ORIGINAL ARTICLE

Thyroid cancer susceptibility polymorphisms: confirmation of loci on chromosomes 9q22 and 14q13, validation of a recessive 8q24 locus and failure to replicate a locus on 5q24


ABSTRACT

Five single nucleotide polymorphisms (SNPs) associated with thyroid cancer (TC) risk have been reported: rs2910164 (5q24); rs6983267 (8q24); rs965513 and rs1867277 (9q22); and rs944289 (14q13). Most of these associations have not been replicated in independent populations and the combined effects of the SNPs on risk have not been examined. This study genotyped the five TC SNPs in 781 patients recruited through the TCUKIN study. Genotype data from 6122 controls were obtained from the CORGI and Wellcome Trust Case-Control Consortium studies. Significant associations were detected between TC and rs965513A (p = 6.35 x 10^{-39}), rs1867277A (p = 5.90 x 10^{-24}), rs944289T (p = 6.95 x 10^{-7}) and rs6983267G (p = 0.016). Rs6983267 was most strongly associated under a recessive model (pG vs GT + TT = 0.004), in contrast to the association of this SNP with other cancer types. However, no evidence was found of an association between rs2910164 and disease under any risk model (p > 0.7). The rs1867277 association remained significant (p = 0.008) after accounting for genotypes at the nearby rs966513 (p = 2.3 x 10^{-13}) and these SNPs did not tag a single high risk haplotype. The four validated TC SNPs accounted for a relatively large proportion (~11%) of the sibling relative risk of TC, principally owing to the large effect size of rs965513 (OR 1.74).

INTRODUCTION

Thyroid cancer (TC) is the most common endocrine malignancy and a complex disease with a largely unknown aetiology.1 TC is characterised by one of the strongest familial relative risks in cancer. First degree relatives of TC patients are up to 8.6 times more likely to develop TC than the general population.2 Most of the genetic variation associated with TC remains uncharacterised, and it is likely to be explained by variants of moderate or low penetrance.

A number of recent studies have identified single nucleotide polymorphisms (SNPs) associated with TC risk on chromosomes 5q24, 8q24, 9q22, and 14q13.3-6 Two of these SNPs, rs965513 (9q22) and rs944289 (14q13), were found through a multi-stage, genome-wide association (GWA) study in the Icelandic population.7 Subsequent replication of association was found in smaller sample sets from Ohio, USA and Spain.

The other three TC SNPs were discovered through candidate gene or SNP approaches.4-6 Rs965513 (9q24) was chosen because it lies within pre-miR-146a, a microRNA upregulated in TC. An association with TC was found in samples from Poland, Finland and Ohio, and the C allele at rs9210164 was found to decrease levels of pre- and mature miR-146A.4 Rs6983267 (8q24) is associated with the risk of several cancers, including those of the prostate, colon and ovary, and was assessed as a TC SNP for this reason. It showed a borderline significant association with TC in the Polish population.6 Rs1867277 was studied because it lies in the 5’ UTR of FOXE1 (or Thyroid Transcription Factor 2), a key gene involved in thyroid organogenesis.7 Rs1867277 and rs965513 are in moderate pairwise linkage disequilibrium (LD) in Europeans (r2 = 0.59, D’ = 0.73, http://www.1000genomes.org/). Rs1867277 was strongly associated with TC risk in Spanish and Italian cohorts.5 None of the three candidate SNP associations has been replicated in independent studies.

The aim of this study was to examine these five TC SNPs in a relatively large UK case-control sample set, to validate or refute their associations with TC in this population, and to estimate the proportion of the familial risk of TC for which they account.

PATIENTS AND METHODS

Study samples

We recruited 781 white UK patients of northern European origin with histologically confirmed non-medullary TC through the Thyroid Cancer Genetic investigation in the UK (TCUKIN) study. In addition to obtaining standard clinicopathological information from medical records and a questionnaire completed by each patient, the
participants donated a blood sample which was used to isolate genomic DNA. The Southampton and South West Hampshire Research Ethics Committee (A) approved the TCUKIN protocol.

**SNP genotyping and control genotype data**

We genotyped the TCUKIN samples at the five SNPs (rs2910164, rs6985267, rs965513, rs1867277, rs944289) using the KAspar system. Probes used to genotype these polymorphisms are shown in supplementary table 1. For comparison, we used available genotype data from 5193 population controls belonging to the National Blood Donor Service (NBS) and the 1958 Birth Cohort (BC58) and 929 cancer-free controls from our ColoRectal Gene Identification study (CORGI). The NBS and BC58 samples had been genotyped with Illumina 1.2M (Hap1.2M) arrays as part of the Wellcome Trust Case Control Consortium 2, and the CORGI controls had been genotyped with Illumina Hap550 arrays (Hap550, N = 932) as part of our ongoing studies in colorectal cancer genetics. Two of the five SNPs (rs2910164 and rs1867277) were not included on the Illumina 1.2M and Hap550 arrays, but had excellent proxy markers that facilitated their imputation. rs2910164 is perfectly tagged on the Hap550s by rs1443434 and on the Hap1.2Ms by rs1867278, rs1443434, rs965513, and rs12348691. These genotypes were imputed using IMPUTE2; all markers had proper_info scores >0.8 and imputation call rates >0.95, suggesting excellent imputation. In order to confirm this excellent tagging/imputation, 94 CORGI controls were genotyped for the TC SNPs using KAspar, resulting in no discordant genotypes for each SNP, whether typed or imputed.

**Quality control and statistical analysis**

General genotyping quality control assessment was as previously described. Duplicate samples were used to check genotyping quality and 100% concordance was found. Samples with multiple missing genotypes were eliminated from the analyses (N = 14). All five SNPs passed our quality control thresholds including call rates >95% and Hardy-Weinberg equilibrium p values >0.05.

Association statistics were obtained on per allele, genotypic and haplotype bases using logistic regression models implemented in PLINK, R, and SNPTEST. Haplotype analyses were carried out with HAPLOVIEW and PLINK. Allelic count association meta-analyses, using the Mantel-Haenszel method, were carried out in STATA. We used the IMPUTE2 software formally to generate rs2910164 and rs1867277 genotypes in the control population, although the fact that perfect proxies were used rendered this task of very limited utility. To test for independence between SNPs, we used conditional logistic regression models. The proportion of the familial relative risk explained by the polymorphisms investigated in the study was estimated using the method reported by Houlston et al.

**RESULTS**

**Single SNP analyses**

Three of the five SNPs examined showed a significant association with TC risk in the UK population (table 1). The strongest associations, on a per allele basis, were observed for the 9q22 SNPs rs965513A (p = 6.55 × 10⁻¹⁴, OR = 1.99, 95% CI 1.77 to 2.21) and rs1867277A (p = 5.90 × 10⁻²⁴, OR = 1.75, 95% CI 1.57 to 1.95). The association at rs944289T on 14q13 was also convincingly replicated (p = 6.95 × 10⁻¹⁷, OR = 1.53, 95% CI 1.18 to 1.48). For rs6985267G, we also found a nominally significant

**Table 1** Association statistics for thyroid cancer risk and genetic variants at chromosomes 5q24, 8q24, 9q22 and 14q13

<table>
<thead>
<tr>
<th>SNP, genotypes and risk allele</th>
<th>Frequency (%)</th>
<th>ORs for genotype or per allele overall (95% CI)</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>rs2910164</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>GG</td>
<td>438 (0.578)</td>
<td>3540 (0.584)</td>
<td>Reference</td>
</tr>
<tr>
<td>CG</td>
<td>271 (0.367)</td>
<td>2178 (0.360)</td>
<td>1.032 (0.876 to 1.214)</td>
</tr>
<tr>
<td>CC</td>
<td>41 (0.054)</td>
<td>339 (0.056)</td>
<td>0.987 (0.862 to 1.138)</td>
</tr>
<tr>
<td>Risk allele (C)</td>
<td>359 (0.238)</td>
<td>2857 (0.238)</td>
<td>1.013 (0.893 to 1.148)</td>
</tr>
<tr>
<td>rs6932627</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>TT</td>
<td>164 (0.218)</td>
<td>1441 (0.236)</td>
<td>Reference</td>
</tr>
<tr>
<td>GT</td>
<td>346 (0.461)</td>
<td>3012 (0.493)</td>
<td>1.010 (0.827 to 1.236)</td>
</tr>
<tr>
<td>GG</td>
<td>241 (0.321)</td>
<td>1662 (0.272)</td>
<td>1.274 (1.027 to 1.583)</td>
</tr>
<tr>
<td>Risk allele (G)</td>
<td>674 (0.449)</td>
<td>5894 (0.518)</td>
<td>1.140 (1.025 to 1.268)</td>
</tr>
<tr>
<td>rs965513</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>GG</td>
<td>187 (0.249)</td>
<td>2748 (0.449)</td>
<td>Reference</td>
</tr>
<tr>
<td>AG</td>
<td>394 (0.525)</td>
<td>2729 (0.446)</td>
<td>2.121 (1.763 to 2.559)</td>
</tr>
<tr>
<td>AA</td>
<td>170 (0.226)</td>
<td>642 (0.105)</td>
<td>3.883 (3.081 to 4.889)</td>
</tr>
<tr>
<td>Risk allele (A)</td>
<td>734 (0.499)</td>
<td>4015 (0.328)</td>
<td>1.981 (1.774 to 2.212)</td>
</tr>
<tr>
<td>rs1867277</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>GG</td>
<td>159 (0.211)</td>
<td>2290 (0.376)</td>
<td>Reference</td>
</tr>
<tr>
<td>AG</td>
<td>398 (0.529)</td>
<td>2879 (0.473)</td>
<td>1.991 (1.638 to 2.428)</td>
</tr>
<tr>
<td>AA</td>
<td>196 (0.260)</td>
<td>918 (0.151)</td>
<td>3.074 (2.446 to 3.864)</td>
</tr>
<tr>
<td>Risk allele (A)</td>
<td>790 (0.525)</td>
<td>4715 (0.387)</td>
<td>1.749 (1.569 to 1.950)</td>
</tr>
<tr>
<td>rs944289</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>CC</td>
<td>87 (0.116)</td>
<td>1003 (0.164)</td>
<td>Reference</td>
</tr>
<tr>
<td>CT</td>
<td>332 (0.441)</td>
<td>2924 (0.478)</td>
<td>1.309 (1.019 to 1.582)</td>
</tr>
<tr>
<td>TT</td>
<td>334 (0.444)</td>
<td>2193 (0.358)</td>
<td>1.755 (1.365 to 2.276)</td>
</tr>
<tr>
<td>Risk allele (T)</td>
<td>1000 (0.664)</td>
<td>7310 (0.597)</td>
<td>1.330 (1.188 to 1.498)</td>
</tr>
</tbody>
</table>

SNP, single nucleotide polymorphism.

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association with TC risk (p=0.016, OR=1.14, 95% CI 1.02 to 1.27, equivalent false discovery rate=0.020). However, the association between TC and rs2910164 was not replicated (P_{allelic}=0.85, OR=1.01, 95% CI 0.89 to 1.14). To test if the strength of these associations were similar in cases with different histological types, we carried out cases-only interaction analyses. We found no differences between cases with papillary and follicular histology (p=0.25 for all markers, data not shown), suggesting that these associations were not restricted to any particular histological type of TC.

### Are there multiple risk alleles on chromosome 9q22?

The 9q22 variants associated with TC risk, rs965513 and rs1867277, were originally reported by independent and non-overlapping GWA and candidate gene studies. Other studies have not considered whether these SNPs represent independent signals of association. We performed unconditional logistic regression analyses incorporating rs1867277 and rs965513 signals of association. We performed unconditional logistic regression analyses incorporating rs1867277 and rs965513 genotypes as variables and sex as a covariate in the model. Both regression analyses incorporating rs1867277 and rs965513.

To test if the strength of these associations were similar in cases with different histological types, we carried out cases-only interaction analyses. We found no differences between cases with papillary and follicular histology (p=0.25 for all markers, data not shown), suggesting that these associations were not restricted to any particular histological type of TC.

### TC is associated with variation at 8q24 under a recessive model

We found evidence that rs6983267G was associated with TC in the UK population (table 1). Interestingly, and unlike previous findings in other cancer types, rs6983267G was associated with TC risk according to a recessive model (tables 1 and 4). We found no difference in risk between non-carriers and heterozygotes (OR=1.01, 95% CI 0.83 to 1.24, p=0.921, table 1), but a significantly increased risk when homozygous carriers (GG) were compared to non-carriers (p=0.016, OR=1.14, table 1), to non-carriers/heteryzogotes (p=0.009, OR=1.27, table 1), and to non-carriers/heterozygotes (p=0.004, OR=1.26, table 4). Wokolorczyk et al had previously found relatively weak evidence of association between TC and the rs6983267 SNP in the Polish population.

### Table 2 Thyroid cancer risk associated with different haplotypes, defined by rs965513 and rs1867277 alleles, at chromosome 9q22

<table>
<thead>
<tr>
<th>Haplotype</th>
<th>rs965513</th>
<th>rs1867277</th>
<th>Frequency</th>
<th>OR (95% CI)</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Cases</td>
<td>Controls</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>G</td>
<td>G</td>
<td>0.413</td>
<td>1.00</td>
<td>Reference</td>
</tr>
<tr>
<td>2</td>
<td>A</td>
<td>A</td>
<td>0.428</td>
<td>2.139</td>
<td>1.902 to 2.407</td>
</tr>
<tr>
<td>3</td>
<td>G</td>
<td>A</td>
<td>0.097</td>
<td>1.189</td>
<td>0.978 to 1.485</td>
</tr>
<tr>
<td>4</td>
<td>G</td>
<td>A</td>
<td>0.061</td>
<td>1.612</td>
<td>1.284 to 2.037</td>
</tr>
</tbody>
</table>

Haplotype frequencies were estimated using Haploview (http://www.haploview.org/). Only samples with full data at both loci were used for the analyses (761 cases and 6085 controls).

### Table 3 Thyroid cancer risk associated with different genotype combinations (diplotypes) at rs965513 and rs1867277

<table>
<thead>
<tr>
<th>rs965513</th>
<th>rs1867277</th>
<th>Frequency (%)</th>
<th>OR (95% CI)</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>GG</td>
<td>GG</td>
<td>123 (0.162)</td>
<td>1917 (0.315)</td>
<td>Reference</td>
</tr>
<tr>
<td>AG</td>
<td>GG</td>
<td>60 (0.079)</td>
<td>744 (0.122)</td>
<td>1.257 (0.897 to 1.747)</td>
</tr>
<tr>
<td>AA</td>
<td>GG</td>
<td>7 (0.009)</td>
<td>73 (0.012)</td>
<td>1.494 (0.568 to 3.333)</td>
</tr>
<tr>
<td>AG</td>
<td>GG</td>
<td>33 (0.043)</td>
<td>355 (0.058)</td>
<td>1.449 (0.939 to 2.183)</td>
</tr>
<tr>
<td>AA</td>
<td>GG</td>
<td>300 (0.394)</td>
<td>1960 (0.322)</td>
<td>2.385 (1.908 to 2.995)</td>
</tr>
<tr>
<td>AA</td>
<td>GG</td>
<td>63 (0.083)</td>
<td>394 (0.065)</td>
<td>2.491 (1.774 to 3.473)</td>
</tr>
<tr>
<td>AG</td>
<td>GG</td>
<td>4 (0.005)</td>
<td>17 (0.003)</td>
<td>3.663 (0.883 to 11.465)</td>
</tr>
<tr>
<td>AA</td>
<td>GG</td>
<td>42 (0.055)</td>
<td>174 (0.029)</td>
<td>3.759 (2.497 to 5.580)</td>
</tr>
<tr>
<td>AA</td>
<td>GG</td>
<td>129 (0.170)</td>
<td>451 (0.074)</td>
<td>4.455 (3.379 to 5.876)</td>
</tr>
</tbody>
</table>

The risk alleles are rs965513A and rs1867277A.
rs2910164 at the pre-miR-146a locus is not associated with TC
Using an allelic model, our study found no evidence of an association between rs2910164 and TC risk ($P_{\text{naive}}=0.85$, table 1). We confirmed the absence of associations between this SNP and TC in genotypic, dominant, recessive and trend models ($p=0.71$ for all models, supplementary table 5). The previous report of an association between rs2910164 and papillary TC risk found for all models, data not shown, supplementary table 3). When the analyses were restricted to the cases that had histologically verified papillary TC we also failed to detect association between rs2910164 and TC ($p=0.32$, table 4).

**Combined effects of rs6983267, rs965513, rs1867277, and rs944289 on disease risk**
We carried out case-only and case-control pairwise analyses between all four risk SNPs associated with TC and found to no evidence for SNP–SNP interaction (details not shown). We then estimated the combined effects of the four SNPs on risk. To incorporate the effects of the two 9q22 markers in the combined risk analyses, we used the estimates obtained in our diplotype analyses (table 3). Using this information, we estimated that the risk for those individuals who are homozygous at 8q24, 9q22 and 14q13—comprising ~1.7% of the UK population—is 9.96-fold higher compared with individuals who do not carry any risk allele at these loci (~1.2% of the population, table 5). The risk homozygote at the four SNPs (17 cases and 41 controls, supplementary table 5) have a 17.03-fold higher chance of having TC (95% CI 3.776 to 159.323) when compared to non-risk homozygote (two cases and 84 controls, supplementary table 5).

**Contribution of 8q24, 9q22, and 14q13 variants to the familial risk of TC**
We have shown that four variants at 8q24, 9q22, and 14q13 are associated with a significantly higher risk of TC in the UK population. We then determined the proportion of the sibling relative risk of TC that they explained (15). Using a TC sibling relative risk of 8.6 (2), we estimated that these four risk variants explain 10.9% of the sibling relative risk of TC. Under a conservative model that assumes the existence of a single risk variant at chromosome 9q22, these loci explain at least 6.6% of the disease heritability (supplementary table 4).

**DISCUSSION**
Using a relatively large sample set in a single, homogeneous European population, we confirmed associations between TC and SNPs on chromosomes 9q22 (rs965513 and rs1867277), 14q13 (rs944289), and 8q24 (rs6983267). However, we failed to replicate an association between SNP rs2910164 on 5q24 and TC risk.

The 9q22 SNPs rs965513 and rs1867277 have not previously been genotyped in the same samples. We have found that there is not a single TC risk haplotype on 9q22 that is perfectly denoted by rs965513 and rs1867277. Conversely, the association cannot be explained entirely by genotypes at only one of the two SNPs, although logistic regression analysis incorporating both SNPs did lead to a considerably reduced association signal for both SNPs, particularly rs1867277. We suggest, therefore, that rs965513 and rs1867277 tag a third variant (or variants) that is the functional variation near FOXE1. Perhaps contrary to the genetic data, Landa et al5 showed that rs1867277A affected FOXE1 transcript levels through the differential recruitment of the USFI/USF2 transcription factors and suggested that rs1867277 was a TC-causal SNP. Fine mapping studies at this site might benefit from the use of non-European samples. For example, the LD between rs965513 and rs1867277 is significantly weaker in populations of African ($r^2=0.01$, D’=0.078) or Asian ($r^2=0.00$, D’=0.014) ancestry (data from the 1000 Genomes Project). Finally, although challenging,16 rare variants at FOXE1 deserve further scrutiny.

Our study validated the association between TC and rs6983267 and extended the range of cancer types associated with this variant. Interestingly, however, we found that rs6983267G is associated with TC risk in a recessive fashion; all other rs6983267 cancer associations follow an allelic dosage model. Recessive cancer predisposition SNPs have rarely been

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**Table 4** Evidence that the association between rs6983267 and thyroid cancer risk is best explained by a recessive model

<table>
<thead>
<tr>
<th>Test</th>
<th>OR (95% CI)</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>GG vs (GT + TT)</td>
<td>1.286 (1.071 to 1.494)</td>
<td>0.004</td>
</tr>
<tr>
<td>GG vs GT</td>
<td>1.282 (1.055 to 1.509)</td>
<td>0.009</td>
</tr>
<tr>
<td>GT vs TT</td>
<td>1.010 (0.827 to 1.236)</td>
<td>0.960</td>
</tr>
</tbody>
</table>

**Meta analysis of UK and Polish studies**

<table>
<thead>
<tr>
<th>Test</th>
<th>OR (95% CI)</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>GG vs (GT + TT)</td>
<td>1.250 (1.089 to 1.435)</td>
<td>7.64x10^-8</td>
</tr>
</tbody>
</table>

Note that the risk allele homozygotes (GG) have significantly higher frequency compared with other genotypes combined (GT + TT).

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**Table 5** Estimates to genotype relative risk at rs6983267, 9q22, and rs944289

<table>
<thead>
<tr>
<th>Locus</th>
<th>Non-carriers Population frequency</th>
<th>Homozygous carriers Population frequency</th>
</tr>
</thead>
<tbody>
<tr>
<td>rs6983267*</td>
<td>0.236 NA NA</td>
<td>1.274 0.272</td>
</tr>
<tr>
<td>9q22†</td>
<td>0.315 2.385 0.322</td>
<td>4.455 0.074</td>
</tr>
<tr>
<td>rs944289</td>
<td>0.164 1.309 0.478</td>
<td>1.755 0.358</td>
</tr>
<tr>
<td>Combined</td>
<td>0.012 3.122 0.210</td>
<td>9.961 0.017</td>
</tr>
</tbody>
</table>

*rs69833237 heterozygous do not have increased risk of thyroid cancer (see tables 1 and 4).
†ORs and frequencies for the 9q22 markers were obtained from the diplotype analysis presented in table 3. Population frequency from non-carriers, heterozygous and homozygous carriers is shown in table 3.
found. In part, this probably reflects suboptimal power in GWA studies and our finding emphasises the role of candidate SNP analyses across cancer types. Given that rs6983267 itself may be functional in predisposition to colorectal and other cancers, one possibility is that the true, recessive functional variation in TC is not rs6983267, but an SNP in strong LD with it.

The first association between TC and common genetic variants was found at the pre-miR-146a locus (4), a micro-RNA that is upregulated in thyroid tumours. There was no good evidence in our study of an association between rs2910164 and TC risk under all models tested, including the heterozygous model that showed the disease association in the study of Jadzewska et al. It is notable that deviations from Hardy-Weinberg equilibrium were present in the case genotypes of Jadzewska et al; it is not clear whether this was the result or the cause of the heterozygote association with TC risk. Other possible explanations for the differences between Jadzewska et al’s study and our own include chance, systematic differences between cases and controls (whether related to ascertainment or technical issues) and population specific effects in either study.

The four validated TC risk SNPs explain an approximately 10% fold differential risk between those with high risk alleles and those with all low risk alleles. Moreover, owing to the large effect size associated with the 9q22 SNPs, the four SNPs explain over 10% of the total sibling relative risk of TC, despite the fact that TC has one of the largest familial relative risks reported for any malignancy. It is highly plausible that future studies involving more than a few thousand cases and controls could identify additional important common risk variants for this common disease.

Author affiliations
1Wellcome Trust Centre for Human Genetics, University of Oxford, Oxford, UK
2Department of Endocrine Surgery, John Radcliffe Hospital, Oxford, UK
3Veilndre Cancer Centre, Cardiff, UK
4Mount Vernon Hospital, Northwood, UK
5Institute of Head and Neck Studies and Education, University Hospitals of Coventry and Warwickshire, Walsgrave, Coventry, UK
6Guys and St Thomas’ NHS Foundation Trust and King’s College London, London, UK
7Weston Park Hospital, Sheffield, UK
8Kent and Canterbury Hospital, Canterbury, UK
9York Hospital, York, UK
10Bristol Hematology and Oncology Centre, Bristol, UK
11Norfolk and Norwich University Hospital Trust, Norwich, UK
12Medway Maritime Hospital, Gillingham, UK
13St James University Hospital, Leeds, UK
14St Mary’s Hospital, London, UK
15Newcross Hospital, Wolverhampton, UK
16NHRI Comprehensive Biomedical Research Centre, University of Oxford, Oxford, UK

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Correction notice
This paper has been corrected since it was first published online. The corresponding author’s name should read Luis G Carvajal-Carmona.

Competing interests
None.

Patient consent
Obtained.

Ethics approval
Ethics approval was provided by the Southamption and South West Hampshire Research Ethics Committee (A).

Contributors
LGGC and IT are the leaders of TCUKIN and designed the study, performed the main statistical analyses and co-wrote the manuscript. AMJ, KMH, LM and MG carried out the experiments and coordinated sample collection. AA carried out most of the statistical analyses. C, H Mehanna, H Mohan, SEMC, JW, EM, AC, MB, TR, CM, PR, GG, DP, and CB are members of the TCUKIN Consortium, coordinated sample collection, and summarised the clinical data. All authors contributed to the final version of the manuscript.

Provenance and peer review
Not commissioned; externally peer reviewed.

Data sharing statement
The data presented in the manuscript are available on request.

References

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**APPENDIX 1**

Collaborators in the TCUKIN Study include: Dr Laura Moss, Velindre Cancer Centre, Cardiff CF14 2TL, UK; Dr Christopher Scrase, The Ipswich Hospital, Ipswich, IP4 5PD, UK; Dr Andrew Goodman, Royal Devon & Exeter Hospital, Exeter, EX2 5DW, UK; Dr Radu Mihaei, John Radcliffe Hospital, Oxford OX3 9DU, UK; Dr James Gildersleeve, Royal Berkshire Hospital, Reading RG1 5AN, UK; Dr Catherine Lemon, Mount Vernon Hospital, Northwood, HA6 2RN, UK; Dr Antony Robinson, Royal United Hospital, Bath BA1 3NG, UK; Dr Caroline Brammer, Newcross Hospital, Wolverhampton WV10 0QP, UK; Dr Georgina Gerrard, St. James University Hospital, Leeds LS9 7TF, UK; Professor Hisham Mehanna, Institute of Head and Neck Studies and Education, University Hospitals of Coventry and Warwickshire, Walsgrave, Coventry CV2 2ZK, UK; Dr Matthew Beasley, Bristol Hematology and Oncology Centre, Bristol BS2 8ED, UK; Dr Hosahalli K Mohan and Dr Susan EM Clarke, Guys and St Thomas’ NHS Foundation Trust and King’s College London, London SE1 9RT, UK; Dr Kate Goodchild, Luton & Dunstable Hospital, Luton LU4 0DZ, UK; Dr Jonathan Wadsley, Weston Park Hospital, Sheffield S10 2SJ, UK; Dr Abdel Hamid, Scunthorpe General Hospital, Scunthorpe DN15 7BH, UK; Dr Danielle Power, St. Mary’s Hospital, London W2 1NY, UK; Dr Elena Maccas, Kent and Canterbury Hospital, Canterbury, CT1 3NG, UK; Dr Jan Sharp, Royal Derby Hospital, Derby DE22 3NE, UK; Mr Andrew Cootsworth, York Hospital, York Y031 8HW, UK; Dr Hamish Courtney, Royal Victoria Hospital, Belfast BT12 6BA, UK; Dr Stephen Whittaker and Dr Katie Wood, Royal Surrey County Hospital, Guildford GU7 7XX, UK; Dr James McCauley, Bradford Royal Infirmary, Bradford BD9 6RJ, UK; Dr Christopher Ashford, Worcestershire Royal Hospital, Worcester WR5 TDD, UK; Dr Tom Roques and Dr Craig Martin, Norfolk and Norwich University Hospital NHS Trust, Norwich NR4 7UY, UK; Dr Vivienne Loo, Broomfield Hospital, Chelmsford CM1 7ET, UK; Dr Jennifer Marshall, Southampton General Hospital, Southampton SO16 6DY, UK; Mr Amy Roy, Derriford Hospital, Plymouth PL6 8DH, UK; Dr Joanna Simpson, The Royal Sussex County Hospital, Brighton BN2 5BE, UK; Dr Nick Rowell, Maidstone Hospital, Maidstone ME16 9EQ, UK; Mr Edward Babu, Hillingdon Hospital, Uxbridge UB8 3NN, UK; Dr Narayanan Srinjan, Royal Shrewsbury Hospital, Shrewsbury SY3 8QZ, UK; Dr Simon Ellenbogen, Tameside General Hospital, Ashton-under-Lyne OL6 9RW, UK; Dr Paul Ryan, Midway Maritime Hospital, Gillingham ME7 5NY, UK; Dr Arshad Jamil, University Hospital North Staffordshire, Stoke on Trent ST4 6GQ, UK.
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