The calcitonin-CGRP gene in the infantile hypercalcaemia/Williams-Beuren syndrome

G A Hitman*, L Garde*, W Daoud†, G J A I Snodgrass†, and R D Cohen*

From the Medical Unit* and Department of Paediatrics†, The London Hospital Medical College, London E1 1BB.

**Summary**

We have investigated 13 families, each of which have one member with infantile hypercalcaemia/Williams-Beuren syndrome (IHWBS), for either a germ cell mutation of, or an association with, the calcitonin-CGRP gene. Restriction fragment mapping studies of the calcitonin-CGRP gene using five restriction enzymes (TaqI, BglII, PvuII, PsiI, and SacI) and region specific probes failed to show any abnormalities of this gene complex. No association of IHWBS with polymorphism of the calcitonin-CGRP/parathormone locus was found. Therefore, although the aetiology of IHWBS may be caused by a new dominant mutation, there is no evidence to implicate major rearrangements of the calcitonin-CGRP and parathormone genes.

The infantile hypercalcaemia/Williams-Beuren syndrome (IHWBS) is a congenital disorder characterised by mental retardation, stenoses of the aorta (especially supravalvular aortic stenosis) and other large vessels, a distinctive facial appearance, and transient but often severe hypercalcaemia in infancy.\(^1\)\(^-\)\(^6\) On reviewing 117 patients with this disorder 29% were found to be hypertensive; although a proportion of these could be explained by renal artery stenoses there is no evidence that this is the only mechanism involved.\(^4\)

The disorder occurs in approximately one in every 20,000 live births and its incidence has remained constant in the UK at about 18 per year for the last 20 years.\(^4\) In British families this disease appears to be sporadic with no other members of the family being affected; the one exception is a pair of identical twins (family 5, fig 1) who are concordant for the disease.\(^4\) Occasional familial cases have been published: one additional pair of monozygotic twins\(^7\) and a pair of second cousins,\(^8\) and Cortada et al\(^9\) described the syndrome in a mother and her dizygotic twin daughters. In this family of Cortada et al the diagnosis has been challenged by Burn, who thought it more resembled Noonan's syndrome.\(^5\)

Supravalvular aortic stenosis (SVAS) occurs both sporadically and within families. The sporadic form can be subdivided into cases of IHWBS or SVAS alone (with normal facies and intelligence). Familial cases follow an autosomal dominant mode of inheritance and usually have normal facial appearance and normal intelligence. Interestingly, the combination of SVAS, pulmonary stenosis, and hypercalcaemia has been reported in a 28 week human fetus investigated for severe non-immune hydrops; the parents were both first cousins suggesting (if this was IHWBS) an autosomal recessive mode of inheritance.\(^10\) These observations give rise to a number of hypotheses to explain the sporadic and possible familial cases. The first is that a germ cell mutation has occurred in one of the gametes or in the fertilised ovum. This might explain the wide range of abnormalities seen in this condition together with its sporadic nature and would be consistent with the identical twins being concordant for the disease. Alternatively, although less likely, infantile hypercalcaemia segregates as a rare autosomal recessive disease which is almost invariably lethal in utero; however, exceptionally, there is a live birth (in less than 1% cases). The latter hypothesis would then explain the apparent 'sporadic' nature of the disease with a few familial cases being described. Against this hypothesis there is no evidence of an increased miscarriage rate in women who have had offspring with IHWBS, although it should be borne in mind that early fetal losses would not be detected.

We have investigated these hypotheses in 13 families with IHWBS by studying the calcitonin-CGRP gene complex on the short arm of chromosome 11.\(^11\)\(^12\) This complex was chosen for several

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FIG 1 Pedigrees studied together with results of the calcitonin-CGRP parathormone gene haplotyping. Letters under symbols=haplotypes which are detailed in table 2. Parental haplotypes AB and CD were derived by combinations of a calcitonin-CGRP TaqI RFLP, a TaqI parathormone gene RFLP, and a parathormone gene PstI RFLP. The parental haplotypes AB and CD are not the same for each family but are abbreviations, the key to which is given in table 2. In family 8 there is recombination between the parathormone PstI and TaqI RFLPs and hence the full data are shown with the pedigree.
TABLE 1 Gene mapping of the calcitonin-CGRP gene complex in IHWBS.

<table>
<thead>
<tr>
<th>Enzyme</th>
<th>Fragment size</th>
<th>Probe</th>
</tr>
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<tbody>
<tr>
<td></td>
<td>A</td>
<td>B</td>
</tr>
<tr>
<td>TaqI</td>
<td>2-9</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>2-6</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>8-06</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>2-5</td>
<td>-</td>
</tr>
<tr>
<td>BglII</td>
<td>2-1</td>
<td>+</td>
</tr>
<tr>
<td></td>
<td>4-1</td>
<td>-</td>
</tr>
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<td></td>
<td>0-45</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>0-6</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>9-4</td>
<td>-</td>
</tr>
<tr>
<td>PvuII</td>
<td>3-4</td>
<td>+</td>
</tr>
<tr>
<td></td>
<td>2-4</td>
<td>+</td>
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<tr>
<td></td>
<td>2-0</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>3-4</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>5-4</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>5-4</td>
<td>-</td>
</tr>
</tbody>
</table>

The resulting DNA fragments were separated by size on a 1% agarose gel and transferred to Gene Screen Plus membranes using an alkaline Southern blot technique as previously described. The membranes were hybridised to 32P labelled gene probes, washed down to a stringency of 0-2 SSC, and gene sequences detected by autoradiography. Fragments were sized by comparison with HindIII digested lambda phage and an internal control.

**Methods**

Thirteen families were identified by the Infantile Hypercalcaemia Foundation all of which had at least one member with IHWBS. All patients with IHWBS had had a documented episode of hypercalcaemia in infancy and met the strict criteria for diagnosis defined by Martin et al. Family pedigrees are shown in fig 1.

DNA was extracted from 10 to 20 ml blood samples (anticoagulated with EDTA) from probands and all available family members, and separately digested with TaqI, BglII, PvuII, PstI, and SacI. The calcitonin-CGRP gene was synthesised using a gene probe technique as previously described. The resulting DNA fragments were separated by size on a 1% agarose gel and transferred to Gene Screen Plus membranes using an alkaline Southern blot technique as previously described. The membranes were hybridised to 32P labelled gene probes, washed down to a stringency of 0-2 SSC, and gene sequences detected by autoradiography. Fragments were sized by comparison with HindIII digested lambda phage and an internal control.

**Gene Probes**

Gene probes used in the study were all kindly donated by Professor R Craig (Courtauld Institute

[Diagram of gene probes used in study]

**Fig 2** The calcitonin-calcitonin gene related peptide (CGRP) complex and gene probes used.

<table>
<thead>
<tr>
<th>Probe</th>
<th>Plasmid code</th>
<th>Insert cutting sites</th>
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<tbody>
<tr>
<td>A</td>
<td>pGEMCAL22</td>
<td>EcoRI, BamHI</td>
</tr>
<tr>
<td>B</td>
<td>pGEMCAL12</td>
<td>PstI</td>
</tr>
<tr>
<td>C</td>
<td>pGEMCAL12</td>
<td>PvuII</td>
</tr>
<tr>
<td>D</td>
<td>pGEMCAL12</td>
<td>PvuII, SacI</td>
</tr>
<tr>
<td>E</td>
<td>pGEMCAL12</td>
<td>PvuII</td>
</tr>
<tr>
<td>F</td>
<td>pGEMCAL12</td>
<td>PvuII</td>
</tr>
</tbody>
</table>

Shaded regions refer to exons. Exons 1 to 3 and 4 are expressed in tissues producing calcitonin and exons 1 to 3 and 5 in those synthesising CGRP. Exon 6 codes for a 3' untranslated region including the poly-A tail.
of Biochemistry, Middlesex Hospital Medical School, London) and those used in the gene mapping study are detailed in table 1 and fig 2. For the construction of calcitonin-CGRP/parathormone gene haplotypes a PstI insert of PhT-B58 (probe F, fig 2) and cDNA parathormone gene probe were used.

Results

Results from the mapping studies are presented in table 1. Restriction sites using the six probes and five enzymes encompassed approximately 13-5 kb in and around the calcitonin-CGRP gene. No differences were observed between the IHWBS children and their families in the size of DNA fragments for this region.

Calcitonin-CGRP/parathyroid gene haplotypes are shown in the families in fig 1 and table 2. The TaqI calcitonin-CGRP polymorphism consisted of two allelic fragments sized 6-5 or 8-0 kb, the parathormone TaqI polymorphism was sized 2-5 and 2-4 kb, and the parathormone PstI polymorphism was sized 2-8 and 2-2 kb.

In three informative families the affected sib was identical to a normal sib for the calcitonin-CGRP/parathormone locus (for example, in family 7 sibs 3 and 4 were both AD, in family 11 sibs 8 and 9 were BC, and in family 13 sibs 3 and 5 were AC). These families therefore provide direct evidence against IHWBS being a recessive disorder at this locus.

Discussion

Restriction mapping studies exclude a germ cell mutation leading to deletions or insertions of over 50 nucleotides affecting the calcitonin-CGRP gene. Restriction mapping would not detect point mutation unless the mutation occurred within the enzyme recognition site. Since the enzymes used recognise four to six base pair sequences and 27 sites were analysed, this analysis cannot exclude a point mutation in the vast majority of sequences within this gene. An alternative to a germ cell mutation would be gene damage during early embryogenesis, in which case analysis of leucocyte DNA would not necessarily show any abnormalities. Using probes E and F a constant band for each enzyme, sized 2-5 kb (TaqI), 5-7 kb (PstI), and 4-2 kb (SalI) was identified which corresponds to a second CGRP gene. The latter has high homology with CGRP and is encoded by a gene separate from the calcitonin-CGRP gene complex. No differences in size of this second CGRP gene was found between the IHWBS children and their sibs.

Family studies provide evidence against this being an autosomal recessive condition with a defect at the calcitonin-CGRP gene complex. Recently, an analysis of the calcitonin-CGRP gene was carried out in a family with autosomal dominant supravalvar aortic stenosis and the authors concluded that this gene was not linked to 'Williams' syndrome'. However, family members with SVAS were of normal intelligence and had normal facies, and therefore their study only rules out linkage of this locus with familial SVAS and not IHWBS.

Subjects with certain defined chromosomal abnormalities on the long arm of chromosome 15, the long arm of chromosome 4, and a translocation of the long arm of chromosomes 9 and 17 have been described as having some features in common with
IHWBS. This might be another starting point to examine loci associated with this syndrome.

Finally, perhaps the collagen genes should be studied given the wide range of connective tissue disorders in this syndrome, together with large vessel disorders similar to those of Marfan's syndrome, although this would not explain the hypercalcaemia or mental retardation. Future studies on the calcitonin-CGRP gene perhaps should include sequencing this gene in DNA obtained from tissues which produce CGRP or calcitonin or both, or long range gene mapping (by pulsed field gel electrophoresis) to search for deletions which might affect gene activity.

This work was supported by grants from the Infantile Hypercalcaemia Foundation and Action Research. We are grateful to Sir George and Lady Cooper for help in identifying and contacting the families, to the families and children for their cooperation, and to Professor R Craig, Middlesex Hospital, for helpful discussions and the provision of gene probes. Genetic advice was kindly given by Dr R M Winter, Kennedy Galton Centre, Clinical Research Centre, Harrow.

References


Correspondence to Dr G A Hitman. Medical Unit, The London Hospital Medical College, Whitechapel, London E1 1BB.