Monozygotic twins with trisomy 18: a report of discordant phenotype

Jerrold S Schlessel, W Ted Brown, Andrzej Lysikiewicz, Russell Schiff, Ann Leslie Zaslav

Abstract

The predicted incidence of liveborn monozygotic trisomy 18 twins is one per million births. The first case of liveborn monozygotic trisomy 18 twins was reported in 1989 and we report a second case in which striking phenotypic discordance existed. The probability of monozygotic trisomy 18 twinning and the mechanisms for phenotypic discordance in trisomic twins is discussed.

Trisomy 18 occurs at a rate of about 0.3 per 1000 newborn babies with a 3:1 preponderance of females to males. The monzygotic twin rate is 3.5 to 4 per 1000 births. The predicted incidence of these events occurring together is approximately one per million. However, the true incidence may be much lower as there has only been one reported observation of trisomy 18 in liveborn monozygotic twins. We report the second case of liveborn monozygotic twins with trisomy 18. Despite identical karyotypes, discordance for major congenital malformations was present.

Case report

A 26 year old white woman, gravida 2, para 1-0-0-1, with a twin pregnancy, was admitted to hospital at 27 weeks' gestation because of premature rupture of membranes and preterm labour. Ultrasonographic examination at this time showed discordant fetal measurements: biparietal diameters 71 and 60 mm, and femur lengths 48 and 44 mm (values for twin A and twin B, respectively). Twin A, who had fetal measurements appropriate for gestational age, had a two vessel umbilical cord and an umbilical artery blood velocity systolic/diastolic ratio (S/D ratio)= 2.5-3. Twin B had a "lemon shaped appearance of the fetal head" (fig 1a) and a cystic spine defect at the level of the lumbar vertebrae consistent with a...
Monozygotic twins with trisomy 18: a report of discordant phenotype

myelomeningocele (fig 1b). Polyhydramnios and a raised S/D ratio of 3.5–4.7 were also noted.

The mother received betamethasone to induce fetal lung maturation and ritodrine for tocolysis to allow for 24 hours of steroid treatment. During the subsequent 24 hours, the patient broke through tocolysis and was delivered by caesarean section.

Twin A was a 923 g female who had no spontaneous respiratory effort. Resuscitation with positive pressure ventilation was instituted immediately after birth and Apgar scores were 7 and 9 at one and five minutes, respectively. Physical examination showed microphthalmia, grossly malformed right ear pinna with absent external auditory canal, right facial skin tag, micrognathia, overlapping of index over third finger, hypoplastic fingernails, a grade III/VI systolic ejection murmur with a single S2, and rocker bottom feet. An echocardiogram showed a hypoplastic right ventricle and a posterior endocardial type ventricular septal defect (fig 2). A small tricuspid valve annulus with tricuspid regurgitation, absent right ventricular outflow tract, pulmonary atresia, and patent ductus arteriosus were also noted. An infusion of prostaglandin E1 was started. A renal sonogram was within normal limits: right kidney length = 3.5 cm and left kidney = 3.1 cm. A bone marrow aspirate and blood sample were obtained four hours after birth for karyotype analysis using standard cytogenetic harvesting and banding techniques.5 6

Twin B was a 666 g female who also had no spontaneous respirations. Identical Apgar scores to those of twin A were assigned to twin B during positive pressure ventilation. Microphthalmia, a reducible omphalocele with herniation of intestine and overlying membrane, a 2 x 2 cm open, leaking, lumbosacral myelomeningocele, overlapping of index over third finger, hypoplastic nails, and rocker bottom feet were noted. Neurosonogram showed a moderate degree of hydrocephalus. A blood sample for karyotyping was drawn.

The diagnosis of trisomy 18 was suspected in both twins in view of the physical findings. The karyotype of twin A was identified at 22 hours of life as trisomy 18 by bone marrow aspirate culture. The karyotypes of twins A and B were trisomy 18 on blood lymphocyte culture. Blood obtained from each twin confirmed monosomy by identical DNA restriction fragment length polymorphisms for hypervariable loci D2S44, D17S79, D14S13, and D18S27 (performed by Lifecodes Inc, Elmsford, NY). The infants were mechanically ventilated and received intravenous hydration, packed red blood cell transfusions, and ampicillin and gentamycin. Twin A died at 36 hours and twin B at 48 hours of life.

Discussion

These infants are the second reported cases of liveborn monozygotic trisomy 18 twins. Growth retardation and structural malformations were detected in one of the twins by fetal ultrasonography, raising the spectre of chromosomal abnormalities. Since the mother was in preterm labour, further prenatal diagnosis was not considered. These twins were discordant for their major congenital malformations, complex congenital heart disease in twin A and

Figure 2. Echocardiogram of twin A showing hypoplastic right ventricle (RV). Arrows indicate RV wall; posterior endocardial ventricular septal defect (VSD); left ventricle (LV); right atrium (RA); left atrium (LA); and patent foramen ovale (PFÖ).
myelomeningocele, hydrocephalus, and omphalocele in twin B, even though concordance for genotype was established by DNA probes.

The first case of trisomy 18 in liveborn monozygotic twins differs from this report in that less striking phenotypic discordance was reported; omphalocele in twin A and clitoromegaly in twin B. Two cases of presumptive monozygotic trisomy 18 twins with only twin A liveborn have been reported. Neither study presented genetic evidence of monozygosity. In one report, one of the twins was viable and the other had major malformations including holocardius. In the other report, the liveborn twin died at 15 minutes of age.

In contrast to the apparent rarity of monozygotic trisomy 18 twinning, monozygotic twinning with trisomy 21 has been reported frequently and discussed extensively. In 1968, Zellweger reported 22 cases of monozygotic twins with Down’s syndrome. Nevertheless, the frequency of liveborn monozygotic twins with trisomy 21 is less than the calculated expectation. It has been suggested that this decrease in the expected frequency of monozygotic twins with trisomy 21 results from increased embryonic or fetal wastage. Fetal wastage has been estimated as 65% in trisomy 21 versus 80% in trisomy 18. This may explain, in part, the very rare occurrence of liveborn monozygotic twins with trisomy 18, as observed here and reported by Shah et al. Although the calculated probability of liveborn monozygotic twins with trisomy 18 is approximately one in one million, the rarity of observations suggest that there may be a further increase in fetal wastage in trisomy 18 in the setting of twins.

The twins in this report were observed to be discordant for phenotypic features as seen in differing major congenital malformations. Two explanations exist for congenital malformation in twins: (1) the germ cell theory, which assumes the defect to be present in the gametes, and (2) the environmental theory, which assumes the defects occur after fertilisation. Since the twins in this report were monozygotic and, therefore, of similar genetic constitution, it seems most likely that intrauterine environmental effects caused the differences in their malformations. Loevy et al noted discordant anomalies in monozygotic twins with trisomy 13 and suggested that the presence of a chromosomal abnormality itself may predispose to a variable range of phenotypic anomalies.

This case of liveborn monozygotic twins with trisomy 18 is the second reported observation. We suggest that the increased fetal wastage seen in trisomy 18 acts to reduce the overall observed incidence of liveborn monozygotic twins with trisomy 18. The predicted incidence of one per million births, based on the probability of monozygotic twinning and trisomy 18 occurring together, may greatly overestimate the true incidence of monozygotic twins with trisomy 18.

Monozygotic twins with trisomy 18: a report of discordant phenotype.

J S Schlessel, W T Brown, A Lysikiewicz, R Schiff and A L Zaslav

*J Med Genet* 1990 27: 640-642
doi: 10.1136/jmg.27.10.640

Updated information and services can be found at:
http://jmg.bmj.com/content/27/10/640

*These include:*

**Email alerting service**

Receive free email alerts when new articles cite this article. Sign up in the box at the top right corner of the online article.

**Notes**

To request permissions go to:
http://group.bmj.com/group/rights-licensing/permissions

To order reprints go to:
http://journals.bmj.com/cgi/reprintform

To subscribe to BMJ go to:
http://group.bmj.com/subscribe/