Refining the genetic location of the gene for X linked hydrocephalus within Xq28

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Abstract
The most common inherited form of hydrocephalus, X linked hydrocephalus (HSAS), is characterised by mental retardation, adducted thumbs, and spastic paraplegia. Genetic analysis has mapped the locus for HSAS to subchromosomal band Xq28 within a region of approximately 2 megabases of DNA. In order to refine the location of the disease gene we have conducted genetic linkage analysis with Xq28 marker loci in four additional HSAS families. A lod score of 4.26 with polymorphic marker DXS52 (St14) confirms the linkage of HSAS to Xq28. Identification of a recombination event between the HSAS gene and Xq28 loci F8C and DXS605 (2-19) reduces the size of the interval likely to contain the disease locus to about 1.5 megabases, the distance between DXS605 and DXS52. The locus for neural adhesion molecule, LICAM, maps within this interval and therefore represents a candidate gene for HSAS.

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X linked recessive hydrocephalus is the most common genetic form of congenital hydrocephalus, occurring in approximately 1/30 000 male births (McKusick 30700). The primary diagnostic features of mental retardation and enlarged cerebral ventricles are often accompanied by adducted thumbs and spastic paraplegia. The frequent observation of aqueductal stenosis led to the acronym HSAS (X linked hydrocephalus with stenosis of the aqueduct of Sylvius) although this malformation is now considered to be secondary to the basic defect. A variety of other cerebral malformations, including absence of the corpus callosum and defects in the septum pelucidum, have occasionally been reported in this condition. Genetic analysis has placed the locus for HSAS in subchromosomal band Xq28 close to polymorphic locus St14 (DXS52) and the gene for coagulation factor VIII (F8C).

HSAS exhibits marked clinical variation within and between families and its features overlap with those of another Xq28 linked disorder, MASA syndrome (mental retardation, aphasia, spastic paraplegia, and adducted thumbs). Since genetic analysis has also placed the gene for MASA syndrome close to DXS52 and F8C it has been suggested that the two disorders are caused by different mutations at the same locus. Indeed hydrocephalus has been observed in at least one MASA syndrome family.

Attempts to refine the location of the HSAS locus within Xq28 have relied on the analysis of rare examples of meiotic recombination between the disease locus and Xq28 markers. A recent summary of these and additional data, incorporating information from 13 HSAS families, concluded that the relevant gene lies between DXS52 and F8C, an interval of about 2 megabases (Mb) by comparison with a physical map of the region.

In order to identify further recombinants we have conducted genetic linkage analysis with Xq28 markers in four additional HSAS pedigrees.

Materials and methods

FAMILIES
Families with at least two affected males in more than one sibship or generation were selected for analysis. No male to male transmission was noted and female carriers were asymptomatic.

Family H1
Subject II.7 was born with a normal head circumference but developed massive hydrocephalus within the first few weeks of life. He died of pneumonia aged 9 months. II.5 died aged 47 and despite a normal head circumference was found to have grossly enlarged cerebral ventricles at necropsy. He was mentally retarded (IQ 45) and suffered from epilepsy and spastic diplegia, requiring crutches from 6 years of age. II.8 is now 49 years old, mentally retarded, has spastic diplegia, and walks with a shuffling gait. His head circumference is within the normal range (55-5 cm) but a brain CT scan showed grossly enlarged ventricles. III.2 had a normal head circumference at birth but developed progressive hydrocephalus requiring shunting. He is now 25 with mild spastic diplegia, mental retardation, and has suffered from epilepsy since the age of 22. DNA was not available for II.7 and only paraffin embedded tissue was available for II.5.

Family H2
Subjects II.9, III.6, and IV.1 were stillborn with hydrocephalus but no further details are available. Fetal scanning of IV.2 at 28 weeks' gestation showed marked dilatation of the lateral and third, but not fourth, cerebral ventricles with the cortical mantle reduced to 3 mm in

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Refining oligonucleotide appropriate using DXS605 F8C VNTR4, DNA merase this crude extract plus of tissue to according DNA of both DNA and paraffin embedded tissue sections were the only source of DNA for IV.6.

Family H3
Patient IV.6 is 6 years old and was born by caesarean section at 35 weeks after prenatal diagnosis of hydrocephalus. He had a grossly enlarged head and a subsequent CT scan showed dilatation of the lateral and third ventricles with a thin cerebral mantle. A ventricular-peritoneal (VP) shunt was fitted. He is mentally retarded, and has adducted thumbs and spastic diplegia. Patient IV.8 died at 2 weeks of age and necropsy showed hydrocephalus.

Family H4
Patient II.1 is now 23 years old, mentally retarded, has adducted thumbs, and walks with a shuffling gait. No scans have been performed. II.4 was born with hydrocephalus, is now 13 years of age, and has received 23 operations since birth for VP shunt maintenance. His head circumference is normal. He is mentally retarded with an unsteady gait, left sided weakness, and adducted thumbs. Patient III.1 was diagnosed prenatally with hydrocephalus and delivered by caesarean section. His head was enlarged and a CT scan showed dilatation of the lateral and third ventricles only. Choroid plexus coagulation and VP shunt insertion were performed. He had adducted thumbs and left sided weakness and died aged 10 months. Necropsy showed collapse of both cerebral hemispheres.

DNA POLYMORPHISMS
Blood samples were obtained with informed consent from available family members and DNA was extracted using routine methods. Where paraffin embedded tissue was the source of DNA, extraction was performed according to the following procedure: 250 mg of tissue (in 10 μm sections) was suspended in 250 μl extraction buffer (75 mmol/l NaCl, 25 mmol/l EDTA). This was subjected to three cycles of heating to 65°C for five minutes plus vortexing for one minute. After collection by centrifugation the sample was incubated for three days at 50°C in the presence of proteinase K (300 μg/ml) and SDS (0.5%). Aliquots of this crude extract were used directly for polymerase chain reactions. Families were typed for DNA polymorphisms DXS52 (St14, VNTR4, F8C (BclI restriction digest4), DXS605 (2–19, EcoRI14), and DXS707 (2–55, Msp116) using PCR and digestion with the appropriate restriction enzyme. The sequences of oligonucleotide primers for DXS605 and DXS707 were a personal communication from Daniella Toniole.

LINKAGE ANALYSIS
Data obtained by polymorphic typing were analysed using the LIPE17 and LINKMAP18 computer programmes for two point and multipoint analyses respectively. Confidence intervals were obtained by taking values of the recombination fraction corresponding to a lod score one unit less than the maximum.

Results
Linkage analysis was conducted on the HSAS families shown in fig 1 using a sample collection of 36 subjects that included 10 affected males and 11 obligate carriers. Two point lod scores for X linked hydrocephalus versus Xq28 polymorphic marker loci are given in the table. No recombinants were observed between DXS52 and the disease locus and a lod score of 4.26 supports the Xq28 location of the HSAS gene. A single recombinant between HSAS and marker loci F8C and DXS605 was observed in family H4 and the haplotypes of this family are shown in fig 2. The relative order of marker loci in Xq28 is derived from Poustka et al. Since the disease segregates with DXS52 in family H4 and not with DXS605 and F8C these data imply that the HSAS gene is located proximal to DXS605. For multipoint analysis we used the marker order DXS52, DXS605, F8C with recombination fractions of 0.05 and 0.001 respectively taken from published linkage data on the assumption that the genetic location of DXS605 is the same as G6PD. DXS707 was not included in this analysis as its location is uncertain. The LINKMAP analysis gave a maximum location score of 24.2 with HSAS coincident with DXS52. This position was favoured by odds of 108:1 compared to a location between DXS605 and F8C, and by odds of 80:1 compared to a location distal to F8C.

Discussion
We have conducted genetic linkage analysis on four new X linked hydrocephalus families. The affected males in our study vary markedly in their clinical presentation both within and between families, an observation that has been made for other HSAS pedigrees.2 The presence of adducted thumbs in addition to spastic paraplegia and mental retardation in two families highlights the overlap of HSAS with MASA syndrome. A lod score of 4.26 obtained using DXS52 indicates close linkage of the disease locus in these families with Xq28 markers. Lack of linkage to Xq28 has been shown for only one out of 13 families analysed by Willems et al13 and absence of recombination with DXS52 implies that the families in our study exhibit the Xq28 linked form of the disorder. There was no evidence of heterogeneity for families H1 to H4 with DXS52 yielding lod scores of 1.25, 1.51, 0.6, and 0.9 respectively.
A single recombination event indicates that the HSAS gene lies proximal to polymorphic loci DXS605 and F8C. Polymorphic locus DXS605 is situated 20 kb downstream of the gene for glucose-6-phosphate dehydrogenase (G6PD), approximately 1.5 Mb distal to DXS52 and 500 kb proximal to F8C. A study by Willems et al. indicated that the HSAS gene lies in the interval between F8C and DXS52, a distance of about 2 Mb. Our results are consistent with this location but narrow down the size of the relevant region to the distance between DXS605 and DXS52, approximately 1.5 Mb. Thus, a direct search for candidate genes should focus on this reduced interval.

The most likely candidate identified to date is the gene for neural cell adhesion molecule, L1CAM. This highly conserved, cell surface glycoprotein is involved in neural cell migration and development of the neuromuscular junction. Since the L1CAM locus is physically located between DXS605 and DXS52, our data indicate that the L1CAM gene should be examined directly in cases of X linked hydrocephalus.

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