Linkage and association between HLA and 21-hydroxylase deficiency

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SUMMARY Congenital adrenal hyperplasia because of 21-hydroxylase deficiency is closely linked to the HLA system. The lod scores in 14 informative families are presented. Apart from linkage, the 21-hydroxylase deficiency is associated with an increase of BW47 antigen and lack of B8 antigen in patients. A family with a possible recombination between the 21-hydroxylase deficiency and the HLA complex was found, thus indicating that the 21-hydroxylase gene lies outside the HLA system and is closely linked to the HLA-DR locus.

Congenital adrenal hyperplasia (CAH) is an autosomal recessive disease which is most commonly caused by 21-hydroxylase (21-OH) deficiency. This deficiency results in decreased cortisol synthesis and secondary increased production of adrenocorticotrophic hormone (ACTH). In its turn, excess ACTH secretion leads to overproduction of adrenal androgens and consequent virilisation. The synthesis of aldosterone involves a 21-hydroxylase step and thus in some patients salt loss occurs in addition to virilisation. The reason why others are not salt losers and may have normal or high aldosterone levels is not well understood. The gene responsible for 21-OH deficiency has been shown to be closely linked to the HLA system, but it is not known whether this gene is structural or regulatory in type. We have confirmed this linkage. Since then, association between 21-OH deficiency and several HLA antigens has been reported. In the present report we describe the haplotypes of 14 families containing at least one affected child and one or more sibs. A further eight affected children have been HLA typed, making 22 unrelated 21-OH deficient children in all.

Patients

Fourteen families with both parents and either two affected sibs or one patient and at least one other sib, totalling 40 children, were HLA typed. Eight children with CAH whose complete families were not available were also HLA typed. Three families (No 12, 13, and 14) were from Pakistan and the rest were English. In two families the parents were first cousins (families 4 and 12) and in one family (family 14) the parents were more distantly related. The patients in families 2, 6, 9, and 11 were non-salt losers as were patients 16 and 18. The rest were salt losers.

HLA typing

HLA typing was performed by the standard NIH two stage micro-lymphocytotoxicity test. One hundred and twelve well defined sera were used to identify most of the HLA-A and -B locus antigens. Splits of HLA-A9 (W23, W24) or of B12 (W44, W45) were not tested for and the HLA-C locus antigens were identified in some subjects only. Monospecific antisera were not available for the BW47 antigen which was detected by a pattern of extra reactions in six antisera mainly for the B7 and B40 cross-reacting groups. Using these sera, BW47 could be detected with confidence in the presence of any other B locus antigen.

Biochemistry

Serum 17-hydroxyprogesterone levels were measured after ether extraction by radioimmunoassay (tritiated hormone supplied by the Radio Chemical Centre, Amersham, and antibodies by Steranti Research Ltd). Samples were taken at zero time and at 30 minutes after 250 mg synacthen had been given intravenously. Urinary 3β-pregnan-3α, 17α, 20α-triol levels were measured using a modification of the method of Bell and Varley. The normal range
for adult males is up to 6 nmol/24 h and the normal range for prepubertal boys aged 5 to 7 years is 0-6 to 1 nmol/24 h. The normal range at 30 minutes for normal prepubertal boys is less than 10 nmol/l (established locally n = 10).

Results

Table 1 shows the HLA-A and -B genotypes of 14 families with two or more children which were used for linkage analysis (table 2). The HLA phenotypes of a further eight children with CAH for whom complete families were not available or who were

### TABLE 1 HLA-A and -B genotypes in 14 families with CAH

<table>
<thead>
<tr>
<th>Family No</th>
<th>Parents</th>
<th>HLA haplotypes</th>
<th>Children's HLA haplotypes</th>
</tr>
</thead>
</table>
| 1         | F: A3, B7/A2, BW35  
M: A3, B14/A2, B5 | a/b | a/c b/c |
| 2         | F: A2, B17/A1, B15  
M: A2, B18/A9, B12 | a/b | a/c a/d b/c |
| 3         | F: A3, BW47/A2, B12  
M: A3, BW47/A7, B7 | a/b | a/c a/c* |
| 4 (C)     | F: A9, B7/A3, B12  
M: A9, B7/A2, B12 | a/b | a/c* b/c |
| 5         | F: A3, B40/A11, B7 | c/d | b/d a/c* a/c |
| 6         | F: A11, BW22/A1, B27  
M: A3, B40/A2, B17 | c/d | a/c* b/d |
| 7         | F: A2, BW35/A7, B8  
M: A1, B15/A1, B5 | c/d | b/c a/c* |
| 8         | F: A3, BW47/A1, B7 | a/b | a/c* b/c |
| 9         | F: A2, BW35/A29, BX  
M: A2, B12/A1, B8 | a/b | a/c* a/c |
| 10        | F: A1, BW47/AW32, B14  
M: A29, B12/A2, B15 | c/d | a/c* a/c |
| 11        | F: A26, B27/A1, B8  
M: A3, B7/A9, B12 | c/d | a/d a/c* |
| 12 (C)    | F: A1, B5/AW25, BW35  
M: A1, B5/A11, BW35 | a/c | b/c b/c/b/a |
| 13 (P)    | F: A9, BX/A11, B40  
M: A9, BW35/A1, B17 | a/b | a/c a/c* |
| 14 (C)    | F: A2, B12/A7, B15  
M: A2, B12/A11, B7 | c/d | a/c* a/c* |

C, consanguineous marriage;  
F, father;  
M, mother;  
*+, affected sib;  
+*, 21-OH deficiency homozygote, clinically 'normal';  
X, unknown antigen;  
T, possible homozygosity or unknown antigens;  
P, Pakistani families.

### TABLE 2 Sum of lod scores for linkage between HLA-B locus and 21-OH deficiency in 14 families

<table>
<thead>
<tr>
<th>Recombination fraction (g)</th>
<th>0.00</th>
<th>0.01</th>
<th>0.02</th>
<th>0.03</th>
<th>0.04</th>
<th>0.05</th>
<th>0.10</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sum of lod scores</td>
<td>0.00</td>
<td>8.625</td>
<td>8.796</td>
<td>8.826</td>
<td>8.811</td>
<td>8.753</td>
<td>8.286</td>
</tr>
</tbody>
</table>

Lod score calculated by using the Z_{1c} score and the C_{1c} corrections.12
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TABLE 4  Biochemical values in members of 2 families

<table>
<thead>
<tr>
<th>Family No</th>
<th>Sib No</th>
<th>Sex</th>
<th>Urinary pregnanetriol (umol/24 h)</th>
<th>Serum 17 hydroxyprogesterone after synacthen 250 µg iv injection 0' 30'</th>
</tr>
</thead>
<tbody>
<tr>
<td>2</td>
<td>5</td>
<td>M</td>
<td>55</td>
<td>70</td>
</tr>
<tr>
<td>8</td>
<td>1</td>
<td>M</td>
<td>0.6</td>
<td>6</td>
</tr>
<tr>
<td>8</td>
<td>2</td>
<td>M</td>
<td>2.1</td>
<td>7</td>
</tr>
</tbody>
</table>

TABLE 5  A Pakistani family (No 12) with an intra HLA recombination

<table>
<thead>
<tr>
<th>Father</th>
<th>HLA-A1, B5</th>
<th>AW25, B40, CW2</th>
<th>a/b</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mother</td>
<td>HLA-A1, B5</td>
<td>A11, BW35, CW4</td>
<td>a/c</td>
</tr>
<tr>
<td>Sib 1</td>
<td>HLA-AW25, B40, CW2</td>
<td>A11, BW35, CW4</td>
<td>b/c</td>
</tr>
<tr>
<td>Sib 2</td>
<td>HLA-AW25, B40, CW2</td>
<td>A11, BW35, CW4</td>
<td>b/c</td>
</tr>
<tr>
<td>Sib 3</td>
<td>HLA-AW25, B40, CW2</td>
<td>A1, B5</td>
<td>b/a</td>
</tr>
<tr>
<td>Sib 4</td>
<td>HLA-A1, B5</td>
<td>A1, BW35, CW4</td>
<td>a/c</td>
</tr>
<tr>
<td>Patient</td>
<td>HLA-A1, B5</td>
<td>A1, B5</td>
<td>a/a</td>
</tr>
</tbody>
</table>

The decrease of Fisher's exact (one-sided)-uncorrected probability for number of observations. NS, non-significant.

TABLE 6  HLA and chromosome 6 markers in family 8

<table>
<thead>
<tr>
<th>Biochemistry</th>
<th>HLA haplotypes</th>
<th>Bf</th>
<th>Glo</th>
<th>C2</th>
</tr>
</thead>
<tbody>
<tr>
<td>Father</td>
<td>NT</td>
<td>A3, BW47, DRW7/A1, B7, DRW2</td>
<td>a/b</td>
<td>F,S</td>
</tr>
<tr>
<td>Mother</td>
<td>NT</td>
<td>(A3), BW22, CW3, DRW6/(A3), BW47, DRW4</td>
<td>c/d</td>
<td>F,S</td>
</tr>
<tr>
<td>Brother</td>
<td>Normal</td>
<td>A3, BW47, DRW7/(A3), BW47, DRW4</td>
<td>a/d</td>
<td>F,F</td>
</tr>
<tr>
<td>Brother</td>
<td>Carrier</td>
<td>A3, BW47, DRW7/(A3), BW22, CW3, DRW6</td>
<td>a/c</td>
<td>F,S</td>
</tr>
<tr>
<td>Patient</td>
<td>21-OH deficiency</td>
<td>A3, BW47, DRW7/(A3), BW22, CW3, DRW6</td>
<td>a/c</td>
<td>F,S</td>
</tr>
</tbody>
</table>

NT, not tested;
Bf, properdin factor B;
Glo, glyoxalase;
C2, C2 component of complement.
significant before correction ($p = 0.002$). As mentioned earlier, in family 8 the patient has become BW47 positive as a result of an apparent paternal recombination. This patient has a normal brother who is a BW47 homozygote.

Discussion

Our study confirms close genetic linkage between the HLA system and 21-OH deficiency, as originally reported by Dupont et al. Both Dupont and we have described recombinants showing that the 21-OH gene segregates with the HLA-B locus. In the present report, we describe a further probable recombinant, this time between the HLA complex as a whole and 21-OH. The family in which the proband has severe salt losing congenital adrenal hyperplasia was HLA typed on three separate occasions with indetical results. The sib who is HLA identical to the proband was clinically completely normal, and while the biochemical results were consistent with the carrier state, they were totally inconsistent with those of an affected homozygote. No other similar recombinations have been reported and it appears that 21-OH is closely linked to the HLA and DR locus. The highest lod scores calculated for the linkage between HLA-B and the 21-OH deficiency gene in 14 informative families is obtained for $\theta = 0.03$. Close linkage has been further confirmed by families in which the parents of an affected child share an HLA haplotype and the affected child or children are HLA homozygous. The linkage between HLA and 21-OH deficiency has been found both in Caucasian and Pakistani families and in salt losing and salt sparing types. The gene for CAH seems, from our data, to be outside the HLA complex and in linkage disequilibrium with HLA BW47 and B8.

The increased frequency of HLA-B47 in CAH has now been described independently by two groups. The significant absence of HLA-B8 among 22 children in our study cannot be explained, but it may be postulated that HLA-B8 is in linkage disequilibrium with another gene which is either lethal in 21-OH homozygotes or completely compensatory. It is noteworthy that a decrease of HLA-B8 is present in most reports published so far (Svejgaard, 1978, personal communication).

The close linkage between HLA and CAH is relevant to both pre- and postnatal diagnosis. For example, we report in family 2 the identification of a previously unsuspected affected male by means of HLA typing. It is now possible to HLA type amniotic cells obtained by amniocentesis in the second trimester. Although amniotic fluid 17-

hydroxyprogesterone assay is quicker and easier, HLA typing will help to confirm or exclude the diagnosis of 21-OH deficiency in a fetus in a family in which CAH has already occurred. Opinions will differ as to whether selective termination of a fetus with 21-OH deficiency is justified in view of the wide variability of clinical manifestation and the relative effectiveness of treatment of affected subjects. However, HLA typing in the second trimester may permit some degree of intraterine treatment of CAH, although by this stage of gestation genital differentiation is far advanced. HLA typing of amniotic cells in the third trimester, or of cord blood lymphocytes, can supplement biochemical diagnosis allowing early diagnosis and treatment of homozygote 21-OH deficients in families in which one case has already occurred. With further experience, HLA typing may be shown to be a reliable way of distinguishing missed cases, carriers, and normal subjects, although at present these distinctions are subject to a small error attributable to genetic recombination.

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References

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