaccepting change A270A (c.810T→G) was found heterozygously in two patients and homozygously in one. This change, and the two likely non-pathogenic missense changes L22V and E259K, have been previously reported in an SNP database (www.ncbi.nlm.nih.gov/SNP). Six intron changes (IVS1–22–C→A, IVS2+75C→T, IVS3–40delCT, IVS4–25G→T, IVS6–7C→T, and IVS9+45A→G) were found in patients with other pathogenic recessive mutations in CYP4V2 described above. These changes were predicted not to affect RNA splicing, as assessed by online splice site prediction software (www.fruitfly.org/seq_tools/splice22), and were interpreted as non-pathogenic polymorphisms.

In the seven Japanese patients homozygous for the frequent IVS6-8_810delinsGC mutation, five non-pathogenic variants located between intron 1 and exon 6 (IVS1–22C→A, IVS2+75C→T, IVS3–40delCT, IVS4–25G→T, and c.775C→A) were observed in the homozygous state. The polymorphisms located in exon 1 (c.64C→G) and in intron 9 (IVS9+45A→G) were frequently identified in a screen of 50-100 normal Japanese subjects, and identified in some, but not all, of the BCD patients (fig 3).

The clinical characteristics of the BCD patients are summarised in table 1. Seven patients complained of night blindness. Visual acuities of the 11 patients ranged from normal (1.0) to severely reduced (0.05). Eight patients had
acuity in at least one eye better than 0.8. Slit lamp examination revealed peripheral corneal crystalline deposits in eight patients. Goldmann kinetic visual fields showed a central, a paracentral, or a ring scotoma in the patients (data not shown). All patients had characteristic numerous small retinal crystalline deposits concentrated in the posterior pole (a representative fundus photograph is presented in fig 4A). There were varying degrees of RPE atrophy and attenuation of the choriocapillaris at the posterior pole; these abnormalities sometimes extended to the midperiphery. Full field ERGs showed variable amounts of rod and cone function ranging from normal to severely reduced even among patients carrying the same mutation (representative ERGs dark-adapted are shown in fig 4B). The b-wave amplitudes for dark adapted single flash rod plus cone responses are summarised in table 1.

DISCUSSION
We identified mutations in the CYP4V2 gene in all 11 unrelated patients we evaluated with typical BCD. These patients were of East Asian or Middle Eastern ancestry. A previous report showed that CYP4V2 mutations were found in 23 of 25 unrelated patients with BCD.10 Together with our results, it is apparent that defects in CYP4V2 account for most, if not all, cases of BCD.

In this screen, we found the homozygous IVS6-8_810delinsGC mutation in seven of eight unrelated Japanese patients with BCD. Consistent with the results in a previous study, which found the same mutation in seven of eight unrelated Japanese BCD patients,10 our findings indicate that this mutation is the major cause of BCD in Japan. The same mutation has been detected in seven of 10 unrelated Chinese patients,17 and in a Chinese patient in this study. Considering that this mutation is frequently identified in both Japanese and Chinese patients and that it has never been detected in BCD patients of other ethnic backgrounds, it is likely that all patients with this mutation descend from a single ancestor. This speculation would be supported by the observation that six closely linked polymorphic markers were identical in a homozygous state in all seven patients with the homozygous IVS6-8_810delinsGC mutation (fig 3). However, the founder of this mutation is probably a very distant ancestor because the region of the conserved linked markers is small, extending only 6.7–17.1 kb (fig 3).23

All patients with the CYP4V2 gene mutation shared similar characteristic fundus features, including crystalline deposits and RPE atrophy. However, the reduction of visual acuity and the amount of visual field loss varied among patients. Full field ERGs, reflecting overall retinal function, showed remarkable variability in amplitudes, as has been previously reported.24 Our results indicate that severity differed even among patients at similar ages with the same IVS6-8_810delinsGC mutation, raising the possibility that factors besides the primary gene defect influence the course of the retinal disease.

It has been reported that corneal crystalline deposits are present in one quarter to one third of patients with BCD.5 We observed them in eight of 11 patients by slit lamp examination. However, as corneal crystalline deposits are very subtle, and in some patients can be detected clearly only by specular microscopy,25 the true number of patients with corneal deposits may be higher.

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