A case control and family based association study of the neuregulin1 gene and schizophrenia

X Zhao, Y Shi, J Tang, R Tang, L Yu, N Gu, G Feng, S Zhu, H Liu, Y Xing, S Zhao, H Sang, Y Guan, D St Clair, L He

Key points

- In a sample of 1724 Han Chinese individuals, 3 ummSNPs (single nucleotide polymorphisms) and 2 microsatellites were genotyped. Case control and TDT analyses were performed to investigate whether the 5′ end of neuregulin 1 (NRG1) was associated with schizophrenia.
- Owing to the low frequency found of the reported at risk haplotype (<1%), no association could be detected with schizophrenia. However, another haplotype was found to be significantly associated with schizophrenia in both case control (p = 0.0057) and TDT analyses (p = 0.0043).
- The results support the view that NRG1 may be an important factor in the aetiology of schizophrenia.

Data from twin, family, and adoption studies provide strong evidence that genetic factors play a major aetiological role in schizophrenia. By a series of linkage studies, chromosome 8p has been implicated as a region harbouring a schizophrenia susceptibility gene.1,4 Recently, Stefansson and colleagues reported that neuregulin 1 (NRG1), located in 8p21-12, may be involved in the aetiology of schizophrenia.1,4 In their linkage and association studies, a 290 kb core at risk haplotype at the 5′ end of NRG1 was found to be strongly associated with schizophrenia in Icelandic and Scottish populations. This haplotype contains the first exon of NRG1, which encodes a part of glial growth factor 2 (GGF2). Deficiency of glial growth factors has been presumed to be implicated in the pathogenesis of schizophrenia.5 Furthermore, NRG1 mutant mice have fewer functional N-methyl D-aspartate (NMDA) receptors than wild type mice, and display stereotypic behavioural abnormalities similar to those of normal mice treated with the psychogenic drug phenylcyclidine.6

This core at risk haplotype was defined by five single nucleotide polymorphisms (SNP8NRG221132, SNP8NRG221533, SNP8NRG241930, SNP8NRG243177, SNP8NRG433E1006) and two microsatellites (478B14-848, 420M91395). The frequency of this haplotype in schizophrenic individuals was higher than in controls; in Icelandic samples the frequency was 15.4 (7.5%; p = 0.000087).7 The first replication using Scottish samples revealed a similar result at 10.2 (5.9%; p = 0.00031).8 Another replication performed by Williams et al with British or Irish samples used one SNP and the two microsatellites of the core at risk haplotype. However, the association was much weaker at 9.5 (7.5%; p = 0.04).9 Yang et al reported other markers located in the middle of NRG1 and associated with schizophrenia, in a Chinese population.10 Another independent analysis using 13 microsatellites found two groups of haplotypes, which were significantly associated with schizophrenia, in a Chinese population.11

All these studies suggest that NRG1 may be a susceptibility gene for schizophrenia. However, there are differences in the findings and more replications are needed. Here we present results from the haplotype analysis using both case control and TDT. In our studies, only three of the five SNPs (SNP8NRG221533, SNP8NRG241930, SNP8NRG243177) and the two microsatellites reported by Stefansson et al were genotyped. The marker SNP8NRG221132 was abandoned because of low heterozygosity. We genotyped more than 200 individuals at this locus, but no allele A was found. In all, 1724 Han people Chinese participated in our research, including 369 subjects with schizophrenia, 299 controls in the case control study, and 352 family trios in the TDT study.

MATERIALS AND METHODS

For the case control investigation, 369 unrelated persons with schizophrenia (178 were male and 191 were female, with a mean age of 41.56 years, SD = 14.35) and 299 control individuals (148 were male and 151 were female, with a mean age of 31.26 years, SD = 9.07) were recruited. The cases included 177 patients from Shanghai and 192 patients from Jilin; the controls comprised 109 people from Shanghai and 190 people from Jilin.

For the TDT study, 352 unrelated schizophrenia probands (187 were male and 165 were female, with a mean age of 23.86 years, SD = 6.60) and their biological parents were recruited. All subjects were Han Chinese in origin. Those with schizophrenia were diagnosed strictly according to the criteria of DSM-III-R (American Psychiatric Association, 1987). Written informed consent was obtained from either the participants or the participants’ relatives, after the procedure had been fully explained.

High molecular weight genomic DNA was prepared from venous blood using the standard phenol chloroform extraction. SNPs were genotyped through TaqMan® technology on an ABI7900 system and probes and primers were designed by the Assay-by-Design® service of Applied Biosystems (San Jose, CA, USA). The standard PCR reactions of 5 µl were carried out using TaqMan® Universal PCR Master Mix reagent kits as in the guidelines. During assay development, microsatellite 478B14-848 was amplified with upper primer 5′-ctg gta tga cca aa- 3′ and lower primer 5′-cat gtc caa ctg aag agg- 3′ and lower primer 5′- cag aga cat cct gta tga cca aa- 3′, microsatellite 420M9-1395 was amplified with upper primer 5′- cag aat ttt cag aat ttc ctt- 3′ and lower primer 5′-att cca gtt aaa tga aag agg c- 3′. Each upper primer was fluorescently labelled. PCR products were electrophoresed on MegaBACE 1000 instruments (Amersham Biosciences, Amersham, UK). In order to compare the results, we aligned our allele histograms with allele histograms for a Chinese population genotyped at deCODE.
Table 1  Allele frequencies of all markers and association analysis of individual markers

<table>
<thead>
<tr>
<th>Marker*</th>
<th>Allele</th>
<th>Case control study</th>
<th>TDT study</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Case†</td>
<td>Control†</td>
</tr>
<tr>
<td>SNP8NRG221533</td>
<td>C</td>
<td>396(0.54)</td>
<td>263(0.52)</td>
</tr>
<tr>
<td>SNP8NRG241930</td>
<td>G</td>
<td>681(0.93)</td>
<td>560(0.95)</td>
</tr>
<tr>
<td>SNP8NRG243177</td>
<td>T</td>
<td>396(0.57)</td>
<td>325(0.56)</td>
</tr>
<tr>
<td>478B14-848</td>
<td>4</td>
<td>427(0.58)</td>
<td>296(0.54)</td>
</tr>
<tr>
<td>420M9-1395</td>
<td>–2</td>
<td>498(0.68)</td>
<td>402(0.67)</td>
</tr>
</tbody>
</table>

*Marker names used by Stefansson et al. 2002. †Allele frequencies are shown in parentheses. ‡Global p values of the χ² statistics. 

For case control analysis, the statistical significance of differences in the allele and haplotype frequency distributions between patients and controls was estimated using the program Clump 2.2.10 The software was also used to compare genotype frequencies between Shanghai samples and Jilin samples. Because no differences were found, samples from the different regions were grouped together in later analyses. Each computation was performed with at least 100 000 simulations. The multiple markers’ haplotype frequencies were estimated using the program EHPLUS.11 12

For TDT analysis, ETDT13 was used to perform the transmission disequilibrium test for single markers. Haplotype analysis was carried out by TRAMSMIT,14 version 2.5.4.

The standardised measure of LD for each pair of markers, denoted as D², was estimated with software 2LD15 for both case control and TDT analysis. All tests were two tailed and significance was accepted at p<0.05. To allow comparisons with previous studies, significance levels were not adjusted for multiple testing.

RESULTS

All the markers showed obvious differences in allele frequencies in our Chinese population, compared with Stefansson’s results.4 5 Allele frequencies and single marker analyses are shown in table 1. None of the five markers revealed significant allele association in case control samples nor transmission distortion in trios.

We calculated linkage disequilibrium (LD) for all marker pairs (expressed in D²). Strong LD was observed in all SNP pairs and microsatellite pairs (D²>0.6), whereas LD between SNPs and microsatellites was much weaker (D²<0.4). The results were consistent in both case control samples and family trios. They also accorded with the results of Tang et al, who suggested that there may be a recombination breakpoint between the two microsatellites 478B14-848, 420M9-1395 (D8S1810, 487-2, 478B14-848, 420M9-1395, DBS1810, 420M9-3663) reported by Tang et al. In their study, the haplotype specified as 20/4/–2/18 (487-2, 478B14-848, 420M9-1395, DBS1810, 420M9-3663) generated the positive association, and in our study the corresponding haplotype with alleles 4/–2 (478B14-848, 420M9-1395) played the same role. The relation between the current and previous studies is shown in table 4.

DISCUSSION

In the current study, we replicate the association between the 5’ end of NRG1 and schizophrenia in a Chinese population. Although the risk haplotype differs from that of Stefansson et al, which was virtually absent in the Chinese population, our results provide important evidence to support the existence of one or more functional variants within this region in both populations. As the given region covers the first promoter of NRG1 and the first exon of GGF2, the

Table 2  Case control analysis of the risk haplotype C/G/T/–/4/2

<table>
<thead>
<tr>
<th>Haplotype</th>
<th>Frequency* (cases)</th>
<th>Frequency* (controls)</th>
<th>p Value‡</th>
<th>Odds ratios‡</th>
</tr>
</thead>
<tbody>
<tr>
<td>5 markers</td>
<td>37.1</td>
<td>27.5</td>
<td>0.0057</td>
<td>1.56</td>
</tr>
<tr>
<td>haplotype</td>
<td>51.1</td>
<td>50.3</td>
<td>(0.00069)</td>
<td>(1.20–2.01)</td>
</tr>
<tr>
<td>3 SNPs</td>
<td></td>
<td>0.91</td>
<td>1.02</td>
<td></td>
</tr>
<tr>
<td>haplotype(CGT)</td>
<td></td>
<td>(0.90)</td>
<td>(0.81–1.28)</td>
<td></td>
</tr>
<tr>
<td>Microsatellites</td>
<td>57.2</td>
<td>48.9</td>
<td>0.0061</td>
<td>1.40</td>
</tr>
<tr>
<td>haplotype(4/–2)</td>
<td></td>
<td>(0.00078)</td>
<td>(1.13–1.72)</td>
<td></td>
</tr>
</tbody>
</table>

*Frequency %. †Values in parentheses are uncorrected p values from χ² distribution. ‡Values in parentheses are 95% confidence intervals.

Table 3  Transmission disequilibrium test of the risk haplotype C/G/T/–/4/2

<table>
<thead>
<tr>
<th>Haplotype</th>
<th>Observed result</th>
<th>Expected result</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>5 markers</td>
<td>249.19</td>
<td>225.25</td>
<td>0.0046</td>
</tr>
<tr>
<td>SNP8NRG243177</td>
<td>375.80</td>
<td>358.86</td>
<td>0.060</td>
</tr>
<tr>
<td>haplotype(CGT)</td>
<td>380.75</td>
<td>359.11</td>
<td>0.018</td>
</tr>
</tbody>
</table>
variant may affect the expression or splicing of the GGF2 isoform. These differences in allele frequencies, LD, and haplotype indicate that there may be more than one functional variant in the region, spanning several hundred kilobases. Alternatively, the same functional variant, which has not been identified, may be carried on a different haplotype in the Chinese population vs Caucasians.

The results of TDT in trios, when compared with the case control study, show a much weaker association. There are two main explanations that could account for our results. First, case control study has more statistical power than TDT to detect associations, because only parents who are heterozygous in given loci provide effective information in TDT. In the present study, heterozygosity of the microsatellites was limited, and the two haplotypes with the highest frequencies accounted for more than 70% of the total; thus the results of case control differed considerably from those of TDT. Secondly, it is possible that part of positive association is contributed by stratification, and TDT may avoid this problem effectively. However, the association detected by our case control study should not be considered a false positive, seeing that significant association was obtained by TDT as well.

The p values of risk haplotypes are also distorted by these factors. However, in our case control study, p values were assessed using the Monte Carlo approach instead of \chi^2 distribution, so that the p values have been corrected. Corrected and uncorrected p values are shown in table 2; the p values of TDT are not corrected.

In conclusion, our results confirm some previous studies and provide further support for the importance of NRG1 in predisposition to schizophrenia. However, the functional variant harbouring in this region is still unknown, and further detailed LD mapping in different populations is essential.

ACKNOWLEDGEMENTS
This work was supported by grants from the National 863 and 973 Projects, the National Natural Science Foundation of China, the Shanghai Municipal Commission for Science and Technology, and the Qiu Shi Science & Technologies Foundation.

**REFERENCES**


**BRAF mutations in cholangiocarcinoma**

Intrahepatic cholangiocarcinoma is the second most common primary hepatic malignant neoplasm, after hepatocellular carcinoma. Retrovirus-associated DNA sequences (RAS), controlled by RAS oncogenes, works at least in part through the mitogen activated protein kinase (MAPK) signal transduction cascade. Signalling through this cascade leads to activation of RAF kinase. Mammalian cells contain three RAF isoforms, A-RAF, B-RAF, and C-RAF. Mutations of BRAF have been found in around 15% of all human cancers, especially in malignant melanomas. Now researchers in Leipzig have studied the role of BRAF in liver tumours.

They looked for BRAF and KRAS mutations in 25 hepatocellular carcinomas and 69 cholangiocarcinomas by direct DNA sequencing after microdissection. MAPK pathway active intermediates were detected using immunohistochemistry. Activating BRAF missense mutations were found in 15 cholangiocarcinomas (22%) and in none of the hepatocellular carcinomas. The mutations were not found in non-neoplastic liver tissue. All BRAF mutations were within exons 11 and 15 and 11 of 15 mutations were in nucleotide 1796 leading to substitution of valine by glutamic acid at position 599.

KRAS mutations were found in 31 cholangiocarcinomas (45%) and none of the hepatocellular carcinomas. Twenty four mutations were of codon 12 and seven of codon 11. Ten were G→A transitions. Two KRAS mutations were found in non-neoplastic tissue. No correlations were observed between BRAF or KRAS mutations and histological or clinical features. Disruption of the Raf/MEK/ERK (MAPK) kinase pathway by either KRAS or BRAF mutation was detected in approximately 62% of all cholangiocarcinomas.

BRAF and KRAS mutations were common in cholangiocarcinoma but were not found in hepatocellular carcinoma.


---

**References**