Instability in the normal CTG repeat range at the myotonic dystrophy locus

We present a family clinically unaffected by dystrophia myotonica (DM), in whom molecular studies detected different CTG repeat numbers in sibs of one generation and also remarkable CTG repeat instability in two consecutive generations in the range of <50 CTG copies. In our family, paternal transmission (father 37 repeats, son 48 repeats) resulted in a maximum increase of 11 CTG repeats in the next generation. This increase has not previously been observed in unaffected families. The intergenerational unstable CTG repeats showed no somatic mosaicism in blood lymphocytes. CTG repeat numbers were defined by polymerase chain reaction (PCR) and a refined Southern protocol.

The CTG repeat region in the 3′ untranslated region of the DMPK gene on 19q13.2-3 is highly variable in DM patients and also in unaffected probands. In the normal population the CTG repeat alleles are stably inherited. Because of our interest in CTG repeat allele frequencies at multiple loci, including the DM locus, in the German population we have asked families to participate in molecular testing. Here we report the results of an eight member, three generation family (fig 1A). Blood samples from subjects I.2 and III.1 were not available. All other family members donated peripheral blood for the investigation after informed consent. No signs of DM were found in the family members after a thorough neurological examination, nor were there any indications of genetically determined disorders. Subject I.2 had died of lung oedema at 65 years. He had no symptoms of DM.

PCR analyses were performed with primers 101 and 102, which flank the DM repeat. PCR products were separated on a 10% polyacrylamide gel and visualised by ethidium bromide staining on an UV transilluminator. Automatic fragment length analysis of 5Cy5′ fluororescently labelled PCR products was performed on ALF (ALF Express™ Personal DNA Sequencer, Pharmacia Biotech) using commercially available external size markers (Pharmacia Biotech) and an internal length standard (PCR product of the probe pSB1.4 containing five CTG repeats). Fragment sizes were calculated with a fragment manager software package (Pharmacia Biotech). For Southern blot analyses we used EcoRI, BgII, BamHI, and radiolabelled probe pSB1.4. Paternity was tested by fingerprinting.

CTG repeat numbers are shown in the pedigree (fig 1A). CTG repeat sizes were confirmed by automatic analysis of the PCR products, except for the repeat number size of the grandmother (I.1). By ALF analysis, the grandmother had 11 and 12 CTG copies. All family members of generation II had one allele with 12 CTG repeats and another allele with different CTG copy numbers: 20 × CTG (II.1), 39 × CTG (II.2), and 37 × CTG (II.3). The mother (II.4) of the child III.2 had two different common alleles: 5 × CTG and 13 × CTG. The child (III.2) had 5 and 48 CTG repeats (fig 1A).

To the best of our knowledge this is the first description of unstable transmission of CTG repeats to all unaffected sibs from the same parents. All members of generation II had more than 19 CTG repeats on this second allele. Additionally, we showed an increase of CTG copies in the third generation (III.2) at the same time. The unstable paternal transmission from father (II.3) to son (III.2) resulted in a maximum increase of 11 CTG copies. This CTG increase is distinctly smaller than usually seen in families with DM heterozygotes.

Large CTG repeats (>42 CTG repeats) are predisposed to instability and the grandchild III.2 is in this range. This fact is very important for genetic counselling. Whether further repeat expansion will occur on transmission to the next generation, followed by the expression of the DM phenotype, remains to be seen. We are currently trying to identify additional family members such as sibs and cousins of I.2 from this pedigree. Longitudinal clinical and molecular studies of probands with critical CTG repeat numbers will be required.

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Figure 1 (A) Family pedigree. Numbers of CTG repeats on both alleles are shown. DNA from subjects I.2 and III.1 was not investigated. (B) Restriction fragment length patterns of high resolution Southern blot analysis with endonuclease BamHI detected with DNA probe pSB1.4 in the family members. BamHI fragments were not separated on the same blot but they are of comparable resolution indicated by the PCR products.


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