Linkage analysis in adenomatous polyposis coli: the use of four closely linked DNA probes in 20 UK families

M B Cachon-Gonzalez, J D A Delhanty, J Burn, K Tsioupra, M B Davis, J Attwood, P Chapman

Abstract

Linkage analysis was carried out on 20 unselected UK families segregating for adenomatous polyposis coli (APC) using four closely linked DNA probes. Significant lod scores were obtained between APC and three markers: \( \pi_{227} (DSS37) \tilde{\theta} = 0.16; C11p11 (DSS71) \tilde{\theta} = 0.10; \) and YN5.48 (DSS81) \( \tilde{\theta} = 0.00. \) The fourth, ECB27 (DSS98), gave low lod scores. The APC gene showed linkage with at least one of the probes used in all families, which is in agreement with previous publications. Combined lod scores are now sufficiently high to allow the use of these probes in presymptomatic diagnosis. Despite the fact that 61% of persons at risk were informative for at least one DNA marker, only 15% were informative with flanking probes. One prenatal diagnosis was performed where the initial request had been for sterilisation.

Adenomatous polyposis coli (APC), also known as familial adenomatous polyposis, is a relatively common autosomal dominant condition. By the middle of the third decade of life most affected subjects have developed about a thousand adenomatous polyps in the colon and rectum; one or more of these will inevitably progress to malignancy unless prophylactic colectomy is carried out. Extracolonic lesions include upper gastrointestinal tract adenomas, osteomas, sebaceous cysts, desmoid tumours, and, the important phenotypic marker, congenital hypertrophy of the retinal pigment epithelium (CHRPE). Colonic polyposis with the addition of the extracolonic lesions was formerly considered to constitute a distinct syndrome first described by Gardner.

The gene for APC has been mapped to chromosome 5, region q21-22, by close linkage to an anonymous DNA marker, C11p11 (DSS71).4 Linkage to the more informative probe, \( \pi_{227} (DSS37) \) was subsequently reported and additional closely linked markers have since been isolated, the most important of which are ECB27 (DSS98), YN5.48 (DSS81), and, more recently, L5.62 and EF5.44. The evidence now available from genetic linkage studies suggests that Gardner’s syndrome maps to the same region of chromosome 5 as APC and prevailing opinion is that the two cannot be clinically distinguished.

Linkage analysis with the probes C11p11, \( \pi_{227} \), ECB27, YN5.48, EF5.44, and L5.62 has been reported in nine Scottish families by Dunlop et al. Recombinants were identified and together with other published data allowed a locus order of centromere-\( \pi_{227}-C11p11-ECB27-L5.62-APC-EF5.44-YN5.48-telomere \) to be proposed. In their study no recombinants were found between the pairs of loci L5.62-ECB27, ECB27-C11p11, C11p11-\( \pi_{227} \), EF5.44-L5.62, \( APC-YN5.48 \), and \( APC-EF5.44 \). However, analysis of an informative Dutch pedigree showed that \( \pi_{227} \) and YN5.48 closely flank the APC locus. It was suggested that this finding would allow prenatal and presymptomatic diagnosis of APC in the majority of families with more than 99-9% reliability.

We report here our results of linkage analysis in 20 UK families using the four closely linked DNA markers, \( \pi_{227}, C11p11, ECB27, \) and YN5.48.
Materials and methods

The 20 families studied were from the Northern Region Polyposis Registry which was established in 1987. In these families, subjects with polyposis were identified from surgical records and the family members at risk were given periodic clinical examinations of the colon together with indirect ophthalmoscopy to look for the presence of CHRPEs, as described. To prevent the inclusion of false positives, bilateral CHRPEs with a total of eight or more typical lesions were considered to be definitively indicative of gene carrier status; where less than eight lesions were present the status was taken as unknown. Only families with at least one affected member expressing the CHRPE phenotype were scored in this way.

Where no information with regard to APC or CHRPE status was available an age dependent penetrance classification (described for the Dutch population\(^4\)) was used, as follows: 6-0% for ages 0 to 12, 32-5% for 13 to 17, 66-5% for 18 to 22, 89% for 23 to 27 years, and 98% for ages 28 and over. No distinction was made between families with respect to extracolonic features apart from CHRPEs.

High molecular weight DNA for restriction analysis was prepared from fresh or frozen blood by standard methods.\(^\text{13}\) Probe DNA clones used were as follows: p227 (DSS37)\(^8\) as a 900 bp HindIII/EcoRI fragment in pAN7; C11p11 (DSS71)\(^4\) as a 3-6 kb EcoRI fragment in pUC8; ECB27 (DSS98)\(^6\) as a 2-8 kb SalI fragment in phage lambda; and YN5.48 (DSS81)\(^8\) as a 2-4 kb TaqI fragment cloned into the AccI site of pUC18.

Purified DNA (5 to 10 μg) was digested with the appropriate restriction endonuclease (TaqI for the probe C11p11, BrXI, BclI, PstI, and MboI for p227, BglII for ECB27, and MspI for YN5.48), fractionated in 0-6 to 1-5% agarose gel, and transferred to Gene Screen Plus nylon filters (Dupont) as recommended by the manufacturers. Before prehybridisation the DNA was cross linked to the membrane by exposing it to 254 nm UV light for three minutes. Prehybridisation was carried out in 0-5% SDS, 10% dextran sulphate, and 0-8 mol/l NaCl for 15 minutes to several hours at 65°C. Probes were radiolabelled with α-\(^32\)P-dCTP (3000 Ci mmol\(^{-1}\)) by the random hexanucleotide primer method\(^\text{15}\) to a high specific activity, and hybridised to the filter at 65°C overnight in the presence of 10 μg ml\(^{-1}\) salmon sperm DNA. Filters were washed to a stringency of 2 × SSC, 1% SDS and autoradiographed at −70°C using Fuji RX-L x ray film.

Linkage analysis was carried out using the LINKAGE group of programs,\(^\text{16}\) within the easier to use computer program LINKSYS.\(^\text{17}\) In family groups where recombination was apparent, paternity was checked with the use of at least three hypervariable DNA probes.\(^\text{18}\) Subjects for whom non-paternity was shown were excluded from the analysis.

Results

Twenty families segregating for the APC gene, comprising 180 typed subjects with 91 meioses, were screened for all the polymorphisms. Allele frequencies were determined in the unaffected spouses and are shown in table 1.

The results of the linkage analysis of APC to each of the four DNA markers, and of these to each other, are summarised in table 2. The peak lod score (\(Z_{\text{max}}\)) and associated recombination fraction (\(\theta\)), indicating the maximum likelihood of marker locations with respect to each other, was calculated using

### Table 1 DNA probes recognising polymorphic loci on chromosome 5q: allele size, frequency in the spouse population, and number of informative families

<table>
<thead>
<tr>
<th>Probe</th>
<th>Enzyme recognising polymorphism</th>
<th>Allele</th>
<th>Size (kb)</th>
<th>Frequency</th>
<th>No of chromosomes</th>
<th>Informative families (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>YN5.48(^8)</td>
<td>MspI</td>
<td>Y1</td>
<td>9-0</td>
<td>0.51</td>
<td>68</td>
<td>55</td>
</tr>
<tr>
<td>ECB.27(^7)</td>
<td>BglII</td>
<td>Y2</td>
<td>8-0</td>
<td>0.49</td>
<td></td>
<td>70</td>
</tr>
<tr>
<td>C11p11(^4)</td>
<td>TaqI</td>
<td>P1</td>
<td>11-9</td>
<td>0.59</td>
<td></td>
<td>65</td>
</tr>
<tr>
<td>p227(^14)</td>
<td>BclI</td>
<td>Z1</td>
<td>4-4</td>
<td>0.20</td>
<td></td>
<td>70</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Z2</td>
<td>3-9</td>
<td>0.80</td>
<td></td>
<td>50</td>
</tr>
<tr>
<td></td>
<td>BrXI</td>
<td>A1</td>
<td>8-0 + 12</td>
<td>0.16</td>
<td></td>
<td>70</td>
</tr>
<tr>
<td></td>
<td></td>
<td>A2</td>
<td>3-0</td>
<td>0.40</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>A3</td>
<td>1-8 + 1-2</td>
<td>0.44</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>PsI</td>
<td>B1</td>
<td>2-7</td>
<td>0.43</td>
<td>68</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>B2</td>
<td>2-3</td>
<td>0.57</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>C1</td>
<td>4-3</td>
<td>0.26</td>
<td>68</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>C2</td>
<td>3-0</td>
<td>0.74</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>MboI</td>
<td>D1</td>
<td>0-55</td>
<td>0-41</td>
<td>66</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>D2</td>
<td>0-45</td>
<td>0-59</td>
<td></td>
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</tbody>
</table>
the LODSCORE program of LINKAGE. Lod scores for each pair of markers at different recombination fractions were calculated using the MLINK program of LINKAGE. The 1 lod unit of support was also determined, as indicated by Conneally et al.19 Only the BclI polymorphism with π227 was used in these pairwise lod score calculations since it is the most informative. As expected, given the small lod scores obtained with the sexes pooled, calculating lod scores with the sexes separated did not give any meaningful information (data not shown).

Several useful recombinants have been detected during the course of this study. One of these is subject IV.3 in pedigree 3708 (fig 1) who is the result of a single recombination event separating ECB27 and π227 from the APC gene and YN5.48; the C11p11 was not informative in this case. This indicates that loci recognised by ECB27 and π227 are situated on the same side of the APC gene. Another recombination event, in a different family, was observed between π227 and ECB27-APC (C11p11 and YN5.48 were uninformative) providing evidence that ECB27 is closer to APC than is π227. An obligate recombinant in a third pedigree between APC and π227, together with C11p11, was also detected, placing these two markers on the same side of the gene. No recombinants between the pairs of markers C11p11-ECB27 and APC-YN5.48 have been found in these families.

Significant lod scores have been obtained between APC and π227, C11p11 and YN5.48, and also between the pairs of markers π227-C11p11, π227-ECB27, and C11p11-ECB27. These data and the information obtained from the recombinants allow us to place the markers π227, C11p11, and ECB27 close together on the same side of the APC gene, which is in agreement with the findings of others.9 Also in agreement is the evidence we have in favour of YN5.48 being very close to APC.

Discussion

This linkage study has used data provided from 20 unselected families ascertained since the setting up of the Northern Region Polyposis Registry in 1987. The identification of a number of recombinants together with the linkage data we have obtained enables our results to be combined with those already published to give an estimate of genetic distance for the markers used and a gene order. The combination of our own and published results for the approximate maximum lod score (Zmax) and associated recombination fractions (θ) is as follows: C11p11-APC Zmax = 15.33, θ = 0.09±0.03; π227-APC Zmax = 19.57, θ = 0.10±0.03;

* Lod score is too small to calculate the confidence limits.

Table 2: Pairwise lod scores for linkage

<table>
<thead>
<tr>
<th>Marker Pair</th>
<th>Zmax</th>
<th>95% confidence limits</th>
<th>θ</th>
</tr>
</thead>
<tbody>
<tr>
<td>APC/π227</td>
<td>2.24</td>
<td>0.16</td>
<td>-∞</td>
</tr>
<tr>
<td>APC/C11p11</td>
<td>2.33</td>
<td>0.10</td>
<td>-1.02</td>
</tr>
<tr>
<td>APC/YN5.48</td>
<td>1.15</td>
<td>0.17</td>
<td>-8.86</td>
</tr>
<tr>
<td>π227/C11p11</td>
<td>4.67</td>
<td>0.00</td>
<td>-6.16</td>
</tr>
<tr>
<td>π227/YN5.48</td>
<td>6.03</td>
<td>0.00</td>
<td>-6.04</td>
</tr>
<tr>
<td>ECB27/YN5.48</td>
<td>2.49</td>
<td>0.12</td>
<td>-3.72</td>
</tr>
<tr>
<td>ECB27/C11p11</td>
<td>3.71</td>
<td>0.00</td>
<td>-3.70</td>
</tr>
<tr>
<td>ECB27</td>
<td>0.27</td>
<td>0.18</td>
<td>-3.08</td>
</tr>
</tbody>
</table>

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ECB27-APC $Z_{\text{max}} = 8.98$, $\hat{\theta} = 0.10^{+9}$; YN5.48-APC $Z_{\text{max}} = 22.57$, $\hat{\theta} = 0.05$. Of note is the fact that the combined approximate maximum lod score for linkage of ECB27 and APC indicates that this marker is probably not as close to the gene as was originally thought. Taking the gene mapping data together with information from constitutional and somatic deletions, the most likely locus order is $\pi 227-C_{11} p11-ECB27-APC-YN5.48$.

Although we chose a cut off point of a total of eight or more CHRPE lesions as being indicative of gene carrier status for the purpose of this linkage analysis, in clinical practice a level of six or more is used in these families as a positive indicator. At risk subjects with three to five lesions can present a problem, as illustrated by fig 2. Based on DNA typing, the results with probes YN5.48 and $n227$ would suggest that subject III.1 is unaffected, but she has four CHRPE lesions in one eye and a single lesion in the other eye. Published data puts YN5.48 telomeric to APC, necessitating a single recombinant if III.1 is affected. This event is not impossible since the distance between these markers could be as much as 17cM, based on a combination of CEPH data for YN5.48-$C_{11} p11^{14}$ and our own data for these two markers and for YN5.48-$n227$. Of course, in the, probably unlikely, scenario that YN5.48 and $n227$ are both centromeric to APC, only a single recombinant would be required. The remaining possibility is that she is not a gene carrier and that five CHRPEs represent the extreme of the normal range.

In all families studied the APC gene has shown linkage with at least one of the probes used. This is in agreement with all published reports that so far there is no evidence for genetic heterogeneity with respect to linkage with markers on chromosome 5. Lod scores are now sufficiently high to allow the use of these probes in presymptomatic diagnosis.

With regard to practicalities, it is also worth noting that despite the fact that 61% of the subjects at risk were informative for at least one DNA marker, only 11 out of 72 (15%) at risk subjects were in fact informative with flanking probes. This means that in most cases presymptomatic diagnosis can only be given with 90% probability at present, based on DNA typing alone. This level of risk proved acceptable to one couple requesting prenatal diagnosis where the initial request had been for sterilisation.

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17 Attwood J, Bryant S. A computer program to make linkage analysis with LIPED and LINKAGE easier to perform and less prone to input errors. Ann Hum Genet 1988;52:259.
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