A nonsense mutation in the tyrosinase gene of Afghan patients with tyrosinase negative (type IA) oculocutaneous albinism

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Abstract
We detected a nonsense mutation in the tyrosinase gene of two Afghan sibs with classical tyrosinase negative (type IA) oculocutaneous albinism. The mutation, a single base substitution at codon 178, creates an amber termination codon that truncates the 529 amino acid tyrosinase polypeptide at this position. The patients' parents are first cousins, and the patients are therefore homoallelic for this mutation.

Oculocutaneous albinism (OCA) is a heterogeneous group of severe, autosomal recessive inherited disorders of pigmentation. In OCA the amount of melanin is reduced or absent in pigment cells of the skin, hair follicle, and eye. Hypopigmentation of the retina is associated with decreased visual acuity, photophobia, and varying degrees of nystagmus. In patients with classical (type IA) OCA, absent melanin biosynthesis results from absent activity of melanocyte tyrosinase (monophenol, L dopa:oxygen oxidoreductase, EC 1.14.18.1). Tyrosinase is a copper containing enzyme that catalyses the first two steps of the melanin biosynthetic pathway: the hydroxylation of tyrosine to dihydroxyphenylalanine (dopa) and the subsequent oxidation of dopa to dopa-quinone.1 2

Several human tyrosinase cDNAs have been cloned3-5 and the 529 amino acid sequence has been deduced. The first 18 N-terminal amino acids constitute a leader peptide which is removed to yield the mature 58 kd highly glycosylated tyrosinase enzyme.6 The deduced amino acid sequence contains five potential asparagine linked glycosylation signals, two potential copper binding sites, and a C-terminal potential transmembrane region.7 The human tyrosinase gene is located on chromosome segment 11q14–q21.8 and consists of five exons spanning more than 50 kb of DNA.9

Seven different mutations of the tyrosinase gene have been described in patients with type IA OCA. These include a frameshift mutation (codon 310)10 and a missense mutation (codon 77 Arg→Gln)10 in Japanese patients, a frameshift mutation (codon 501)11 and three different missense mutations (codon 81 Pro→Leu,12 codon 373 Thr→Lys, and codon 383 Asp→Asn)13 in Caucasian patients, and one missence mutation (codon 89 Cys→Arg)14 in an American Black. Here we describe an eighth tyrosinase gene mutation associated with type IA OCA, an amber nonsense mutation at codon 178 (Trp→Ter), in two sibs of Afghan ethnic origin. The patients' parents are first cousins and both patients are homoallelic for the codon 178 nonsense mutation.

Materials and methods
DESCRIPTION OF PATIENTS
The two patients, a brother (patient 1) and sister (patient 2), are the offspring of a consanguineous marriage of first cousins of Afghan ethnic origin. The patients' parents, a brother, and three sisters are darkly pigmented. Although hairbulb tyrosinase activity was not determined, both affected children exhibit classical features of type IA oculocutaneous albinism, completely lacking apparent melanin pigment in their hair, skin, and eyes. Their irides are pale blue and transilluminate completely, and their retinas are unpigmented. Their visual acuity is greatly reduced, with foveal hypoplasia, constant nystagmus, and photophobia. Patient 1 also has trisomy 21.

Enzymatic assay of freshly epilated anagen hairbulbs from patient 1 showed no detectable tyrosinase activity.
POLYMERASE CHAIN REACTION (PCR) AND DNA SEQUENCING
Genomic DNA was isolated from peripheral blood leucocytes of both patients and tyrosinase gene fragments corresponding to the five exons plus flanking sequences of the male sib (patient 1) were PCR amplified from 0.1 μg DNA in 100 μl volumes of 10 mmol/l Tris-HCl, pH 8.3, 50 mmol/l KCl, 1.5 mmol/l MgCl_2, 200 μmol/l of each dNTP, 100 μg/ml gelatin, 100 pmol of each primer (table), and 2.5 U Taq DNA polymerase (Perkin-Elmer Cetus, Norwalk, CT) using an automated thermal cycler (Coy Laboratory Products Inc, Ann Arbor, MI). Each of 40 cycles consisted of 30 seconds at 94°C, one minute at 50°C, and two minutes at 72°C. Amplification products were gel purified, subcloned into bacteriophage vectors M13mp18 or mp19, and their nucleotide sequences were determined.

RESTRICTION ENZYME CLEAVAGE ANALYSIS
Tyrosinase gene exon 1 DNA fragments of both patients and also of eight normal persons of northern European Caucasian origin were amplified by PCR using oligonucleotide primers P11 and P12 (table). PCR amplification products were then digested with XbaI and analysed by polyacrylamide gel electrophoresis for the presence of the codon 178 nonsense mutation. The exon 1 PCR products of the two patients were also digested with MboI to analyse a polymorphism at codon 192.

Results
IDENTIFICATION OF THE TYROSINASE GENE CODON 178 MUTATION
To define the molecular basis of the type IA (tyrosinase negative) OCA in the two sibs of Afghan origin, we analysed their tyrosinase genes by PCR amplification, molecular cloning, and subsequent DNA sequencing. Each of the five tyrosinase exons plus adjacent non-coding sequences was amplified from genomic DNA of patient 1 using oligonucleotide primers (table) derived from the DNA sequence of the normal human tyrosinase gene. As shown in fig 1, nucleotide sequence analysis of the PCR products showed only a single difference from normal, a base substitution at codon 178: TGG (Trp) → TAG (Ter). This amber codon would result in termination of translation of the corresponding tyrosinase mRNA at this position.

The coding region of normal human tyrosinase genes exhibits two common non-pathological polymorphisms. Codon 192 can encode either tyrosine (TAT) or serine (TCT) and codon 402 can encode either arginine (CGA) or glutamine (CAA). The codon 192 polymorphism is also an RFLP for the restriction enzyme MboI (TAT: no MboI site, TCT: MboI site). Therefore, 338 bp PCR products corresponding to part of exon 1 were amplified from genomic DNA of both patients using primer pair P11/P12, digested with MboI, and were analysed by polyacrylamide gel electrophoresis for the presence of the polymorphic MboI site (data not shown). Both probands were found to be homozygous for TCT (Ser) at this codon. The polymorphism at codon 402 is not an RFLP; therefore, we sequenced six independent exon 4 M13 subclones from patient 1 and found that he is also homozygous for codon 402 CGA (Arg).

Oligonucleotide primers used for amplification of tyrosinase gene exons by PCR.

<table>
<thead>
<tr>
<th>Primer</th>
<th>Sequence</th>
<th>Position*</th>
<th>Sequence amplified</th>
</tr>
</thead>
<tbody>
<tr>
<td>P1</td>
<td>5'-GCTCTTTAAGTGAGATATC-3'</td>
<td>-40→-21†</td>
<td>Exon 1 plus adjacent regions and promoter</td>
</tr>
<tr>
<td>P2</td>
<td>5'-TTATACCTGCGCTGAAGAG-3'</td>
<td>1543-1562c</td>
<td></td>
</tr>
<tr>
<td>P3</td>
<td>5'-CAGAGGGAGCTTCAACACG-3'</td>
<td>1644-1663</td>
<td>Exon 2 plus adjacent regions</td>
</tr>
<tr>
<td>P4</td>
<td>5'-AAGCTGCAAGAAATCTGTAAATGC-3'</td>
<td>2049-2068c</td>
<td></td>
</tr>
<tr>
<td>P5</td>
<td>5'-GAGTCTCATATCAGAAGATGGA-3'</td>
<td>2153-2172</td>
<td>Exon 3 plus adjacent regions</td>
</tr>
<tr>
<td>P6</td>
<td>5'-TTAATAATCCATGAGCAGCAGT-3'</td>
<td>2470-2498c</td>
<td></td>
</tr>
<tr>
<td>P7</td>
<td>5'-TTCATATATGCTTATTTTT-3'</td>
<td>2514-2533</td>
<td>Exon 4 plus adjacent regions</td>
</tr>
<tr>
<td>P8</td>
<td>5'-TAAGATTGGTGTATTCACT-3'</td>
<td>2842-2861c</td>
<td></td>
</tr>
<tr>
<td>P9</td>
<td>5'-CCTCAAGAGCTGGAAGAGGG-3'</td>
<td>2963-2982</td>
<td>Exon 5 plus adjacent regions</td>
</tr>
<tr>
<td>P10</td>
<td>5'-GAGTGCATATCATGTTAGATT-3'</td>
<td>3661-3680c</td>
<td></td>
</tr>
<tr>
<td>P11</td>
<td>5'-GCAAGTTTGCTTTTGGGGA-3'</td>
<td>979-998</td>
<td>Fragment of exon 1</td>
</tr>
<tr>
<td>P12</td>
<td>5'-CAAGAAGAGTCTATGCCAGG-3'</td>
<td>1297-1316c</td>
<td></td>
</tr>
</tbody>
</table>

*Nucleotide positions refer to sequence of Giebel et al.†This primer was derived from the sequence of Kikuchi et al; its position is enumerated relative to the sequence of Giebel et al. c indicates that primer sequence is complementary to that in fig 2 of ref 8.
This enabled us to confirm the presence of this mutation in the patients' genomic DNA. A 338 bp fragment of the tyrosinase gene exon 1 was PCR amplified from genomic DNA of the two sibs with OCA and from eight unrelated, normal, Caucasian subjects using primer pair P11/P12. The fragments were then digested with XbaI and analysed by polyacrylamide gel electrophoresis (fig 2). The 338 bp exon 1 PCR products of all eight normal subjects contained no XbaI site and thus were not cleaved by XbaI. However, the 338 bp fragments of both OCA patients were completely cleaved by XbaI to 223 bp and 115 bp fragments. This is consistent with their both being homoallelic for the codon 178 TGG→TAG nonsense substitution.

Discussion

Tyrosinase negative (type IA) oculocutaneous albinism, characterised by absent melanocyte tyrosinase activity, results from tyrosinase gene mutations that abolish tyrosinase enzymatic activity.9-14 We have identified a new tyrosinase gene mutation in two Afghan sibs with type IA OCA. Both patients are homoallelic for a single base substitution (TGG→TAG) within codon 178, resulting in the change of a tryptophan codon to a translational termination codon. The resultant truncated 177 amino acid tyrosinase polypeptide would lack both of the putative copper binding sites and the putative transmembrane segment,3 and therefore would have no tyrosinase enzymatic activity and might even be subject to rapid
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proteolytic degradation. The codon 178 substitution creates a new XbaI restriction site, permitting simple identification of this substitution by either allele specific oligonucleotide hybridisation or XbaI cleavage analysis.

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