Polymorphic acetylation of sulphamethazine in a Zimbabwe population

CHARLES F B NHACHI
From the Department of Clinical Pharmacology, Godfrey Huggins School of Medicine, University of Zimbabwe, Avondale, Harare, Zimbabwe.

SUMMARY Sulphamethazine, 8 mg/kg body weight, was administered orally in tablet form to 100 healthy volunteers and total and free sulphamethazine were determined in the six hour urine sample. The bimodal population frequency distribution for percentage acetylated sulphamethazine showed 42 of the tested population to be fast and 58 to be slow acetylators, that is, an estimation of $q=0.72\pm0.3$ as the frequency of the allele controlling slow acetylation. The study also revealed ample evidence that the assay of the drug in urine can be done in a significantly shorter time.

It has been established by a number of investigators that there is a genetic polymorphism in man for the acetylation of sulphamethazine.1-3 Subjects are generally classified as slow or rapid acetylators4-6 and the proportion of slow to rapid acetylators varies among different peoples,7 but still exhibits bimodality even among regional groupings of a population.8 9

The present study was carried out to establish the existence of genetic polymorphism in a Zimbabwean population. The study also presents evidence that the determination of the acetylator phenotype using the method of Bratton and Marshall as given by Varley10 can be achieved, even when the incubation time is reduced to 40 minutes and using a very small dose of sulphamethazine.

Subjects, materials, and methods

One hundred healthy volunteers (all native Zimbabweans) were employed in the study. The subjects, both male and female, were between the ages of 20 and 38 years and were drawn from the Medical School students, technicians, and hospital nurses.

Six hour urine samples were collected and the volume measured. Total and free sulphamethazine concentrations were determined, using the method of Varley,10 on the day of collection. The percentage of sulphamethazine acetylated was determined using the formula

$$\frac{T-F}{T} \times 100\%$$

where $T=$total sulphamethazine excreted after six hours, and $F=$free (unacetylated) sulphamethazine excreted after six hours.

The percentage of sulphamethazine was also determined at different periods of acid hydrolysis in a boiling water bath for 10 subjects, who had been classified as slow (4) and fast (6) acetylators.

Results

The distribution of slow and fast acetylators of sulphamethazine in urine six hours after administration are shown in the figure and table 1.

Received for publication 8 July 1986.
Revised version accepted for publication 14 November 1986.
TABLE 1 Distribution of the acetylator phenotype among 100 subjects (70 men, 30 women).

<table>
<thead>
<tr>
<th></th>
<th>Fast</th>
<th>Slow</th>
</tr>
</thead>
<tbody>
<tr>
<td>% acetylators</td>
<td>42</td>
<td>58</td>
</tr>
<tr>
<td>% women acetylators</td>
<td>8</td>
<td>22</td>
</tr>
<tr>
<td>% men acetylators</td>
<td>34</td>
<td>36</td>
</tr>
</tbody>
</table>

With one degree of freedom and \( \alpha=0.05 \), \( \chi^2=3.841 \), there is no significant difference in phenotype distribution between the sexes.

TABLE 2 Percentage of urinary sulphanmethazine acetylated as determined after different periods of acid hydrolysis of urinary acetyl sulphanmethazine.

<table>
<thead>
<tr>
<th>Subjects</th>
<th>Minutes</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>5</td>
</tr>
<tr>
<td>A</td>
<td>4</td>
</tr>
<tr>
<td>B</td>
<td>86</td>
</tr>
<tr>
<td>C</td>
<td>33</td>
</tr>
<tr>
<td>D</td>
<td>10</td>
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<td>E</td>
<td>70</td>
</tr>
<tr>
<td>F</td>
<td>60</td>
</tr>
<tr>
<td>G</td>
<td>37</td>
</tr>
<tr>
<td>H</td>
<td>9</td>
</tr>
<tr>
<td>I</td>
<td>39</td>
</tr>
<tr>
<td>J</td>
<td>40</td>
</tr>
</tbody>
</table>

satisfactory separation (not completely resolved) of fast and slow acetylator phenotypes was obtained. (The phenotype is classified as slow if the percentage of acetylated sulphanmethazine is less than 70% and fast if more than 70%).

As has been shown in previous studies, the urinary elimination of total sulphanmethazine was significantly faster in fast acetylators (mean 5.1, 1.7 SEM) than in slow acetylators (mean 3.2, 1.6 SEM).

Table 2 shows that in the majority of cases and for practical purposes (qualitatively), the apparent percentage of urinary sulphanmethazine acetylated changes very little, if at all, after 40 minutes of acid hydrolysis in a boiling water bath.

Discussion

Although the existence of a genetic polymorphism in man in general, and in an African population in particular, for the acetylation of sulphanmethazine is well known, this study is the first to be done in Zimbabwe.

That the distribution of acetylation of sulphanmethazine exhibits bimodality is confirmed by the results of the present study. However, as can be seen in the figure, there is an overlap. The six hour urine sample has been shown in other studies to give efficient discrimination between slow and fast acetylators. Analysis of a six hour plasma sample for percent acetylation would have given a better resolution between phenotypes.

The method for determining the total amount of sulphanmethazine excreted in urine (based on the Bratton Marshall method as given by Varley) involves breaking down the acetyl derivative before the colour reaction. This is achieved by subjecting the urine sample to acid hydrolysis, that is, adding hydrochloric acid and heating the sample in a boiling water bath for one hour. Although this method was used in the present study, it was shown that 40 minutes of hydrolysis is enough to give an efficient discrimination between slow and rapid acetylators, as indicated in table 2.

In remote centres of most developing countries, where spectrophotometry is not automated, saving 20 minutes on every batch of assays (not more than 10 samples per batch) is a very important factor indeed.

Determination of the acetylator phenotype in a Zimbabwean population was a worthwhile exercise in view of the fact that slow acetylators of certain drugs, for example, isoniazid, are more susceptible to developing peripheral neuropathy than rapid acetylators of the drug.

Among other things, the present study provides evidence for the determination of the acetylator phenotype using a very low dose of sulphanmethazine (8 mg/kg) and the feasibility of carrying out the assay in a shorter time. Also, the frequency of the allele controlling slow acetylation, which is estimated at \( q=0.72 \pm 0.3 \), is similar to values which have been found in Kenya, Uganda, and northern Nigeria.

References

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Correspondence and requests for reprints to Dr C F B Nhachi, Department of Clinical Pharmacology, Godfrey Huggins School of Medicine, PO Box A178, Avondale, Harare, Zimbabwe.
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C F Nhachi

doi: 10.1136/jmg.25.1.29