Linkage analysis in blepharophimosis-ptosis syndrome confirms localisation to 3q21-24

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Abstract

Blepharophimosis-ptosis is an autosomal dominant disorder in which previous chromosome rearrangements have suggested a putative gene location on the long arm of chromosome 3. This paper confirms the location at 3q21-24 with linkage studies in two large families. A lod score of 3.2 was found with D3S1237.

The blepharophimosis-ptosis syndrome (BPES) was first described in 1841 by Fredrich August von Ammon. It consists of blepharophimosis, ptosis, epicanthus inversus and some extraocular features such as learning difficulties and female infertility. The term blepharophimosis describes the reduction in the horizontal diameter of the palpebral fissure. This is associated with dysplasia of the eyelid owing to fibrosis and impairment of the levator palpebrae superioris muscle. There may also be amblyopia and blockage of the lacrimal puncta. Surgical correction is available to improve the cosmetic appearance of the eyes.

Mild psychomotor retardation has been observed in some patients with one report giving a mean IQ score of 86. However most affected subjects have no significant learning difficulties. Infertility has been reported frequently in females but not in affected males. It has been suggested that there may be two subtypes depending on whether female infertility is present or not.

The condition is an autosomal dominant trait. In 1991 Fukushima et al. published a case report of blepharophimosis sequence associated with a de novo balanced translocation 46,XY,t(3;4)(q23;p15.2) and suggested that the locus might be assigned to 3q23. Following this Jewett et al. reported a further case associated with an interstitial deletion at 3q22. The aim of our study was to confirm the putative location by linkage studies using two large families with BPES.

Materials and methods

The families were contacted and a full history and examination carried out on each member of the families. Inner canthal, outer canthal, and interpupillary distances were measured.

Figure 1 Pedigrees of family A and family B (twins IV-6 and IV-7 are dizygous).
using a metal rule and appropriate photographs were taken. A review of previous hospital records was carried out where there had been hospital investigations. Up to 20 ml of blood in potassium EDTA was taken from affected and unaffected members of the families for DNA analysis. Loose buccal cells obtained by a sucrose mouthwash were used for DNA extraction from those who did not consent to venepuncture.

DNA was extracted from whole blood by the method described by Bertelson et al. Specific target sequences in the region of 3q2 were amplified using the polymerase chain reaction. In view of the previous chromosome rearrangements the CA repeat primers D3S196, D3S1237, D3S1238, and D3S1309 were selected and obtained from Research Genetics (table). Polymerase chain reaction (PCR) was performed as reported previously. Amplified products were separated using 10% PAGE, and visualised with silver stain. Lod scores were generated using the Mlink section of the LINKAGE package version 5.03. Pedigrees were entered into the LINKAGE program using the Megabase interface. The incidence of BPES was taken to be one in 10 000 live births and complete penetrance was assumed. Allele frequencies were obtained from the genome database.

Results

There were nine affected subjects alive in family A and four affected members in family B (fig 1). They all had the classical appearance of blepharophimosis-ptosis as illustrated in figs 2, 3, and 4.

In family A two affected females, II-6 and III-4, had passed through their reproductive years and had reduced fertility. In III-4 menarche was delayed at 17 years. Initially she had a normal menstrual pattern until 25 years, but after this the frequency of the menstrual cycles decreased. Extensive investigations including pelvic ultrasound, hormone studies, and pituitary function tests were undertaken. The ultrasound scans showed she had one ovary of reduced size. She was treated with Clomid (clomiphene) and Pergonal (menotrophin) without success. IV-4 had menarche at 12 years and at her present age of 16 years has regular periods and has not yet been investigated further. The remaining three affected females are prepubertal.

One affected member of family A (IV-1) is a 9 year old boy with severe learning difficulties. He developed fits at the age of 10 months. CT brain scans have shown dysgenesis of the corpus callosum and enlarged lateral ventricles. He also has irregular teeth with a midline incisor (fig 4).

The results of the linkage analyses are shown in the table. The maximum lod score was 3·21 for D3S1237. Since no recombinations were observed, the recombination fraction 0 = 0. The segregation of the haplotypes is shown in fig 3.
**Discussion**

This syndrome gives a very characteristic facial appearance, but it is not the only syndrome with blepharophimosis or prosis. Blepharophimosis has been reported in the Marden-Walker syndrome and the Ohdo syndrome. In these syndromes there are many associated features and developmental delay is more significant than in BPES. The degree of developmental delay that occurs in BPES is usually relatively mild and not associated with structural brain abnormalities. The patient reported here (family A, IV:1) is unusual in having severe learning difficulties and a midline developmental defect manifest in dysgenesis of the corpus callosum and a single incisor. Clayton-Smith et al described a boy with blepharophimosis, agenesis of the corpus callosum, and mental retardation, but the features in this case more closely resemble the Ohdo syndrome than BPES. Thus the association between BPES and midline cerebral abnormalities has not been observed previously, and suggests this may be the extreme end of the phenotypic spectrum.

There is no suggestion of reduced fertility in males, although no specific clinical investigation has been conducted on this. However, in females reduced fertility is well recognised and is a significant part of the clinical management of the disorder. In the families reported, most of the females have been prepubertal, but III:4 in family A has been extensively investigated and appears to have primary ovarian failure. This finding is similar to published reports where investigations have been carried out. Smith et al reported two sisters with BPES in whom one sister had ovaries with a large number of primordial follicles on ovarian biopsy but was resistant to hormonal stimulation, and the other affected sister had no follicular development in the ovary and had a premature menopause. It appears the mechanism of reduced fertility in BPES may be similar to the ovarian dysgenesis seen in Turner's syndrome, but is the result of an autosomal gene rather than X chromosome loss.

Many single gene syndromes have first been mapped through their associations with chromosomal rearrangements. There is considerable evidence from chromosomal rearrangements that the gene for BPES is localised to the long arm of chromosome 3 at 3q23. However, it is interesting that Warburg et al reported a patient with many similar features to BPES in whom there was a deletion of 7q34. What is particularly interesting about this patient is that there was a single incisor, suggesting a partial holoprosencephaly sequence. In our family A patient IV:1 also has a single incisor and agenesis of the corpus callosum. We therefore feel that partial holoprosencephaly may be part of the BPES. It has already been suggested that BPES could be a contiguous gene syndrome with several adjacent genes deleted. It is difficult to see how the abnormality in the embryonic development of the eyelids and the ovarian failure could be related embryologically, and the possibility of a contiguous gene syndrome is very attractive. However, Vasalli et al have reported a single gene in the mouse, which affects both eyelid development and ovarian function. The gene produces the betaB subunit for activin and inhibin. It is located on mouse chromosome 9 and, as the human homologous region is 2cen-q13 rather than 3q23, it is not a candidate gene for BPES.

The linkage data show that the lod scores were above 3 for primers D3S1237 and D3S1238. They are located at 3q21-q22 and 3q22-q24 respectively. No recombinations were observed between any of the markers and the disorder. It is surprising that no recombinations were seen between D3S1237 and D3S1238 considering the distance between
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them is in the order of 32 cM. The lack of recombination makes fine mapping more difficult unless further families with this rare disorder are studied. From the linkage data presented the gene locus for BPES is likely to be 3q22-q23, which correlates well with the putative localisation from chromosome re-arrangements. A very recent paper by Small et al has also shown linkage of blepharophimosis-ptosis syndrome to 3q. These pedigrees used the same recombination with the markers used and the locus is placed proximal to D3S1309. The authors suggest that this places the BPES gene locus at 3q22-23 but D3S1309 maps to 3q21 and the most likely location for the BPES gene is given as near the RHO/ACPP markers at the proximal side of band 3q13. Although there are some discrepancies between the cytogenetic and genetic maps, this does not tie in with the most recent suggested cytogenetic location of the BPES gene. Further analysis of families with this syndrome, especially with more proximal markers, will improve the localisation.

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