Mutation screening of Mre11 complex genes: indication of RAD50 involvement in breast and ovarian cancer susceptibility

K Heikkinen, S-M Karppinen, Y Soini, M Mäkinen, R Winqvist

J Med Genet 2003;40:e131 [http://www.jmedgenet.com/cgi/content/full/40/12/e131]

Approximately 5–10% of all breast and ovarian cancers are due to genetic predisposition.1 Germinal mutations in two major susceptibility genes, BRCA1 and BRCA2, explain most of the hereditary ovarian cancer cases, but worldwide have been observed only in a fraction of the breast cancer families; in Finland, about 20%.2–4 A small portion of the remaining cases, perhaps 1–2%, appears to be caused by germine mutations in other cancer susceptibility genes such as TP53, PTEN, AR, and ATM.5 Recently, moderate or low penetrance germine mutations such as 1100delC of CHK2 have also been implicated,6 suggesting that in many cases the phenotypic expression of the disease is accounted for by the combined effect of multiple but more subtle mutations in different target genes.6–7 Despite all these discoveries, many breast cancer families, especially those with fewer than four cases and/or with later disease onset, have cancers due to mutations in still unknown genes. As the protein products of the genes so far indicated are central players in the pathways involved in cell cycle checkpoint functions and in the sensing, transduction, and repair of DNA lesions,8 other similarly acting genes may represent new potential candidates for breast and/or ovarian cancer susceptibility.

The Mre11 complex, composed of the proteins MRE11, NBS1, and RAD50, is a vital component in the DNA damage response and is at least partially responsible for connecting DNA damage detection to DNA repair and cell cycle checkpoint functions. The early association of the complex with damaged DNA suggests its involvement in sensing DNA double strand breaks (DSBs). In eukaryotes, DSB repair is handled by two different pathways, homologous recombination (HR) and non-homologous end joining (NHEJ), and the initial step in both is the recognition and signalling of the damage by the Mre11 complex.8–10 In addition to DNA damage detection and repair, the complex has an important role in co-ordinating checkpoint responses and DNA replication,11–14 and is also required for the maintenance of telomeres.15

The sites important for the composition and various functional activities of the Mre11 complex have been characterised (summarised in fig 1). Because of its independent interaction with both NBS1 and RAD50, MRE11 has been viewed as the core of the complex, whereas the interaction between NBS1 and RAD50 is indirect and mediated through MRE11.16–17 The sites in MRE11 responsible for binding to NBS1 and RAD50 have both been assigned to the aminoterminal half of the protein.16–18 In NBS1, the MRE11 binding domain has been located to the extreme carboxyterminus of the protein,16 and the corresponding site in RAD50 has been located to a 40 residue coiled coil region adjacent to the ABC domain.19–20 Additionally, several other important domains for the complex function have been identified. The DNA nuclease activity is restricted to MRE11 and specified by phosphodies-

Key points

- The Mre11 complex has an essential role in maintaining genomic integrity and preventing cells from malignancy. The complex components interact with several proteins (for example BRCA1, ATM and CHK2) already connected to hereditary breast and ovarian cancer susceptibility, which makes MRE11, NBS1, and RAD50 suitable candidate genes for predisposition to these cancers.
- We screened the affected index cases of 151 families with signs of hereditary susceptibility to breast and/or ovarian cancer for germline mutations in the Mre11 complex genes. Seven novel heterозygous alterations were observed, three of which were considered potentially disease-related: MRE11 913C>T (Arg305Trp), NBS1 448C>T (Leu150Phe) and RAD50 687delT (stop codon at 234).
- RAD50 687delT is expected to cause major truncation in both known protein isoforms and occurs in multiple individuals of two families. Curiously, RAD50 687delT was also present in some of the controls, but at a lower frequency, probably referring to incomplete penetrance. The MRE11 and NBS1 alterations affected highly conserved amino acids and were seen in single cancer cases, but they were both absent in 1000 healthy controls.
- Our results suggest that these three mutations in the Mre11 complex genes associate with hereditary susceptibility to breast and/or ovarian cancer, and may be important in other types of cancer too. This is the first time to our knowledge that germline mutations in RAD50 have been linked to genetic disease in humans.

Abbreviations: AT, ataxia telangiectasia; ATLD, ataxia telangiectasia-like disorder; BASC, BRCA1 associated genome surveillance complex; BRCT, BRCA1 carboxyterminus; CSGE, conformation sensitive gel electrophoresis; DSB, double strand break; FHA, forhead associated; HR, homologous recombination; NBS, Nijmegen breakage syndrome; NHEJ, non-homologous end joining.
subcellular localisation after DNA damage.\textsuperscript{17} \textsuperscript{22} RAD50 in turn consists of bipartite N- and C-terminal ATPase segments, which assemble into a single ABC-type cassette at the end of the predicted antiparallel coiled coil.\textsuperscript{23} A functionally important zinc hook has been identified in the central portion of the coiled coil domain.\textsuperscript{24} This zinc hook is thought to mediate RAD50 dimerisation, which allows the required molecular flexibility of the Mre11 complex and keeps it functionally assembled during the DNA recombination and repair process.\textsuperscript{14}

The Mre11 complex proteins interact with several important protein partners, many of which have been related to cancer predisposition. For instance, following ionising radiation, NBS1 is phosphorylated in an ATM dependent manner to ensure proper response to DNA damage.\textsuperscript{15} In turn, MRE11 phosphorylation relies on NBS1.\textsuperscript{24} Co-operation between a functional Mre11 complex and ATM is also required for activation of the CHK2 kinase in response to DNA damage.\textsuperscript{15}

In addition, MRE11, NBS1 and RAD50 are linked through protein–protein interactions with BRCA1. After exposure to ionising radiation, BRCA1 forms nuclear foci that co-localise with the Mre11 complex.\textsuperscript{25} Along with BRCA1, ATM, and other proteins, MRE11, NBS1, and RAD50 are also part of a multi-subunit protein complex referred to as the BRCA1 associated genome surveillance complex (BASC).\textsuperscript{26} Defects in the DNA damage response and repair processes causes chromosomal instability that may result in cancer predisposition. For two of the Mre11 complex genes, namely MRE11 and NBS1, the connection to cancer has already been established: germline mutations in \textit{MRE11} are associated with the ataxia telangectasia-like disorder (ATLD).\textsuperscript{27} whereas defects in \textit{NBS1} cause Nijmegen breakage syndrome (NBS).\textsuperscript{21} Cells derived from ATLD and NBS patients display similar characteristics as those from ataxia telangectasia (AT) patients, and AT due to germline mutations in the \textit{ATM} gene has been associated with an increased risk of developing various malignancies; breast cancer, lymphoma, and leukaemia in particular.\textsuperscript{28} As yet, germline mutations in \textit{RAD50} have not been connected to genetic diseases in humans. However, in mice certain germline \textit{RAD50} mutations result in growth defects and cancer predisposition.\textsuperscript{29} Because mutations in \textit{ATM}, \textit{CHK2}, and \textit{BRCA1} have all been previously linked to hereditary predisposition to breast and/or ovarian cancer\textsuperscript{17} it is possible that germline mutations in the individual Mre11 complex genes may also be directly involved in predisposition to these cancers. The suggested connection has been supported by a recent study, where the \textit{NBS1} 657del5 founder mutation was found to be responsible for the occurrence of a small but significant portion of familial breast cancer cases.\textsuperscript{30}

On these grounds, we tested 151 index patients from Finnish families with signs of hereditary susceptibility to breast and/or ovarian cancer for possible germline mutations in the \textit{MRE11}, \textit{NBS1}, and \textit{RAD50} genes. The results of our study indicate novel disease associated alterations.

**MATERIALS AND METHODS**

**Subjects and unaffected controls**

Index cases diagnosed with breast or ovarian cancer of 151 families originating from northern Finland were selected for the screening of germline \textit{MRE11}, \textit{NBS1}, and \textit{RAD50} mutations. Of the studied families, 95 were associated with breast, 29 with breast–ovarian, and 4 with ovarian cancer. These families met the criteria for either high (76 families) or moderate (52 families) genetic susceptibility to breast and/or ovarian cancer. Inclusion criteria for the families were one or more of the following: (a) two to three or more cases of breast and/or ovarian cancer in first or second degree relatives; (b) early disease onset (<35 years); (c) bilateral breast cancer; or (d) multiple primary tumours including breast or ovarian cancer in the same individual. The high risk families mainly included three or more cancer cases. All of the high risk families had been screened for germline mutations in \textit{BRCA1, BRCA2, CHK2}, and \textit{TP53}, and of eleven of them were found to be \textit{BRCA1} or \textit{BRCA2} mutation positive.\textsuperscript{31} \textsuperscript{32} Besides high or moderate risk families, 23 families showing single cases of breast or ovarian cancer along with multiple cases (two or more) of other kinds of cancer in first or second degree relatives were included in the study. Additionally, 1000 anonymous cancer free individuals were used as controls, and the occurrence of novel, potentially pathogenic alterations was also investigated in 192 breast cancer patients without family history of the disease. Both groups derived from the same geographical area as the studied families. All patients had given their informed consent for obtaining pedigree data and blood specimens for a study on cancer susceptibility gene mutations. Approval to perform the study was obtained from the ethics board of the Northern Ostrobotnia Health Care District and the Finnish Ministry of Social Affairs and Health.

**Screening for germline mutations**

DNA from blood lymphocytes was extracted using the standard phenol–chloroform method or the Puregene D-50K purification kit (Gentra, Minneapolis, MN, USA). The coding regions and exon–intron boundaries of the \textit{MRE11}, \textit{NBS1}, and \textit{RAD50} genes were screened for germline mutations by conformation sensitive gel electrophoresis (CSGE), which has been shown to be a cost efficient way to scan genes with high sensitivity and specificity.\textsuperscript{43} \textsuperscript{33} Samples with band shift were reamplified and the sequencing analysis was performed with the Li-cor IR 4200-S DNA Analysis system (Li-cor Inc., Lincoln, NE, USA) using the SequiTherm EXE\textsuperscript{TM} II DNA Sequencing kit \textit{LC} (Epicentre Technologies, Madison, WI, USA). Oligonucleotides for CSGE and sequencing were designed by using Primer3 software based on sequence information obtained from publicly available databases. Oligonucleotide sequences and PCR conditions are available upon request.

**LOH analysis**

Loss of heterozygosity (LOH) analysis was performed to assess whether the tumours of the patients displaying the novel \textit{MRE11} and \textit{RAD50} germline mutations had lost the wildtype allele. The flanking microsatellite DNA markers used for \textit{MRE11} were D11S4176 and D11S1757; and for \textit{RAD50} D5S2110, D5S2057, D5S1984, D5S2002, and D5S2117. The PCR products were analysed with the Li-cor DNA Analysis system using the IRD800 labelled forward primer.
Evaluation of RAD50 687delT expression at the mRNA level

Expression of the mutated allele at the mRNA level was evaluated for RAD50 687delT. Using the FastTrack® 2.0 Kit (Invitrogen, Carlsbad, CA, USA), mRNA from lymphoblast cell lines was isolated, following which cDNA synthesis was carried out with the GeneRacer™ kit (Invitrogen). The use of the GeneRacer™ Kit ensured that only transcripts containing full length cDNA ends were used in PCR analysis. Primers used for the cDNA specific amplification of RAD50 exon 5 were: forward 5’-TCATTGTCAAGTTTCATTATTTTAGAG-3’ (located in exon 4) and reverse 5’-TCATTGTCAGTTTCATTATTTTAGAG-3’ (located in exon 6).

RESULTS

Novel germline alterations

The mutation analysis of the MRE11, NBS1, and RAD50 genes revealed several germline alterations. Eleven heterozygous changes in protein encoding regions were observed (summarised in tables 1 and 2). Seven of the alterations were novel and three of these were considered potentially disease related. In addition, a large number of different apparently harmless alterations occurring in intron regions was seen (data not shown).

MRE11 and NBS1

In the MRE11 gene, a single mutation in exon 10 was found in one index case with ovarian cancer (case 322, fig 2A). This novel single base change, a transition C→T at position 913, resulted in substitution of arginine to tryptophan at codon 305. The position of the involved arginine is extremely conserved between several species, even those distantly related (fig 4). Additionally, the flanking regions on both sides of this residue are well conserved, suggesting functional importance.

In contrast to MRE11, multiple alterations within the protein encoding region of NBS1 were detected, but only one was considered potentially pathogenic. This novel change, a C→T transition at nucleotide 448, results in substitution of leucine to phenylalanine at codon 150 and was displayed by one index case diagnosed with breast cancer (case 277, fig 2B). Interestingly, this alteration occurs in the middle of the highly conserved BRCT domain, where the leucine residue at position 150 is conserved also in the mouse, rat, and chicken orthologues (fig 4).

RAD50

In RAD50, four novel germline changes affecting exon regions were discovered (tables 1 and 2). Of these changes, His68His appears to be a harmless polymorphism, but the roles of Ile94Leu and Arg224His alterations remain more difficult to determine. Both of these substitutions affect conserved residues, the former in the remarkably well conserved aminoterminal ATPase domain and the latter in the coiled coil domain, but they also occurred in the healthy controls, although at lower frequency. Interestingly, one of the breast cancer families displaying the RAD50 Ile94Leu variant also showed partial co-segregation with the previously identified CHK2 Ile157Thr unknown variant. Until further clarifying studies have been carried out, the cancer predisposing role of these two RAD50 variants remains unknown. However, at least one of the identified RAD50 alterations, 687delT, is likely to be disease related.

The RAD50 687delT truncation mutation

The RAD50 687delT mutation in exon 5 leads to a premature stop at codon 234 and affects both isoforms of the protein.

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### Table 1: Observed sequence variants occurring in the exon regions of the Mre11 complex genes

<table>
<thead>
<tr>
<th>Gene</th>
<th>Exon</th>
<th>Nt change</th>
<th>Effect on protein</th>
<th>Allele frequency*</th>
</tr>
</thead>
<tbody>
<tr>
<td>MRE11</td>
<td>10</td>
<td>913C→T</td>
<td>Arg305Trp</td>
<td>0.7% (1/151) – (0/192) – (0/1000)</td>
</tr>
<tr>
<td>NBS1</td>
<td>4</td>
<td>448C→T</td>
<td>Leu150Pro</td>
<td>0.7% (1/151) – (0/192) – (0/1000)</td>
</tr>
<tr>
<td>RAD50</td>
<td>5</td>
<td>687delT</td>
<td>Frameshift → stop codon at 234</td>
<td>1.3% (2/151) – (0/192) – 0.6% (6/1000)</td>
</tr>
</tbody>
</table>

*Heterozygotes.

### Table 2: Observed sequence variants occurring in the exon regions of the Mre11 complex genes

<table>
<thead>
<tr>
<th>Gene</th>
<th>Exon</th>
<th>Nt change</th>
<th>Effect on protein</th>
<th>Hereditary cases</th>
<th>Sporadic cases</th>
<th>Controls</th>
</tr>
</thead>
<tbody>
<tr>
<td>NBS1</td>
<td>2</td>
<td>102G→A</td>
<td>Leu34Leu†</td>
<td>39.1% (59/151)</td>
<td>ND</td>
<td>ND</td>
</tr>
<tr>
<td></td>
<td>5</td>
<td>553G→C</td>
<td>Glu185Gln†</td>
<td>43.7% (66/151)</td>
<td>ND</td>
<td>ND</td>
</tr>
<tr>
<td></td>
<td>10</td>
<td>1197T→C</td>
<td>Asp399Asp†</td>
<td>50.3% (76/151)</td>
<td>ND</td>
<td>ND</td>
</tr>
<tr>
<td></td>
<td>11</td>
<td>1720A→C</td>
<td>Leu574ILE</td>
<td>0.7% (1/151)</td>
<td>1.3% (4/307)‡</td>
<td>ND</td>
</tr>
<tr>
<td>RAD50</td>
<td>13</td>
<td>201G→A</td>
<td>Pro272Thr</td>
<td>37.1% (56/151)</td>
<td>ND</td>
<td>ND</td>
</tr>
<tr>
<td></td>
<td>2</td>
<td>204C→T</td>
<td>His68His</td>
<td>0.7% (1/151)</td>
<td>ND</td>
<td>ND</td>
</tr>
<tr>
<td></td>
<td>3</td>
<td>280A→C</td>
<td>Ile94Leu</td>
<td>1.3% (2/151)</td>
<td>0.3% (3/1000)</td>
<td>ND</td>
</tr>
<tr>
<td></td>
<td>5</td>
<td>671G→A</td>
<td>Arg224His</td>
<td>2.0% (3/151)</td>
<td>0.9% (9/1000)</td>
<td>ND</td>
</tr>
</tbody>
</table>

*Heterozygotes.
†Reported by Varon et al (1998) or the SNP database.
‡Only 307 controls were tested for this alteration.
ND, not done.
The deletion was found in two separate families (fig 3). In family 5, the index case and her sister (cases 828 and 474), both diagnosed with breast cancer, were carriers of the mutation. In family 6, the index (case 505) with ovarian cancer carried $\text{RAD50}^{687\text{delT}}$, but had also tested positive for the $\text{BRCA1}^{3745\text{delT}}$ mutation in one of our previous studies.4 In addition, the deceased mother with ovarian cancer was an obligatory carrier of at least the $\text{RAD50}^{687\text{delT}}$ mutation. In all the other cases tested, both the $\text{RAD50}$ and the $\text{BRCA1}$ truncation mutation segregated incompletely with the disease.

The observation of unaffected $\text{RAD50}$ truncation mutation carriers in family 6, and also the surprisingly high prevalence at the population level (0.6%) suggested not only that $\text{RAD50}^{687\text{delT}}$, if truly pathogenic, exhibits low penetrance, but also that it might predispose carriers to cancer without clustering of the disease in the pedigrees. In order to test this hypothesis, we screened 192 breast cancer cases without family history of the disease for the $\text{RAD50}$ truncation mutation. In parallel, we also screened the same cases for the $\text{MRE11}$ and $\text{NBS1}$ mutations, as the disease segregation of these two alterations could not be confirmed from the pedigrees. As none of the tested alterations was observed in the studied cohort, it may suggest that these changes are mainly associated with patients with a family history of cancer.

As $\text{RAD50}^{687\text{delT}}$ is expected to cause truncation of most of the protein product, we evaluated whether these mutant transcripts are eliminated by nonsense mediated decay. Lymphoblast cell lines generated from blood specimens of three different mutation positive individuals (fig 3: family 5, case 828; family 6, cases 505 and 061) was used to produce mRNA for the cDNA synthesis. The results from sequencing analysis unambiguously demonstrated that in all studied cases both the wild type and the truncation allele were still present in the tested mRNA pool (data not shown).

**MRE11 and RAD50 LOH analysis**

In order to test the nature of the observed mutations, we investigated whether the tumorigenesis thought to be
associated with the novel germline mutations involved loss of the corresponding wild type allele. LOH analysis was performed on the available tumour specimens from four different patients (cases 322, 828, 474, and 505; figs 2 and 3) displaying either the MRE11 or RAD50 mutation, but no LOH was observed (data not shown). Owing to the lack of suitable tumour material the corresponding LOH analysis for the NBS1 mutation could not be performed.

**DISCUSSION**

The Mre11 complex is an integral part of the proper DNA damage response and repair of double strand breaks, essential for the wellbeing of any living organism. Failure in these highly conserved processes leads to the loss of genomic integrity and consequently to an elevated risk of cancer.

The cellular functions of the Mre11 complex are indispensable, as seen in mice, where major deficiencies in the Mre11 complex lead to embryonic lethality and only hypomorphic mutations appear to be viable. In humans, only a small number of hypomorphic germline mutations affecting the MRE11 and NBS1 genes are currently known, mainly associating with the severe and cancer prone phenotypes of ATLD and NBS, respectively. Disease related germline mutations in RAD50 have not to our knowledge previously been reported.

In the current study, we have examined the prevalence of NBS1, MRE11, and RAD50 germline mutations in index cases with breast or ovarian cancer from 151 families. Of the novel heterozygous sequence alterations observed, Arg305Trp (MRE11), Leu150Phe (NBS1), and 687delT (RAD50) potentially relate to cancer susceptibility (table 1). As the LOH analysis of tumours from MRE11 or RAD50 mutation carriers showed retention of the wild type alleles, it may indicate that these germline mutations are sufficient to cause the observed adverse phenotypic effect. However, silencing of the wild type allele by epigenetic mechanisms (for example by promoter methylation) cannot be excluded.

The association of the novel MRE11 and NBS1 missense mutations with hereditary cancer susceptibility is supported by their absence in both the controls and the unselected breast cancer cases. In addition, both amino acid substitutions occur in highly conserved protein domains. In MRE11, the Arg305Trp substitution takes place at the very end of the first of two major protein domains, replacing the conserved positively charged hydrophilic arginine with the neutral and highly hydrophobic tryptophan. Because of its position in the protein, it is possible that the Arg305Trp substitution may affect the structural organisation of the functionally active L-shaped groove located between domains 1 and 2, and perhaps lower the affinity of MRE11 for DNA binding. The situation could be similar to that indicated for the Asn117Ser mutation in ATLD; despite the fact that the change occurs relatively remote from the active site, it has been suggested to disturb the macromolecular interaction of MRE11. In NBS1, the Leu150Phe substitution involves the highly conserved BRCT domain (fig 4), where the conserved leucine is replaced with the biochemically non-similar phenylalanine. Interestingly, another amino acid substitution affecting a conserved residue in the BRCT domain, Ile171Val, has been associated with acute lymphoid leukaemia. Additionally, recent site specific mutagenesis experiments by Cerosaletti & Concannon indicate that changes at the conserved residues GlyGly136–137GluGlu or Tyr176Ala are capable of disrupting Mre11 complex nuclear focus formation and blocking NBS1 phosphorylation after irradiation, emphasising the functional importance of this region and especially the role of the conserved amino acids in the BRCT domain. In order to obtain proof of the proposed deleterious nature of the MRE11 and NBS1 missense mutations discovered in the current study, the biochemical effects of these alterations has to be demonstrated by functional assays. Furthermore, additional case-control series are needed to confirm their possible role of as cancer susceptibility alleles.

Of the novel germline alterations observed in this study, RAD50 687delT was probably the most dramatic as it is predicted to cause about 80% shortening of both isoforms of the protein. The truncated protein would have retained only the N-terminal portion of the sequence necessary for binding of the RAD50 dimer to MRE11, but would have lost the corresponding and equally important C-terminal site. Based on studies of lymphoblast cell lines we know that the deletion allele is expressed at the mRNA level, but it is currently unknown if it actually gives rise to a truncated protein, how stable this product would be, and whether it has effects on the function of the wild type protein, perhaps by disturbing the functionally important dimerisation of RAD50. If a shortened protein actually is produced, but is unstable, the effect is likely to be that of a null mutation. This dosage variation due to haploinsufficiency may have a direct association with the corresponding LOH analysis for the NBS1 mutation could not be performed.

<table>
<thead>
<tr>
<th>MRE11</th>
<th>PDE motif IV</th>
<th>PDE motif V</th>
<th>Arg305Trp</th>
</tr>
</thead>
<tbody>
<tr>
<td>hs</td>
<td>V125H-QNRSKHG-TNF1PEQGLDDFIDLV-VWGCHEEC-KVTAPK-TKEQQL-FY11S-QPQSSS-VTSLS-PGEAVXKHV-GLLRK-171X5-C128</td>
<td></td>
<td></td>
</tr>
<tr>
<td>mm</td>
<td>V125H-QNRSKHG-TNF1PEQGLDDFIDLV-VWGCHEEC-KVTAPK-TKEQQL-FY11S-QPQSSS-VTSLS-PGEAVXKHV-GLLRK-171X5-C128</td>
<td></td>
<td></td>
</tr>
<tr>
<td>rg</td>
<td>V125H-QNRSKHG-TNF1PEQGLDDFIDLV-VWGCHEEC-KVTAPK-TKEQQL-FY11S-QPQSSS-VTSLS-PGEAVXKHV-GLLRK-171X5-C128</td>
<td></td>
<td></td>
</tr>
<tr>
<td>NBS1</td>
<td>Leu150Phe</td>
<td>Leu150Phe</td>
<td>Leu150Phe</td>
</tr>
<tr>
<td>hs</td>
<td>109 EYPEIVC-VCSCLDSVQKTALNQAILGLQGFTVNWTTEECHRKMMSVVKTVKICAGRVPKFPYKTEFELKAVGSK 196</td>
<td>136-137</td>
<td>171-176</td>
</tr>
<tr>
<td>mm</td>
<td>109 EYPEIVC-VCSCLDSVQKTALNQAILGLQGFTVNWTTEECHRKMMSVVKTVKICAGRVPKFPYKTEFELKAVGSK 196</td>
<td>136-137</td>
<td>171-176</td>
</tr>
<tr>
<td>gg</td>
<td>109 EYPEIVC-VCSCLDSVQKTALNQAILGLQGFTVNWTTEECHRKMMSVVKTVKICAGRVPKFPYKTEFELKAVGSK 196</td>
<td>136-137</td>
<td>171-176</td>
</tr>
</tbody>
</table>

Figure 4 Novel amino acid substitutions in MRE11 and NBS1, and comparison of sequence homology between Homo sapiens (hs), Rattus norvegicus (mm), and Gallus gallus (gg). The sites showing changed residues are boxed. The amino acids conserved in all four species are drawn in blue, biochemically similar residues in green, and dissimilar ones in black. The sequence shown for MRE11 extends to its last two phosphodiesterase motifs (PDE IV and V) and for NBS1 corresponds to its BRCT domain. In NBS1 the two sites (136–137 and 176) marked with an asterisk (*) have been found to be critical for Mre11 complex nuclear focus formation and phosphorylation of NBS1 after irradiation, and the one marked with an arrowhead (171X5) has been associated with acute lymphoid leukaemia.
influence on the activities in DNA damage recognition and response pathways, and could modify the cancer risk by increasing genetic instability or by altering the effect of the normal DNA damage response.\textsuperscript{41} However, the possibility that RAD50 687delT is a hypomorphic mutation, analogous to the common Slavic NBS founder mutation NBS1 657delE,\textsuperscript{42} also cannot be excluded. Interestingly, according to the recent study by Gorski \textit{et al}, the frequency of the NBS1 657delE mutation in the population controls they used is comparable with that of RAD50 687delT in our study (both 0.6%).

The RAD50 687delT mutation was observed altogether in seven individuals, two of whom showed breast and two ovarian cancers, all at relatively young age (41–50 years old). However, in family 6 (fig 3 right) the situation is less straightforward, as at least one of the affected individuals (case 505) showed parallel involvement of the previously identified BRCA1 3745delT truncation mutation.\textsuperscript{43} In addition, there were several unaffected carriers of both mutations, but the three daughters of case 505 found positive for either the RAD50 or the BRCA1 mutation, or both, may not yet contribute to the understanding of the disease penetrance owing to their relatively young age. Although at present it is difficult to determine the exact role of these two germline mutations in this family, there are several scenarios worth exploring. It is possible that both RAD50 and BRCA1 act independently on the resultant phenotype. On the other hand, co-occurrence of RAD50 687delT could also strengthen the effect of the BRCA1 mutation. Interestingly, a recent comparative genome hybridisation targeted linkage analysis has revealed a possible BRCA1 penetrance modifier locus on 5q in a location close to RAD50.\textsuperscript{44} Besides penetrance modification, the RAD50 mutation may also in some way influence the tissue specificity of a BRCA1 associated malignancy. However, it is also possible that in the genetic background provided by this particular family the mutation in RAD50 does not significantly increase the risk of cancer in BRCA1 5745delT carriers.

The surprisingly high frequency of RAD50 687delT mutation carriers among the healthy population controls and its absence from the breast cancer cases without familial disease background suggests that RAD50 687delT may be a new low penetrance susceptibility allele associated particularly with hereditary predisposition to cancer. At present it is unknown whether the RAD50 687delT allele exists outside Finland, as it is a well known fact that many disease related gene defects can be either unique or highly enriched in the Finnish population, owing to founder effects, isolation, and genetic drift.\textsuperscript{45} However, the situation could also be similar to that observed for the cancer predisposing CHK2 1100delC allele, which is enriched in southern Finland,\textsuperscript{46} but also occurs at significant frequencies in many other European populations (for example, the UK). In order to obtain more information about the geographical occurrence and the overall relevance of the RAD50 687delT mutation, the intention is to study additional breast, ovarian, and other hereditary cancer materials along with suitable control specimens, initially from the Nordic countries. In addition, functional studies are needed to reveal the exact details about the molecular consequences of the RAD50 687delT mutation. In conclusion, this is the first time that germline mutations in RAD50 have been linked to genetic diseases in humans. The RAD50 deletion allele reported in this study provides evidence that deleterious germline mutations in the third member of the Mre11 complex exist, are viable, and appear to associate with predisposition to cancer. Additional investigations will be needed to confirm and follow up these preliminary leads on the role of Mre11 complex gene mutations in hereditary susceptibility to cancer.

ACKNOWLEDGEMENTS
The authors wish to thank Drs G Blanco, U Puistola, J Leisti and J Ignatiu, and Nurse O Kajula for their help in patient contacts. We are grateful to Dr A Borg and K Rapakko MSc for comments on the manuscript, and to Dr M Alline for contribution in the early phases of this study. The expertise and helpfulness of Dr M Heikkinen in several clinical issues is highly appreciated. We are most thankful for the skilful assistance of K Outila in performing part of the mutation screening work. We wish to thank the Academy of Finland, University of Oulu, Oulu University Hospital, Nordic Cancer Union, and the Cancer Foundation of Northern Finland for providing the means for the current investigation. The kind participation of all patients has been of utmost importance for performing this study.

ELECTRONIC DATABASE INFORMATION

The accession numbers are: AP0009786, NM_005590 and NM_005591 for MRE11; AB013139 and NM_002485 for NBS1; NT_007072, NM_05732 and NM_133482 for RAD50.


Authors’ affiliations
K Heikkinen, S-M Karppinen, R Winquist, Departments of Clinical Genetics
Y Soini, M Mäkinen, Pathology, University of Oulu, Oulu University Hospital, Oulu, Finland

Correspondence to: Dr R Winquist, Department of Clinical Genetics, Oulu University Hospital, PO Box 24, FIN-90029 OYS, Finland; robert.winquist@oulu.fi

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K Heikkinen, S-M Karpinnen, Y Soini, M Mäkinen and R Winqvist

*J Med Genet* 2003 40: e131
doi: 10.1136/jmg.40.12.e131

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