Carrier detection and early diagnosis of Wilson’s disease by restriction fragment length polymorphism analysis

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SUMMARY Wilson’s disease, a rare autosomal recessive disorder, has been recently mapped to the long arm of chromosome 13 (q14.1). In this study, we carried out linkage analysis between three chromosome 13 DNA markers, D13S1, D13S10, D13S2, the locus for the red cell enzyme esterase D (ESD), and the Wilson’s disease locus (WND) in 17 Wilson’s disease families of Italian descent, mostly from Sardinia. We confirmed a tight linkage [\(\theta=0.00, Z(\theta)=4.07\)] between the WND and ESD loci, and provided suggestive evidence for linkage [\(\theta=0.00, Z(\theta)=1.70\)] of the WND locus with D13S10. Multipoint linkage analysis indicated the following order: centromere-D13S1-D13S10-WND-ESD-D13S2. RFLP analysis at these two loci in our families allowed us either to define the carrier status (50%) or to exclude the homozygous state (25%) in the great majority of unaffected sibs.

Wilson’s disease, or hepatolenticular degeneration, is a rare, autosomal recessive disorder characterised by impaired biliary excretion and defective incorporation of copper into ceruloplasmin. This results in hepatic and extra-hepatic copper accumulation, low plasma concentration, and increased urinary excretion.1 Clinical presentation occurs in the first or second decade of life and is characterised either by liver disease or extrapyramidal syndrome. The variability of the clinical picture and age at presentation suggests genetic heterogeneity.2 The molecular basis of the disease is not known. Chelation therapy with penicillamine is highly effective, especially when initiated before irreversible organ damage has occurred.

Preclinical diagnosis, based on low ceruloplasmin levels and increased urinary copper excretion, is now completely reliable and should be confirmed by invasive methods such as liver biopsy. Carrier detection, by evaluation of ceruloplasmin levels and incorporation of 64Cu into ceruloplasmin, does not give a definite result and prenatal testing is not available. Progress in gene mapping of Wilson’s disease should improve the diagnostic capabilities.

A tight linkage between the WND and ESD loci on chromosome 13 has recently been detected.3 Further studies confirmed this linkage and also detected a tight linkage between WND and the polymorphic region D13S10 which is detected by the single copy probe 7D2.6 The incidence of Wilson’s disease in the Sardinian population is around 1 to 3/10^4 livebirths,7 an order of magnitude higher than in other Caucasian and non-Caucasian populations (1/10^5).1 In this study, we carried out linkage analysis...
between the WND locus and several chromosome 13 polymorphic loci in 17 Italian families, mostly from Sardinia. Our results confirmed linkage between the WND locus and the ESD and D13S10 loci and showed the feasibility of early diagnosis and carrier detection by DNA analysis.

Patients and methods

Patients

This study included 17 Italian families with Wilson’s disease, 14 from Sardinia and three from the mainland. Nine of these families had multiple cases (fig 1). Out of 26 patients investigated, 19 were younger and seven older than 14 years. In the group of young patients, 16 had chronic liver disease and three were asymptomatic. In the group of older patients, two had the extrapyramidal syndrome, two had chronic liver disease, and three were asymptomatic.

Diagnosis was based on reduced serum ceruloplasmin and copper levels, high urinary copper excretion, and copper accumulation in the liver, evaluated by biopsy. The pertinent laboratory data on the 26 patients evaluated are presented in table 1.

Serum ceruloplasmin levels, investigated in 34 parents, were normal in 31 and reduced in three.

DNA analysis

Leucocyte DNA extraction and restriction endonuclease analysis were carried out as previously described.8

<table>
<thead>
<tr>
<th>TABLE 1 Pertinent laboratory data of 26 Wilson’s disease patients.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Serum ceruloplasmin (mg/dl)</td>
</tr>
<tr>
<td>Serum copper (mg/dl)</td>
</tr>
<tr>
<td>Copper excretion (µg/24 h)</td>
</tr>
<tr>
<td>Liver copper (µg/g dry weight)</td>
</tr>
</tbody>
</table>

Numbers indicate mean [1 SD] followed by the range of normal values.

FIG 1 Pedigrees of the 17 Wilson’s disease families investigated.
Three anonymous probes, p7F12, p7D2, and p9D11, which detect RFLPs at the chromosome 13 loci D13S1, D13S10, and D13S2 respectively, were used to hybridise Southern blots of DNA samples from members of the Wilson's disease families. In addition, an ESD cDNA probe was used to detect RFLPs at the ESD locus. The ESD isoenzymes were also determined according to Hopkinson et al.9

**LINKAGE ANALYSIS**

Pairwise linkage analysis between the WND locus and each of the chromosome 13 polymorphic loci in 17 informative families was performed by the lod scores method, using the MLINK program from the linkage package,10 assuming a recessive model with complete penetrance and a population frequency of the WND gene of 0.002.7 Genotypes from different polymorphisms detected by single copy probes were combined into chromosomal haplotypes. Separate lod scores for sex and one lod unit confidence intervals for the estimated recombination fractions were calculated as recommended at HGM8.11

Multipoint linkage analysis to determine the most likely order of the four marker loci and the relative position of the Wilson's disease locus among the marker loci was performed using the ILINK and LINKMAP programs from the LINKAGE package.

**TABLE 2 RFLPs at four loci on chromosome 13 in 17 Italian families.**

<table>
<thead>
<tr>
<th>Locus</th>
<th>Restriction enzyme</th>
<th>No of subjects</th>
<th>Heterozygosity (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>D13S1</td>
<td>MspI, TaqI, BclI</td>
<td>34</td>
<td>58.8</td>
</tr>
<tr>
<td>D13S10</td>
<td>TaqI, DraI</td>
<td>26</td>
<td>42.3</td>
</tr>
<tr>
<td>ESD†</td>
<td>ApaI</td>
<td>34</td>
<td>67.5</td>
</tr>
<tr>
<td>D13S2</td>
<td>MspI, TaqI</td>
<td>32</td>
<td>62.5</td>
</tr>
</tbody>
</table>

*ESD isoenzymes were also investigated.

**TABLE 3 Linkage analysis between WND and four marker loci on chromosome 13.**

<table>
<thead>
<tr>
<th>Loc1</th>
<th>Loc2</th>
<th>Sex</th>
<th>Z(δ)</th>
<th>δ</th>
<th>CI</th>
</tr>
</thead>
<tbody>
<tr>
<td>WND</td>
<td>ESD</td>
<td>M</td>
<td>4.7</td>
<td>0.00</td>
<td>0.00-0.084</td>
</tr>
<tr>
<td>WND</td>
<td>D13S1</td>
<td></td>
<td>2.35</td>
<td>0.049</td>
<td>0.003-0.284</td>
</tr>
<tr>
<td>WND</td>
<td>D13S1</td>
<td></td>
<td>1.85</td>
<td>0.00</td>
<td>0.0-0.224</td>
</tr>
<tr>
<td>WND</td>
<td>D13S2</td>
<td></td>
<td>1.46</td>
<td>0.09</td>
<td>0.002-0.299</td>
</tr>
</tbody>
</table>

**Results**

**RFLP ANALYSIS**

We studied the heterozygosity of the parents of patients with Wilson's disease at four chromosome 13 polymorphic loci, D13S1, D13S10, D13S2, and ESD, by RFLP analysis (table 2). More than one RFLP was analysed for each locus. In addition, ESD, serum isoenzyme polymorphism was also investigated. The highest percentage (67.5%) of heterozygosity was detected at the ESD locus and the lowest (42%) at the D13S10 locus.

**LINKAGE ANALYSIS**

Lod scores [Z(δ)] at various recombination fractions (δ) between WND and the four marker loci are reported in table 3. Significant linkage [Z(δ)=4.07] at an estimated recombination frequency δ=0.00 was found for ESD, and suggestive evidence was found for D13S10 [Z(δ)=1.85, δ=0.00]. Separate analysis for sex did not show any significant differences in recombination between males and females.

Multipoint linkage analysis of the four markers, performed to establish the most likely relative position of the four loci on the long arm of chromosome 13, indicated that the most likely order is: centromere-D13S1-D13S10-ESD-D13S2. Since our data on the recombination frequencies among marker loci, based on a small number of families, were subject to large sampling errors, we used the published genetic map6,12 and the corresponding estimates of the recombination frequencies between each pair of loci for the purpose of establishing the relative position of the WND locus. Results of four overlapping four point analyses performed, excluding one marker at a time, indicated that the WND locus is likely to be located distal to D13S10, but it might be either proximal or distal to ESD. The two most favoured orders are D13S1-D13S10-WND-ESD.
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D13S2 and D13S1-D13S10-ESD-WND-D13S2, with a relative likelihood of the first order over the second of approximately 4:1, at a recombination frequency of 0.07 between WND and D13S10 and of 0.03 between WND and ESD.

Carrier detection of Wilson's disease by DNA analysis

All members, affected and unaffected, of the 17 Wilson's disease families were studied at the four polymorphic loci. Because of the high recombination between D13S1 and D13S2 and the WND locus (θ=0.049 and 0.09 respectively, table 3) only ESD and D13S10, which showed the lowest recombination fraction with WND in our study, were used for carrier detection.

The RFLPs at these polymorphic loci were combined to obtain a chromosomal haplotype. Haplotype analysis may avoid misdiagnosis resulting from recombination between WND and one of the other two loci, an example of which is illustrated in fig 2.

The linkage phase was established in both parents in three families (completely informative families), in one parent only in 11 (partially informative families), and in neither in three (uninformative families) (table 4).

In eight families with multiple cases, the affected sibs shared identical chromosome 13 haplotypes which were never detected in unaffected sibs.

Discussion

In this study, which included a large number of

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**TABLE 4** Informativeness of the Wilson's disease families at the ESD and D13S10 polymorphic loci.

<table>
<thead>
<tr>
<th>Families (17)</th>
<th>With 1 case (9)</th>
<th>With 2 cases (8)</th>
</tr>
</thead>
<tbody>
<tr>
<td>One parent informative</td>
<td>4</td>
<td>7</td>
</tr>
<tr>
<td>Both parents informative</td>
<td>2</td>
<td>1</td>
</tr>
<tr>
<td>None</td>
<td>3</td>
<td></td>
</tr>
</tbody>
</table>

**TABLE 5** Carrier detection by RFLP analysis in Wilson's disease families.

| No of families | 17 |
| No of possible carriers investigated | 36 |
| Carrier status excluded | 2/36 |
| Carrier status identified | 16/36 |
| Undefined | 18/36 |

By haplotype analysis, we investigated 36 possible carriers in the 17 Wilson's disease families. We were able to identify 16 carriers and to exclude the carrier status in two. Carrier detection was not possible in the remaining 18. However, in 11 of them haplotype analysis allowed the exclusion of the homozygous state (table 5).

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**FIG 2** Pedigree of a Wilson's disease family with two affected members. The different bars indicate the chromosomal haplotype obtained analysing four chromosome 13 loci. The different alleles at each locus investigated are indicated with numbers only in the parents and in the member indicated by the arrow, where a double recombination occurred between ESD and D13S10-D13S1 in the proximal region and D13S2 in the distal region.
families with Wilson’s disease from a single population, we confirmed a tight linkage (θ=0.00) with a lod score of 4.07 between the gene for Wilson’s disease and the ESD locus. This study also provides suggestive evidence for linkage [θ=0.00, Z(θ)=1.85] between the WND and the D13S10 loci. Haplotype sharing in affected sib pairs in multicase families further supports the linkage of WND to ESD and D13S10.

Multipoint linkage analysis between several chromosome 13 markers and the WND locus indicates that the most likely order is as follows: centromere-D13S1-D13S10-WND-ESD-D13S2. This order corresponds to the less favoured one proposed by Bowcock et al. (D13S10-WND-ESD-D13S2 or D13S10-ESD-WND-D13S2) on the basis of their analysis of five families with Wilson’s disease from different populations.

The extreme variation in the clinical picture and the marked difference in age at presentation suggest that Wilson’s disease may be heterogeneous at the molecular level. This heterogeneity could be allelic or non-allelic. Non-allelic heterogeneity should be excluded before applying, at the clinical level, polymorphism analysis based on linkage between markers and the disease locus detected in some families. All the 17 families investigated in this study were independently ascertained and in all of them the WND locus was linked to chromosome 13 markers. From these data it seems reasonable to assume that, at least in the Sardinian population, Wilson’s disease is largely caused by mutation(s) at the WND locus located on chromosome 13.

The p7D2 and ESD gene probes are informative in approximately 35-3% and 64-7% of matings and, given the linkage to the WND locus, would provide the tools for preclinical diagnosis, heterozygote detection, and prenatal diagnosis.

RFLP analysis at these two loci in our families allowed us either to define the genotype (50%) or to exclude the homozygous state (25%) in the large majority (75%) of unaffected sibs. However, recombination between WND and either ESD or D13S10 may occur and result in misdiagnosis. Although in our samples the maximum likelihood estimate of the recombination distance between WND and the two linked RFLP markers is 0.00, the 90% confidence interval for the recombination distance should be used for the conservative estimate of the probability of an at risk fetus being affected with Wilson’s disease. On the assumption that ESD and D13S10 flank WND, by using both polymorphic loci considered as haplotypes in informative families, the risk of a sib sharing the same haplotypes as the proband would be adjusted from 25% to 98-2%, while the probability of being heterozygous would be 1.7%. If the order of loci is D13S10-ESD-WND false negative and false positive diagnoses would be in the order of 11.3% and 5.8% respectively.

Prenatal diagnosis of Wilson’s disease will be greatly facilitated when new probes linked to the WND locus are available and the order of the probes along chromosome 13 is defined.

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References


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