Meiotic recombination in the β globin gene cluster causing an error in prenatal diagnosis of β thalassaemia

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SUMMARY In the course of a prenatal diagnosis for β thalassaemia by linkage analysis of restriction fragment length polymorphisms, a homozygous β thalassaemia fetus was misdiagnosed as β thalassaemia trait.

Extensive studies of the polymorphic sites within the β globin gene cluster in all the members of the family resulted in the conclusion that the paternal chromosome 11 of the newborn was different from that expected. Paternity was confirmed by HLA typing and blood group studies.

The analysis of another polymorphic locus on chromosome 11 within the family was in agreement with the possibility of a crossing over between the two paternal chromosomes in a region 5' to the β gene, previously indicated to contain a 'hot spot' area for recombination.

This report underlines the risk of performing prenatal diagnosis using restriction polymorphisms 5' to the β gene.

First trimester prenatal diagnosis is accomplished in a variety of hereditary disorders by linkage analysis using restriction fragment length polymorphisms (RFLPs) as markers of the chromosome carrying the abnormal gene.1-3 This indirect method is convenient in diseases heterogeneous at the molecular level, since it does not require direct identification of the molecular lesions and has been successfully applied to a wide range of diseases.3-6

β thalassaemia shows remarkable molecular heterogeneity: in Italy at least eight defects have been described7 and the majority of patients are compound heterozygotes for two different mutations.8 To circumvent the problem of simultaneous identification of two different thalassaemic defects in the fetus, a plan of prenatal diagnosis was organised in northern Italy based upon indirect assay using polymorphic restriction sites in the β globin cluster.9,10

This procedure always carries some degree of risk of recombination between the mutated gene and the polymorphisms analysed. Considering the distance between the polymorphic sites used and the β gene, the theoretical risk should be approximately 1:2000.11 Nevertheless, previous population studies on the association of polymorphic sites in this region suggested that the recombination rate in a 9 kb region 5' to the β gene could be greater than expected.7,12,13 Furthermore, at least two examples of meiotic recombination have been described up to now.14,15

In this paper we report an error which occurred during prenatal diagnosis of β thalassaemia by RFLP linkage analysis: a homozygous β thalassaemic child was born who had been diagnosed as β thalassaemia trait in the first trimester of pregnancy. The results obtained by studying the inheritance of several RFLPs in chromosome 11 within the family suggest that meiotic recombination in the father is the most likely explanation of this event.

Material and methods

DNA was prepared from peripheral blood of an Italian couple at risk for β thalassaemia, who

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requested prenatal diagnosis in the seventh week of gestation, and of their child with Cooley’s anaemia (fig 1).

DNA (10 μg) was digested with restriction endonucleases HindIII, HincII, AvaiI, and BamHI. DNA fragments were separated on 0.8% agarose gels and transferred onto nitrocellulose filters. The filters were hybridised to genomic probes corresponding to the globin genes. Eight polymorphic sites in the β like globin cluster\(^12\) were used to assess the feasibility of prenatal diagnosis (fig 2).

Fetal DNA was prepared from chorionic villi\(^17\) obtained by the aspiration technique in the ninth week of gestation. Both HindIII γ and HincII ψβ sites, 5′ to the β gene, were analysed in fetal DNA.

Other polymorphic sites studied were the TaqI 5′δ, the PstI at the δ gene level, and the HindIII 3′β\(^18\) in the β globin gene cluster and the polymorphic Ha-ras locus on chromosome 11 3′ to the β globin gene (fig 2); the latter was analysed by the AvaII restriction enzyme.\(^19\)

Paternity was assessed by traditional blood grouping and HLA typing.

Haemoglobin isoelectric focusing\(^20\) and globin chain synthesis\(^21\) were performed on the cord blood.

**Results**

The feasibility of prenatal diagnosis was 100% with the use of either HindIII γ or HincII ψβ polymorphic sites: the analysis of both these polymorphisms led to a diagnosis of a heterozygous β thalassaemia fetus (fig 1).

At birth, no haemoglobin A was detectable on isoelectric focusing of the cord blood, and globin chain synthesis on the peripheral blood of the newborn revealed no β chain production. Moreover, within three months II.2 developed a clear picture of Cooley’s anaemia, requiring blood transfusions.

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**FIG 1** Family pedigree. Informative polymorphic sites used for prenatal diagnosis (HindIII γ and HincII ψβ) are indicated. + indicates the presence and – the absence of the polymorphism.

**FIG 2** Map of the β globin gene cluster and of the Ha-ras locus on chromosome 11. Arrows indicate the polymorphic restriction sites examined. 1 = HincII ε, 2 = HindIII γγ, 3 = HindIII Aγ, 4 = HincII ψβ, 5 = HincII 3′ψβ, 6 = AvaiI ψβ, 7 = TaqI 5′δ, 8 = PstI δ, 9 = AvaII β, 10 = HindIII 3′β, 11 = AvaII 3′β. The sites 1–6, 9, and 11 are usually studied to assess the feasibility of prenatal diagnosis of β thalassaemia. Below: results of polymorphism analysis in all the members of the family (see fig 1). + = presence and – = absence of the polymorphism. Numbers indicate different sized Ha-ras AvaII alleles: 1 = 1.5 kb, 4 = 2.1 kb, 12 = 3.1 kb.
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To rule out technical errors, blood was again obtained from all the members of the family and the newborn; previous results were confirmed.

Non-paternity was taken into account and investigated by HLA typing and blood group studies: the segregation of alleles at several polymorphic loci (HLA-A, B, C, DR, ABO, Rh, Mn) was in agreement with correct paternity.

The pattern of polymorphic sites in the β globin cluster showed that II.2 had inherited the maternal β thalassaemia chromosome, as expected, and a paternal chromosome different from that inherited by the similarly affected sister (II.1) (fig 2).

Interestingly, the HindIIIγ and HincIIψβ polymorphisms in the paternal chromosome of II.2 correspond to those present in the 5' region of the 'normal' chromosome of I.1. Since II.2 has no normal β genes, this event could have occurred because of a meiotic recombination between the two paternal chromosomes.

To confirm this possibility, the pattern of inheritance of a polymorphic locus 3' to the β gene (the Ha-ras proto-oncogene) was studied. Different Ha-ras alleles are observed in normal populations, related to tandemly repeated sequences of variable length 3' to the coding sequences of the Ha-ras locus.19 This was also the case in our family, since the parental chromosomes could be distinguished by this polymorphism (fig 2). This study showed that II.1 and II.2 have inherited the same maternal and paternal Ha-ras alleles.

In order to define the boundaries of the hypothesised crossing over, other polymorphisms in the β globin region were examined. However, this did not provide any additional information, since the father was homozygous for the presence of the Taql 5'δ and the absence of the Pstδδ and the HindIII 3'β sites (fig 2).

Discussion

In the family studied, two sisters, both homozygous for β thalassaemia, have unequivocally different genetic markers in the region of the paternal chromosome 11, 5' to the β thalassaemia gene. As non-paternity can be confidently excluded, there are a number of different possibilities that could theoretically explain this fact.

The first is a new β thalassaemia mutation in the 'normal' chromosome of the father. We have no direct proof that the β thalassaemia mutation inherited from the father is indeed the same in II.1 and II.2, so the hypothesis of a new β thalassaemia mutation in the 'normal' paternal chromosome cannot be ruled out. However, this possibility is unlikely, since new mutations in β thalassaemia are extremely rare and up to now only one case has been demonstrated at the molecular level.22

A more realistic explanation is a meiotic recombination, either a gene conversion or a crossing over between the two paternal chromosomes II.

A single gene conversion event does not explain the simultaneous variation at the HindIII γ and HincII ψβ sites, since they are at least 6 kb apart. On the other hand, a conversion of the normal gene into a β thalassaemia one is ruled out by the Ha-ras polymorphism inheritance within the family. As shown in fig 2 the same Ha-ras alleles were found in I.1 and II.2, suggesting that both have inherited the same paternal thalassaemic chromosome. This finding is in agreement with the hypothesis of a crossing over in the region of the paternal chromosome 5' to the β gene. Since the other polymorphisms examined between the ψβ and β gene were not informative, the boundaries of the hypothesised crossover remain wide (fig 3): the event could have occurred at any site in the region between the ψβ gene and the β thalassaemia mutation, joining the 5' region of the originally 'normal' chromosome to the β thalassaemic chromosome.

We could not establish whether crossing over took place in II.2 or in II.1, since no other member of the paternal family was available to assess the usual polymorphism pattern linked to the β thalassaemic mutation.

At least two other recent cases of recombination in the approximately 9 kb 'hot spot' region of the β

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\begin{array}{cccc}
\varepsilon & G_Y & A_Y & \psi\beta & \beta \\
I-1 & - & + & + & + & T & 1 \\
 & - & + & - & + & N & 4 \\
II-1 & - & + & + & + & T & 1 Pat \\
 & + & + & - & - & T & 12 Mat \\
II-2 & - & + & + & + & T & 1 Pat \\
 & + & - & - & - & T & 12 Mat \\
\end{array}
\]

FIG 3 Schematic representation of the chromosomes II of I.1, II.1, and II.2. The difference in the polymorphisms 5'β in II.1 and II.2 is evident. The boundaries of the hypothesised crossover between the paternal chromosomes are indicated by a dotted line.
22 Chehab FF, Honig GR, Kan YW. Spontaneous mutation in beta-thalassaemia producing the same nucleotide substitution as that in a common hereditary form. Lancet 1986;1:3–6.

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