Evaluation of molecular genetic diagnosis in the management of familial adenomatous polyposis coli: a population based study


Abstract
A population based clinical and molecular genetic study of familial adenomatous polyposis coli (FAPC) was performed to investigate the value of molecular genetic analysis and ophthalmological assessment in the presymptomatic diagnosis of FAPC. The point prevalence of affected patients was $2.62 \times 10^{-3}$ (1/38000) and the minimum heterozygote prevalence was estimated at $3.8 \times 10^{-3}$ (1/26000). Eight of 33 (24%) probands were new mutations. Forty-eight asymptomatic relatives at 50% prior risk aged between 10 and 40 years were assessed for risk modification with linked DNA markers: in nine subjects (18%) the family structure was unsuitable for linkage based analysis, but 32 subjects were informative with a panel of intragenic and closely linked markers (25 had a combined age/DNA related risk of $<1\%$ (low risk group) and seven were at high risk (DNA predicted risk $>99\%$)). Ophthalmological assessment for CHRPEs showed that 27/43 (63%) affected patients and high risk relatives and 0/18 low risk relatives had more than three CHRPEs. Interfamilial variation in CHRPE expression was apparent. This study has shown that DNA based risk modification with intragenic and closely linked DNA markers is informative in most FAPC families. In addition to the clinical benefits of presymptomatic diagnosis for FAPC, the reduction in screening for low risk relatives (365 person years in the present study) means that molecular genetic diagnosis of FAPC is a cost effective procedure.

Methods
PATIENTS
Affected patients and at risk relatives were ascertained by contacting gastroenterologists, surgeons, and general practitioners in East Anglia (population 2 058 951). All affected patients had $>100$ polyps. At risk relatives were offered screening (by annual sigmoidoscopy or three yearly colonoscopy) starting between 11 and 15 years of age and continuing to the age of 40 years. Ophthalmological assessment for CHRPE was performed by a single ophthalmologist. At risk subjects aged between 11 and 40 years were offered DNA based risk modification when the family structure was suitable for genetic linkage analysis.

Molecular Genetic Analysis
Blood was collected from relevant family members and genomic DNA prepared from peripheral blood lymphocytes by standard techniques. Five polymorphic DNA markers were investigated: (1) a $3'\ Ssp1$ polymorphism (DP2.5) in the APC gene, $^{16}$ (2) an exon 11 RsaI polymorphism (APC-E11) in the APC gene, $^{11}$ (3) a microsatellite polymorphism (L5.71) at the MCC locus (Jones and Nakamura, in preparation), (4) a $Msp1$ RFLP at D5S81 (YN5.48), $^{12}$ and (5) a $BglII$ RFLP at D5S98 (ECB27) $^{13}$ DNA was digested with the appropriate restriction enzyme according to the

Familial adenomatous polyposis coli (FAPC) is a dominantly inherited disorder characterised by the development of numerous (at least 100) colorectal polyps. Age at onset is variable, but polyps usually develop in teenage years and penetrance is almost complete at the age of 40 years. $^{1}$ Colorectal carcinoma develops inevitably unless prophylactic colectomy is performed. Polyps may also occur in the upper gastrointestinal tract, and although the risk of malignant transformation is probably lower than for colonic adenomas, duodenal carcinoma is now emerging as an important cause of death in patients with FAPC. $^{7}$ Extraintestinal manifestations of FAPC include epidermoid cysts, osteomas, desmoid tumours, and, most frequently, congenital hypertrophy of the retinal pigment epithelium (CHRPE).

FAPC is caused by mutations in the APC gene in chromosome 5q21-q22. $^{5,6}$ Presymptomatic bowel screening of at risk relatives considerably reduces morbidity and mortality from FAPC, $^{67}$ and the efficiency of screening can be enhanced by reliable presymptomatic diagnosis of FAPC gene carriers with linked DNA markers and ophthalmological assessment for CHRPEs. $^{6,8}$ However, presymptomatic detection of FAPC gene carriers is limited by (1) significant recombination between the DNA markers in general use, and (2) uncertainty about the sensitivity and accuracy of CHRPEs. The isolation and characterisation of the APC gene $^{6,5}$ should lead to more accurate presymptomatic diagnosis by the identification of new intragenic and very closely linked DNA markers. We report a population based study of the clinical and molecular genetics of FAPC.
Summary of the results for presymptomatic diagnosis for FAPC in East Anglia. Full details are given in the text.

16 aged <10 years and not studied

9 family structure unsuitable for linkage analysis

64 relatives at 50% risk aged <40 years

39 relatives at 50% risk aged 10-40 years investigated with linked DNA markers

4 results not yet available or declined presymptomatic test

25 LOW RISK (combined risk <1%)

7 HIGH RISK (DNA risk >99%)

3 RISK NOT MODIFIED

Manufacturers' recommendations. For ECB27 and YN5.48 agarose gel electrophoresis Southern transfer, and hybridisation with 32P labelled probe was performed as described previously. For two intragenic markers (APC-exon 11 RsaI and DP2.5) and the CA repeat polymorphism in L5.71, DNA was amplified by the polymerase chain reaction under the appropriate conditions (Jones and Nakamura, in preparation) and the alleles resolved in an agarose gel for the intragenic markers and on a 6% polyacrylamide/6 M urea sequencing gel for L5.71. Families were initially investigated with the two intragenic polymorphisms, and then with the highly informative dinucleotide repeat L5.71 which maps approximately 300 kb centromeric to the APC gene. Uninformative families were then studied with YN5.48 and ECB27. The locus order is: ECB27-L5.71-APC-YN5.48. DNA related risks were calculated using the MLINK program with the following recombination fractions: APC exon 11 RsaI 0 = 0.001, DP2.5 0 = 0.001, L5.71 0 = 0.005, YN5.48 0 = 0.02, and ECB27 0 = 0.03. A final carrier risk was derived by a Bayesian calculation combining the age related and DNA based risks. The age related risk was based on the age at last sigmoidoscopy/colonoscopy and the published age dependent penetrance classes.

Results

**CLINICAL GENETIC ANALYSIS**

At the start of the study there were 54 affected patients and 64 relatives aged <40 years at 50% prior risk (that is, had an affected parent) from 33 kindreds living in East Anglia. The crude prevalence of FAPC patients in East Anglia was 2.62 × 10^-3 (1/38 000). By estimating the number of gene carriers among asymptomatic at risk relatives and the prevalence of asymptomatic new mutation FAPC patients aged <20 years, it was calculated that the heterozygote prevalence in East Anglia is at least 3.8 × 10^-3 (1/26 000). Eight of 54 affected patients represented new mutations.

Ophthalmological assessment was performed in 39 affected patients and 22 at risk relatives informative for DNA analysis (see below) living in East Anglia. There were 30/39 (77%) affected patients and 7/22 (32%) at risk relatives with one or more CHRPEs, and 24/39 (57%) and 3/22 (14%) respectively had four or more CHRPEs. Of the 22 relatives informative for DNA analysis who underwent ophthalmological assessment, 18 were found to be at low risk (combined age and DNA related risk of <1%) and four were high risk (99%). The frequency of CHRPEs varied between families. In the 14 families in which at least one affected patient had more than three CHRPEs 27/29 (93%) affected patients/high risk relatives and 0/12 low risk relatives had more than three CHRPEs. In the nine families in which none of 13 affected patients had more than three CHRPEs, none of the at risk relatives (one high, six low risk) had more than three CHRPEs.

**CYTOGENETIC ANALYSIS**

High resolution cytogenetic analysis was performed in four mentally retarded FAPC patients. Chromosomes were normal in three patients, but one affected patient with moderate mental retardation, minor dysmorphic features, and multiple (>10) CHRPEs showed an apparently balanced complex translocation: 46,XY,t(5;12)(16)pter-q22::12q22-12qter;12pter-q22::5q32-5q22::16q12-16qter;16pter-q22::5q32-5qter,t(9;1)(q33;q14). Both parents had normal chromosomes and phenotype.

**GENETIC LINKAGE ANALYSIS**

Twenty-two of 26 kindreds had a suitable structure for linkage based risk modification. These kindreds contained 39 subjects at 50% prior risk aged between 10 and 40 years and 38 of these requested investigation for risk modification by linked DNA markers. After molecular investigation with the protocol described previously, 32 relatives were informative, three are still being studied, and DNA testing did not significantly alter the risk of three subjects (figure). The three uninformative subjects (S1, T1, and T2) were from two families in which the family structure was not optimal for linkage based analysis. In the 32 informative relatives, all those at low risk had a combined age/DNA related risk of <1% and all high risk subjects had a DNA predicted risk of >99%. The results of DNA marker analysis for individual relatives are shown in the table. The dinucleotide (CA) repeat polymorphism in L5.71 was the most informative, with 77% (17/22) of subjects investigated being informative. The two intragenic polymorphisms (exon 11 RsaI and DP2.5) were informative in 55% (18/33) and 46% (13/28) of subjects, but the two polymorphisms were in linkage disequilibrium (observed haplotype frequencies in 59 individual chromosomes: 1/1 = 0/59, 1/2 = 23/59, 2/1 = 31/59, 2/2 = 5/59, χ^2 = 38.4, p < 0.0001), which reduced the overall informativeness of these two markers.

No evidence of FAPC has been detected in
the 25 low risk subjects; 18 of these have undergone ophthalmological assessment with negative findings (fewer than four CHRPEs) (see above). Four high risk subjects have been investigated by sigmoidoscopy or colonoscopy, two had no polyps at the ages of 14 and 16, and polyps were detected in two aged 18 and 27 years.

Discussion

EPIDEMIOLOGY

The ascertainment of affected patients (2.62 \times 10^{-5}) in East Anglia compares favourably to that in north-east England (2.29 \times 10^{-5}),\textsuperscript{8} Finland,\textsuperscript{7} Denmark (2.6 \times 10^{-5}),\textsuperscript{16} and Western Australia (2.59 \times 10^{-5}).\textsuperscript{15} The prevalence of FAPC appears to be increasing because of better case ascertainment and increased survival as a result of more patients being diagnosed early through presymptomatic screening programmes.\textsuperscript{8} We estimated a minimum heterozygote prevalence of 1/26000 (3.8 \times 10^{-5}). Our estimate that 24% of probes represented new mutations is similar to that at the Cleveland Clinic (22%),\textsuperscript{18} but less than the 46% of probes in the Netherlands reported by Vasen et al.\textsuperscript{6} Although it has been suggested that new mutations may be associated with a more severe phenotype,\textsuperscript{18} a meaningful comparison is difficult because isolated cases will be diagnosed later.

CARRIER DETECTION

Although four patients with FAPC and cytogenetically visible chromosome 5q deletions have been reported,\textsuperscript{19-22} to our knowledge we have reported the first FAPC patient with a reciprocal translocation apparently involving the APC gene. Although chromosomal aberrations are rare in FAPC, cytogenetic analysis should be performed in all mentally retarded FAPC patients and those with a family history of mental handicap. Only a minority of such patients will prove to have a chromosomal rearrangement, but the benefits of identifying such patients are considerable.

Most reports of DNA based presymptomatic diagnosis in FAPC are of selected families,\textsuperscript{15,23,24} but such studies do not accurately reflect the range of families seen in routine clinical practice. Population based studies are more representative, but MacDonald et al\textsuperscript{20} found that only 46% of at risk relatives in the English West Midlands had a suitable family structure for genetic linkage analysis, only 12% of subjects tested were informative for flanking markers, and the recombination fractions for the DNA markers studied ranged from 3 to 10%. We used recently identified intragenic and closely linked DNA markers and found that in those families with a suitable structure for linkage based analysis, the status of most at risk relatives can be established accurately. However, 18% (9/48) of at risk relatives were from families in which the family structure was unsuitable for this type of analysis or in which DNA from key family members was not available. Presymptomatic diagnosis by direct mutation detection may be applicable in such families and we have characterised the APC gene mutations in 15/24 East Anglian kindreds analysed.\textsuperscript{25} However, the
spectrum of APC gene mutations is extremely heterogeneous and although some clustering of APC gene mutations exists (for example, at codons 1309 and 1061),25-27 the complete detection of APC gene mutations would represent a formidable amount of work. The best strategy for the molecular genetic investigation of FAPC is not yet established but we suspect that genetic linkage analysis will be the initial investigation in families with a suitable structure. The observation that most APC gene mutations result in a truncated protein suggests that the most promising diagnostic approach might be the use of antibodies against the APC gene product.

The presence of multiple CHRPEs can be used as a biomarker for the presymptomatic diagnosis of FAPC. The specificity of four or more CHRPEs for FAPC appears to be high. Burn et al. found that 0/04 normal subjects and we found 0/18 low risk relatives had more than three CHRPEs. However, Cachon-Gonzalez et al. reported a single person with five CHRPEs who was at low risk with flanking DNA markers (PI227 and YN5.48). CHRPE analysis provides a method of independently confirming the results of molecular genetic analysis, but further studies are needed to determine the molecular basis for interfamilial differences in the frequency of CHRPEs.

COST–BENEFIT ANALYSIS
We have not attempted to perform a detailed cost–benefit analysis of our programme (such an analysis would have to quantify the benefit of reassuring at risk relatives as well as the prevention of colorectal cancers, etc); however, it is possible to estimate the reduction in screening produced by identifying low risk subjects. We estimate that by discontinuing screening in the 25 low risk relatives we will screen 365 person years of unnecessary screening. The cost of a single colonoscopy has been estimated as £107,28 so for a screening programme based on three yearly colonoscopy (the most frequently used protocol in East Anglia), a saving of 365 person years translates into a future financial saving of approximately £13 000 at current costing.

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