Linkage mapping and phenotypic analysis of autosomal dominant Pallister-Hall syndrome

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Abstract

Pallister-Hall syndrome is a human developmental disorder that is inherited in an autosomal dominant pattern. The phenotypic features of the syndrome include hypothygalamic hamartoma, polydactyly, imperforate anus, laryngeal clefting, and other anomalies. Here we describe the clinical characterisation of a family with 22 affected members and the genetic mapping of the corresponding locus. Clinical, radiographic, and endoscopic evaluations showed that this disorder is a fully penetrant trait with variable expressivity and low morbidity. By analysing 60 subjects in two families using anonymous STRP markers, we have established linkage to 7p13 by two point analysis with D7S691 resulting in a lod score of 7.0 at θ=0, near the GLI3 locus. Deletions and translocations in GLI3 are associated with the Greig cephalopolysyndactyly syndrome. Although Greig cephalopolysyndactyly syndrome has some phenotypic overlap with Pallister-Hall syndrome, these two disorders are clinically distinct. The colocalisation of loci for these distinct phenotypes led us to analyse GLI3 for mutations in patients with Pallister-Hall syndrome. We have previously shown GLI3 mutations in two other small, moderately affected families with Pallister-Hall syndrome. The linkage data reported here suggest that these larger, mildly affected families may also have mutations in GLI3.

Methods

PEDIGREE ASCERTAINMENT AND PHENOTYPIC ANALYSIS OF FAMILY 1

The members of family 1 were examined by one of the authors (17 cases) or a referring physician (five cases). This research study was reviewed and approved by the NIH National Cancer Institute Institutional Review Board. Subjects were determined to be affected using published criteria for PHS. Outside clinical records and copies of imaging studies were obtained from personal physicians with the consent of the subjects. The subjects were evaluated by physical examination, magnetic resonance imaging of the CNS with a GE 1.5 Tesla scanner, flexible fibreoptic direct laryngoscopy, and limb radiographs.

Linkage analysis

Peripheral blood samples were obtained from 44 members of family 1 and 16 members of a previously reported family. Germic DNA was purified by affinity chromatography from RBC lysed whole blood (Qiagen Corp). Markers were selected from the CEPH database or Marshfield screening set 6.0 (Research Genetics, Inc) and amplified with α³²PdCTP incorporation and scored manually. Markers were...
selected from the set beginning with chromosomes that had been partially analyzed in previous linkage exclusion work. The linkage analysis calculations were done with FASTLINK (version 3.0P), which is a faster version of LINKAGE, using an autosomal dominant model with full penetrance, a mutant allele frequency of 0.0001, and gender independent recombination frequencies. These initial assumptions of penetrance, gene frequency, and gender independence were based on our experience with this disorder and were considered working hypotheses. Because these initial assumptions yielded positive results, alternative hypotheses were not considered. The multilocus calculations were done in parallel using FASTLINK on top of the TreadMarks distributed shared memory system running on an IBM SP2 parallel computer at the Division of Computer Research and Technology of NIH. Haplotype analysis was performed manually by minimising recombinants. YAC-based physical mapping was performed as previously described.

Results

CLINICAL ANALYSIS OF FAMILY 1

The proband (IV.8, fig 1) is a 21 month old child who was noted to have ptosis and polydactyly at the time of birth. This finding prompted an ophthalmological and neurological evaluation including a cranial MRI examination. The cranial MRI showed that she had a $2 \times 2.5 \times 3$ cm mass in the area of the hypothalamus that did not enhance with gadolinium (fig 2). The MRI finding was compatible with a hypothalamic hamartoma.

The family history showed that she had 21 relatives with polydactyly in an autosomal dominant pattern (fig 1). The pedigree includes nine obligate heterozygotes, all of whom have some manifestation of the disorder. The gender ratio of the affected members is six males to 16 females (however, note that family 2 has nine affected males and only four females). There is one instance of male to male transmission (from III.20 to IV.30).

Figure 1 The pedigrees of the families (1 and 2) used in the linkage analysis. This figure also shows the haplotype analysis for markers D7S521, D7S691, D7S678, D7S621, and D7S1818. The asterisk denotes an allele in an offspring of an affected parent homozygous for that allele. Note that the haplotypes in family 2 are arbitrarily set based on the offspring of subject IV.10.

Figure 2 Sagittal cranial magnetic resonance image of subject IV.8. The arrowhead indicates the mass of the hypothalamic region, diagnosed as a hamartoma by the MRI signal characteristics.
reported to have had polydactyly but no supporting documents are available.

The affected members of family 1 are in good general health and have normal intellectual development. One sib of the proband has learning disabilities but is not developmentally delayed. All members of family 1 have normal craniofacial features including head circumference and interpupillary distances. Four members of family 1 have had psychiatric diagnoses including depression and schizophrenia. However, one of those four is unaffected by PHS. III.17 was diagnosed with gelastic epilepsy at 7 years of age (gelastic epilepsy is a rare partial complex seizure disorder that is characterised by uncontrolled laughing in the ictal phase). She is now treated with gabapentin and has been free of seizures for several years. IV.18 was admitted to hospital for headache and depression at 15 years of age. An MRI scan showed a mass in the hypothalamus and she underwent a biopsy of that lesion. The histology of that tissue was reported as a hamartoma.

Twelve of the 22 affected persons in family 1 have undergone cranial MRI imaging either as a part of this study or for medical indications. Of those 12 persons, all but two had obvious masses of the tuber cinereum that were compatible with a hypothalamic hamartoma. These masses ranged from less than 1 x 1 x 1 to 2.5 x 3.0 x 3.0 cm in dimension (table 1). In all cases, the pituitary gland was present and normally formed. The masses were isointense to gray matter on all pulse sequences and did not enhance with gadolinium. Of the two persons who did not have an obvious hypothalamic mass, one (II.2) has a slight protrusion of the posterior hypothalamus that could be a normal variant. The second person (III.23) has no evidence of a hypothalamic mass. No scan showed evidence of increased intracranial pressure or mass effect. There was no clinical evidence of optic neuropathy, visual field cuts, or endocrine dysfunction. There is no clinical evidence of progression or expansion of the hamartomas, consistent with previous experience with this disorder.

Polydactyly was present in all 22 affected persons, which is expected since it was used as the primary feature for patient recruitment into the study (table 1). The polydactyly was markedly variable in family 1 and included postaxial polydactyly types A (fully formed supernumerary digit) (fig 3A) and B (digitus minimus) as well as central polydactyly (fig 3B). Most people had four limb polydactyly although one person had polydactyly limited to the hands (III.17). Most affected persons in family 1 had
Table 2  Two point linkage scores

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**Figure 4**  Foot radiograph of III.26 showing an apparent vestigial metatarsal bone lying between the third and fourth metatarsals.

**Discussion**

**CLINICAL CONSIDERATIONS**

The diagnosis of PHS in family 1 was made after the evaluation of IV.8, who presented with ptosis and polydactyly. According to available medical records and the medical history, the diagnosis of PHS was not considered previously in this family although two other people (III.17 and IV.18) had been diagnosed with polydactyly and hypothalamic hamartomas. The CNS manifestations of the disorder in family 1 are variable, but range in severity from moderate to asymptomatic. The presence of gelastic epilepsy in III.17 is probably the result of the hamartoma because of the rarity of this form of epilepsy and its frequent association with hypothalamic hamartomas. The rarity of symptoms referable to the hamartoma in family 1 and the lack of improvement in those with sporadic and familial hamartomas who have undergone extirpation of hamartomas for various symptoms suggest that operative intervention is rarely indicated.

A characteristic of the early case reports of PHS was the association of the hamartoma with endocrine dysfunction. This association ranged from panhypopituitarism to isolated precocious puberty. Of the 10 known or reported families with inherited PHS, only one case has been associated with endocrine dysfunction. The findings in family 1 are consistent with the association of endocrine dysfunction with sporadic, but not familial,

**LINKAGE ANALYSIS**

Initial linkage was noted to marker D7S672 with a lod score of 1.09 at θ=0.25. Two point linkage analyses with nearby markers showed close linkage to D7S691 with a lod score of 7.0 at θ=0 (table 2). The peak two point lod score was 8.4 at θ=0.025 with D7S678. Multipoint analyses of this region were performed using D7S21, D7S678, D7S691, and D7S621. These analyses gave a peak lod score of 13.5 near D7S691 on 7p13 (fig 5). Haplotype analysis using the Génethon map marker order showed that there were three double recombinants in these two families. We hypothesised that the genetic ordering of markers was incorrect and pursued physical mapping to attempt to resolve these double recombinants. YAC based physical mapping of this region showed a different order for these markers, shifting D7S678 in a centromeric direction. The Génethon order was tel-D7S678-D7S521-D7S691-cen, whereas the physical order placed D7S678 centromeric of D7S691. Analysis of the reordered haplotypes in the two families showed that this marker order eliminated double recombinants in the region (fig 1). The YAC based physical map of this region, including ESTs and genes (GLI3 and inhibin βA, INHBA) is shown in fig 6. Specifically, the deduced physical order is tel-D7S521-INHBA-D7S691-GLI3-D7S678-D7S621-D7S1818-cen (the latter two markers were ordered in the experiments but are not shown in fig 6). Thus, on genetic and physical grounds, INHBA and GLI3 could not be excluded as candidate genes for this disorder.

Flexible fibreoptic direct laryngoscopy was used to examine the larynx of 10 persons in family 1. Two of the 10 subjects had a bifid epiglottis and one person had a misshapen, angled epiglottis. One subject with bifid epiglottis also had a small posterior laryngeal cleft. These malformations were not associated with aspiration, recurrent pneumonia, or other voice or swallowing disorders in these patients.
PHS. None of the 22 affected persons in family 1 has documented endocrine dysfunction and there is no history of precocious puberty.

The finding of epiglottic clefts in family 1 is remarkable because it is an extraordinarily rare malformation. There are 16 case reports of bifid epiglottis tabulated in a recent review. A number of the older case reports of PHS have included descriptions of epiglottic and laryngeal clefts. This epiglottic malformation, together with the metacarpal or metatarsal polydactyly, and the hypothalamic hamartoma can be used to determine the embryological timing of the developmental disturbance in this disorder. The epiglottis, limb paddle, and hypothalamus all form during the seventh week of gestation. Thus the gene that causes this disorder must act during or before that time to alter these early developmental processes. The issue of whether sporadic and familial PHS are a single entity or are aetiologically heterogeneous has been debated elsewhere. However, clinical data cannot definitively address this hypothesis, which must await molecular analysis.

LINKAGE ANALYSIS

The mapping of the PHS phenotype to a delimited region within 7p13 raised the possibility that PHS and Greig cephalopolysyndactyly syndrome (GCPS) were allelic disorders. GCPS has been shown to be the result of translocations or deletions in the GLI3 gene that cause an abnormal phenotype by the mechanism of haploinsufficiency. Although PHS and GCPS are clearly distinct phenotypes, the similarity of the disorders is intriguing. Both have polysyndactyly, craniofacial features, and are inherited in an autosomal dominant pattern. Nevertheless there are major differences between the disorders. Central polysyndactyly has not been associated with GCPS. Preaxial duplication of the toes has been described in a single, atypical, sporadic case of PHS. Hypertelorism and frontal prominence, which are common in GCPS, are not reported in PHS. No cases of GCPS have been reported to have a hypothalamic hamartoma. There is significant variability in the polydactyly in both families analysed in this study. However, other than mild widening of the hallux phalanges on x ray, this variability does not include partial duplications of the thumbs or big toes. Therefore, given the findings of colocalisation and some phenotypic overlap of the two disorders, we concluded that PHS and GCPS were clinically distinct phenotypes and that GLI3 was a candidate gene for PHS on clinical and genetic grounds.

![Figure 5](image1.png)

**Figure 5.** Multipoint linkage results of two separate four point analyses of PHS and three STRP markers. The X axis of the graph is non-linear and shows five linkage scores in each interval. The lengths of the intervals are specified on the X axis. The two candidate genes, INHBA and GLI3, are shown in the intervals that they have been assigned by physical mapping: The notch in the results at 55.3 cM is the result of uninformative genotypes at D7S621.

![Figure 6](image2.png)

**Figure 6.** YAC based physical map. Shown here is a portion of the YAC contig map for the region of chromosome 7p15 described in the text. This map was constructed as part of a larger effort to derive a complete physical map of chromosome 7. Along the top are listed the STSs. For the Genethon genetic markers, the genetic map position (in cumulative cM from pter) are indicated above the marker name. Uncharacterised ESTs are designated by a $ sign. Marks that are not uniquely ordered on the physical map have an asterisk between them. Note that D7S251, D7S2469, D7S1423, D7S2454, and D7S2548 are not uniquely ordered on the Genethon map. Marker D7S678, which the Genethon map places telomeric (left) of D7S321, physically maps adjacent to D7S2428. This ordering of markers eliminates apparent double crossovers in HIL25 and IVS in family 1 and V1 in family 2 (see fig 1). Only a minimal set of YACs are depicted in the figure. A complete view of the contig map is available in the Genomes Division of Genbank (see http://www.ncbi.nlm.nih.gov/), while additional details about the chromosome 7 physical map are available at http://www.ncbi.nlm.nih.gov/DIR/GTB/CHR7. Relevant information about the YACs and STSs is available in Genbank or GDB.
On the basis of these data we have performed and previously reported mutation screening of GLI3 in PHS patients. We have found frameshift mutations in GLI3 in two small families (two and four affected, respectively) but no mutation has been identified in the families analysed in this linkage study. The determination of mutations in these other two families coupled with the linkage reported in the two large families in this paper strongly suggests that mutations in GLI3 are responsible for autosomal dominant PHS in the larger mildly affected families reported here. That GLI3 mutations also cause PHS in the severe sporadic cases remains to be determined. Taken together, the results showing that GCPS is caused by haploinsufficiency, that small families with moderate PHS have frameshift mutations, and that large mildly affected PHS families are tightly linked to GLI3 suggest that detailed genotype-phenotype correlations will provide important insights into human development and the function of GLI3 in normal development.

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