Neonatal spinal muscular atrophy with diaphragmatic paralysis is unlinked to 5q11.2-q13

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Abstract
Two sibs affected by the severe neonatal form of spinal muscular atrophy (SMA) with diaphragmatic paralysis are described. The two sibs were discordant for the haplotypes determined by DNA markers flanking the SMA locus. This supports non-linkage of SMA to chromosome 5 in this family and indicates that the uncommon SMA type I variant associated with early onset respiratory failure maps outside the 5q11.2-q13.3 region.

Spinal muscular atrophy (SMA) is a group of autosomal dominant, X linked, and autosomal recessive diseases with severe, intermediate, or mild phenotypes. The autosomal recessive forms (MIM 253300, MIM 253400, MIM 253550) are classified as type I, type II, and type III according to clinical manifestations and age of onset. The localisation of the SMA locus has allowed prenatal diagnosis in at risk families with at least one affected child. It has been estimated that only 5% of all SMA type I families are not linked to 5q. However, definite proof of this figure is still lacking and other authors suggest that a consistently higher proportion of SMA type I cases are unlinked to 5q. Cobben et al reported a family which suggested the existence of a second locus for autosomal recessive SMA type I. We report on two sibs, born to unaffected parents, with neonatal SMA type I associated with early onset respiratory distress related to diaphragmatic paralysis, in which DNA analysis suggested non-linkage to chromosome 5q.

Case reports

Case 1
A 3200 g female infant was delivered at term to a primiparous 28 year old woman. The parents were healthy and unrelated with no history of muscle disorders. At birth, the infant appeared apnoeic without response to stimuli and was immediately intubated. Cyanosis cleared promptly with bag ventilation with 2% oxygen but several attempts at extubation failed because of lack of respiratory effort, and, therefore, mechanical ventilation needed to be undertaken. There was no evidence of skeletal muscle weakness and deep tendon reflexes were normal. Chest radiological examination showed marked raising of both hemidiaphragms with absence of diaphragmatic motion on fluoroscopy (fig 1).

At 2 months of age, the baby was still unable to ventilate and clinical evidence of progressive muscular weakness was present. At 3 months of age, muscle wasting, lingual fasciculations, hypotonia, absent motility, and areflexia were clearly indicative of a severe neuromuscular impairment. Limb muscle weakness was predominantly distal with paralysis of the extensors of the hands and feet. The arms were abducted and internally rotated at the shoulders assuming a "jug handle" position. There was a gradual decrease in leg movements with bilateral development of pes equinus. Respiratory failure was associated with a narrowed thorax, pectus excavatum, and flaring of the lower ribs. Feeding and swallowing were impossible.

Muscular enzymes were only slightly raised (CK 230 IU/l and LDH 462 IU/l). An ele-
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Figure 2 Intercostal muscle: comparison of cases 1 and 2 (HE) shows the histological change in time of neurogenic atrophy. Increase of anisocytosis of muscle fibres and fat infiltration are evident.

Tromyogram showed no fibrillation at rest but small groups of fibres were discharging during movement and this was considered consistent with the diagnosis of diffuse denervation. Nerve stimulation of IPS and EPS showed a reduction of the registered MAP.

A biopsy of the left quadriceps was performed and studied using histological, histochemical, and histoenzymatic techniques. The presence of clusters of large rounded fibres and scattered atrophic fibres is consistent with SMA.

The patient died at 7 months of age after several episodes of pneumonia. Necropsy showed widespread neurogenic atrophy of skeletal muscle (fig 2). The diaphragm was thin and membranes showed severe reduction of discernible muscle fibres. Macroscopic appearance of the pons, cerebral bulb, and spinal cord was markedly atrophic (data not shown). There was a severe depletion of the number of anterior horn cells associated with degenerative changes, neuronophagia, and gliosis (fig 3).

The patient was originally diagnosed as having SMA type I and the mother requested for prenatal diagnosis in the second pregnancy. The parents were informed of the limitations of prenatal diagnosis based on a linkage analysis in nuclear SMA families.\(^{9,12}\) Notwithstanding this, the mother requested first trimester monitoring of the fetal genotype and continued the pregnancy.

**CASE 2**

A 3300 g male infant was delivered after an uneventful pregnancy at 41 weeks of gestation. At one minute, the Apgar score was 2 and the baby was intubated. At five minutes, the Apgar score was 7 but there was no respiratory effort and manual ventilation was needed. On initial examination, bilateral fixed flexion of the elbow and wrist joints with ulnar deviation of the hands was noted. The infant was hypotonic, with absent tendon reflexes and a weak response to stimuli. A chest x ray showed an unusually high diaphragm bilaterally (data not shown). Clinical and radiological features were highly suggestive of the same disease documented in the dead first sib. Therefore it was decided that respiratory support by mechanical ventilation was not indicated, and the infant died one hour after birth.

A muscle biopsy of the left quadriceps showed groups of atrophic and scattered normal fibres (data not shown).

Necropsy showed a markedly raised thin and velar diaphragm. Histological examination of different skeletal muscles showed large groups of atrophic fibres (fig 2). This pattern was most severe in the diaphragm in which a few normal sized fibres were visible. The brain and cerebral bulb appeared normal. Histological examination of the thalamus, cerebral bulb, and medulla oblongata showed severe neuronal loss, especially in the anterior horns. A few of the remaining neurones showed marked enlargement and degenerative changes associated with occasional neuronophagia (data not shown).

**DNA STUDIES**

DNA from a frozen biopsy of case 1 was available and used for molecular analysis, according to Lo Cicero et al.\(^{9}\) Genomic DNA from a chorionic villus sample (CVS) was obtained at 12 weeks during the mother’s second pregnancy and indirect prenatal diagnosis was performed using a set of microsatellite markers flanking the SMA locus on 5q.\(^{9}\)

The results of the DNA analysis are shown in fig 4. Complete informativity was obtained using the markers D5S125, D5S435, D5S557, D5S39, and D5S127. This analysis predicted that the fetus had a wild type genotype, having
inherited the parental haplotypes unlinked to the chromosome 5q SMA locus. A recombination event was also detected between D5S112 and D5S39, which was not considered relevant because it was outside the SMA locus (fig 4, II.2) Microsatellite alleles were re-evaluated in lymphocyte and fibroblast DNA of the newborn (case 2), and complete concordance with the CVS results was obtained. This excludes misdiagnosis owing to laboratory contamination, sampling errors, or PCR artefacts. Paternity testing using three DNA polymorphisms (D1S80, APOB, HLADQ2) was performed and showed no evidence of non-paternity.15

Discussion

Severe neonatal respiratory failure, associated with radiological evidence of bilateral diaphragmatic weakness and evagination, is not a typical feature of SMA type I. On the other hand, the diagnostic criteria of SMA, reported by the International SMA Consortium,16 do not exclude SMA patients with diaphragmatic paralysis. In the first patient, clinical manifestations of Werdnig-Hoffmann disease became apparent at 2 months of age, but some limb deformities suggestive of a neuromuscular disorder were present in the second sib at birth. The diagnosis of SMA was confirmed in both instances by muscle biopsy and postmortem examination that showed (1) neurogenic muscle atrophy; (2) decrease in the motor neurons of the anterior horns of the spinal cord; (3) degenerative changes and neuronophagia; and (4) proliferation of astrocytes and microglia. This unique variant of SMA, associated with early respiratory muscle involvement and weakness and atrophy of the distal musculature, has previously been observed.15-19 Moreover, it has been suggested that other infants with early onset fatal respiratory disease of unknown aetiology may also have this unusual variant of SMA.20

The mapping of the SMA locus to chromosome 5 has shown genetic homogeneity between “classical” SMA phenotypes including the severe form or Werdnig-Hoffmann disease, the mild form or Kugelberg-Welander disease, and the intermediate type III form. This has ruled out non-allelic heterogeneity to explain the interfamilial and intrafamilial clinical heterogeneity.8,11,22 In the absence of significant genetic heterogeneity within the autosomal recessive SMAs, the DNA markers in the chromosomal region 5q12-13 are usually used for prenatal and presymptomatic diagnosis in informative families.9,11,23,24 A prenatal risk calculation which includes genetic heterogeneity can be applied and is acceptable in the majority of cases.9,12 However, the recent observation of SMA type I unlinked to 5q and the occurrence of de novo deletions of the 5q13 region indicate the need for caution in interpreting the results provided by linkage studies.8,11,25,26

The family reported in the present study is unlinked to chromosome 5q markers. This indicates that the rare SMA variant associated with paralysis of the diaphragm maps outside the 5q11.2-13.3 region. It also implies that this clinical form is genetically different from “classical” SMA types. The two sibs of the present family had inherited the four parental chromosomes 5, excluding recombination events. Furthermore, clinical features were distinct from those expected to occur in other autosomal recessive diseases associated with evagination of the diaphragm or in other similar progressive motor neurone diseases.18

This finding has relevant implications for genetic counselling and prenatal diagnosis in SMA families.

We thank Dr E Bertini for the useful discussion. This work was supported by Telethon Italy (Grant no 296), ASM (Milan), and MURST (40% and 60%).

References


Figure 4 Pedigree and DNA results in the family. The numbers under the pedigree symbols refer to alleles of the DNA markers DSS125, DSS435, DSS557, DSS39, and DSS127. The SMA locus on 5q is located between DSS435 and DSS557.
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