Confirmation of Association Between ABO Blood Groups and Salivary ABH Secretor Phenotypes and Electrophoretic Patterns of Serum Alkaline Phosphatase

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Arfors, Beckman, and Lundin (1963), who had studied Swedish twin subjects, first pointed out that (1) serum alkaline phosphatases are present in either one or two electrophoretic bands which are under genetic control, (2) that their patterns are closely associated with the ABO blood groups in that the second electrophoretic alkaline phosphatase band is rarely found in adult subjects possessing an A1 or an A2 gene.

Beckman (1964) studied an extensive Brazilian family material comprising 468 adults and 839 children. It was found that all of them with an extra serum alkaline phosphatase band were secretors of ABH blood group substances. There was no correlation with secretion of Lewis substances. Only one adult of the 468 examined possessing an A1 or A2 gene showed two alkaline phosphatase bands; whereas over half the subjects of blood groups O and B showed this character.

Beckman (1964) concludes also that the family studies show that the appearance of the second electrophoretic alkaline phosphatase band is dependent on at least one additional factor besides the ABO and secretor genes.

The purpose of this communication is to confirm in the British population the associations described by Beckman (1964).

Materials and Method

Sera were from random British (white) blood transfusion volunteers.

Salivas were collected by having the subject spit into a test-tube. The quantity collected varied from 2 to 5 ml. Artificial stimuli to salivation were not employed. The specimens were boiled for 10 minutes on a water bath within an hour after collection, centrifuged at about 2,000 r.p.m. for 5 minutes, and the supernatants were stored at −20°C. when not in use.

Starch was the hydrolysed variety for gel electrophoresis from Connaught Medical Research Laboratories, University of Toronto, Canada.

Colouring agent. Fast Red TR salt (Gurr).

Electrophoresis. A gel was prepared by dissolving 242 g. starch in 200 ml. of Tris-HCl buffer 0·05 M, pH 8·8. A 19 cm. long horizontal gel was used, and during electrophoresis the gel and wicks were covered with parafilm. The current was 20 mA, voltage 260 V, and the time 3½ hours. Electrophoresis was conducted at +4°C. In the electrode compartments Tris-HCl buffer 0·30 M, pH 8·6 was used.

The sliced gel was stained for alkaline phosphatase by incubation for one hour at 37°C. with the following modification of the technique of A. L. Latner (personal communication, 1964):

10 mg. sodium naphthyl phosphate in 100 ml.
10 mg. fast red TR salt 0·1 M borate
20 mg. MgCl2 6 H2O 0·10 buffer pH 9·6.

During incubation the staining medium was renewed four times (to counteract the inhibition produced by the diazo-salt).

The second half of the sliced gel was stained with a saturated solution of Amido Black 10B in a mixture of methanol 200 ml., water 200 ml., glacial acetic acid 40 ml., to check upon the electrophoretic mobilities of serum protein bands.

ABO Blood Grouping. This was carried out by means of the standard tube techniques.

Determination of Salivary ABH Secretor Status. All salivas were tested for H, A, and B activity quantitatively by means of a doubling dilution agglutination-inhibition technique with 0·145 M sodium chloride as the diluent. Standard salivas from known O, A, and B secretors and non-secretors were included in each day’s estimations.
**Blood Groups, Secretor Phenotypes, and Serum Alkaline Phosphatase**

**TABLE**

<table>
<thead>
<tr>
<th>Blood Group</th>
<th>Salivary ABH Secretor Status</th>
<th>Serum Alkaline Phosphatase Bands</th>
<th>Two Distinct Bands</th>
</tr>
</thead>
<tbody>
<tr>
<td>O Secretor</td>
<td>4</td>
<td>6</td>
<td>43</td>
</tr>
<tr>
<td>O Non-secretor</td>
<td>12</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td>A Secretor</td>
<td>12</td>
<td>6</td>
<td>4</td>
</tr>
<tr>
<td>A Non-secretor</td>
<td>7</td>
<td>2</td>
<td>2</td>
</tr>
<tr>
<td>B Secretor</td>
<td>12</td>
<td>6</td>
<td>4</td>
</tr>
<tr>
<td>B Non-secretor</td>
<td>1</td>
<td>1</td>
<td>1</td>
</tr>
</tbody>
</table>

**Discussion**

The results are shown in the Table and they show that the observations of Beckman (1964) are confirmed on a random sample of the British population.

The site of origin of the main alkaline phosphatase band which occurs in all subjects is the liver. The second band which occurs in some adult subjects only is derived from the jejunal mucosa (Hodson, Latner, and Raine, 1962; Cunningham and Rimer, 1963; Haije and De Jong, 1963). Very rarely healthy adults have a third band derived from bone.

In children the second (jejunal) band is more marked and in young children the bone band is commonly evident.

The cause of the variation in the intensity of the second band observed in the present experiment is not known. It could be merely technical or a variability due to environmental or genetic causes.

The findings of Arfors et al. (1963) and Beckman (1964), which are confirmed here, are to be considered very important as they represent a pleiotropism (two or more characters which are controlled by a single gene) at a biochemical level.

**Summary**

This short communication confirms in the British population results found previously on Swedes and Brazilians. Attention is hereby drawn to the association existing between (1) electrophoretic serum and alkaline phosphatase bands and (2) the ABO blood group and salivary ABH secretor polymorphisms.

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